

**Approach for Developing
Water Quality Goals
for United Keno Hill Mines**

Report Prepared for:

**Elsa Reclamation and
Development Company Ltd.
Whitehorse, YT**

Report Prepared by:

**Minnow Environmental Inc.
2 Lamb Street
Georgetown, Ontario
L7G 3M9**

June 2011

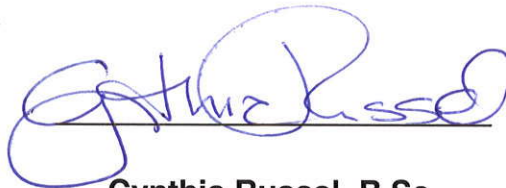
Approach for Developing Water Quality Goals For United Keno Hill Mines

Report Prepared for:

Elsa Reclamation and Development Company Ltd.

Report Prepared by:

Minnow Environmental Inc.



**Cynthia Russel, B.Sc.
Project Manager**



**Patti Orr, M.Sc.
Project Principal**

June 2011

EXECUTIVE SUMMARY

United Keno Hill Mines Limited and UKH Minerals Ltd. were the previous owners of the properties located on and around Galena Hill, Keno Hill and Sourdough Hill, collectively known as the Keno Hill Mining Property. Although the mine has not operated since 1989, abandoned adits (more than 44), buildings/structures, and waste dumps associated with the site represent sources of contaminants to the downstream watersheds. The most significant of these sources include the lime-treated discharge from the tailings pond system, Galkeno 900 Adit, Galkeno 300 Adit, Bellekeno 600 Adit, Silver King Adit and Valley Tailings. The influence from these sources is largely limited to the tributaries that drain the properties (Christal, Flat and Lightning creeks), although some influence on water and sediment quality can be measured further downstream in the South McQuesten River (Minnow 2008, 2009).

In June 2005, Alexco Resources Corp. was selected as the preferred purchaser of the UKHM assets. As required in the purchase agreement, Alexco formed a subsidiary company – Elsa Reclamation and Development Company Ltd. (ERDC), to own and manage the site. Another requirement of the purchase agreement is for ERDC to prepare and implement, to the satisfaction of the Governments, a Reclamation Plan to address historical mining liabilities on the UKHM claims. Funding for the development and implementation of the Closure Plan is primarily from the Government of Canada (represented by Indian and Northern Affairs Canada (INAC) with some cost sharing by ERDC.

Under the purchase agreement, ERDC is allowed to resume production at a historic mine by declaring it as a Production Unit. The terrestrial liability associated with historical mine operations within the Production Unit remains with the Government of Canada, however, ERDC becomes responsible for water related liabilities in addition to any new terrestrial liabilities associated with the redevelopment of mine operations within the Production Unit. Alexco Resources operates the Bellekeno Mine and is responsible for the BK625 treatment facility and new terrestrial liabilities associated with Bellekeno operations.

One of the mandates of ERDC is to develop a Reclamation Plan for the “Existing State of the Mine” such that historical mining liabilities may be addressed and future environmental conditions anticipated. ERDC is currently in the process of developing this reclamation plan.

As part of the development and implementation of a closure plan, ERDC, INAC, Yukon Government (YG), First Nations and other interested groups will need to establish criteria on which to evaluate closure conditions and set expectations for environmental performance within the downstream receiving environment. Based on a review of water quality data within the watersheds affected by historical UKHM operations, it is unlikely that concentrations of

key mine related contaminants (cadmium and zinc) will achieve any site specific water quality objective (SSWQO) developed through standard approaches (CCME 2003). Therefore, an alternative approach for setting water quality expectations should be considered for these historically affected areas.

EDRC retained Minnow Environmental Inc. to develop an approach to water quality goals and assessment in Christal and Flat creeks and the South McQuesten River. The approach provided herein is a conceptual level approach to water quality evaluation that needs to be considered by the various stakeholders associated with UKHM prior to proceeding with final objectives and goals for the various affected watercourses. This Framework will be reviewed by stakeholders (INAC, First Nations and YG) to arrive at final water quality objectives for the historic properties prior to the selection of closure options. Ultimately, specific details of an approved approach to water quality evaluation will need to be incorporated into future water license requirements.

Recognizing that the UKHM receiving environment has been effected over many years and decades by various mining operations, the objective of this study was to develop an approach to assessing water quality downstream of UKHM that would serve to protect existing biota within the immediate receiving environment (Flat and Christal creeks) and provide for no further degradation of the South McQuesten River relative to upstream conditions. In order to achieve this objective, water quality data was reviewed relative to the toxicological thresholds in the proposed Canadian Water Quality Guidelines (CWQG; Environment Canada 2009a, b), and upstream or regional background conditions.

South McQuesten River

In mid 2007, it was noted that concentrations of various metals including cadmium and zinc had increased upstream of UKHM on the South McQuesten River (KV-1). Further examination found that concentrations of cadmium and zinc have been above both the existing and proposed future CWQG in the South McQuesten River since mid-2006. Furthermore, cadmium and zinc concentrations showed an increasing trend both upstream of UKHM at KV-1 and downstream at KV-4 and KV-5 between 2006 and 2009. Comparison of the slopes of these increases determined that the concentrations of cadmium and zinc are increasing at the same rate downstream (KV-4 and KV-5) as upstream (KV-1) indicating that the source of the increase in water concentrations in the South McQuesten River is upstream of UKHM.

When background concentrations exceed a CWQG, a statistic describing the upper range of background concentrations (e.g. 95th percentile) can be used as a SSQWO (CCME 2003).

However, it is uncertain whether or for how long concentrations upstream of UKHM (KV-1) will continue to increase, so it is not currently possible to establish a single numerical value that represents the upper limit of upstream/background conditions. This means that an alternative approach must be considered for evaluating the influence of UKHM in downstream areas of the South McQuesten (KV-4, KV-5). Therefore, it is recommended that downstream water quality objectives for cadmium and zinc and possibly other contaminants be linked to concentrations upstream of UKHM (i.e., KV-1) such that no further degradation of water quality occurs downstream of UKHM (KV-4 and KV-5) relative to upstream conditions.

Flat Creek and Christal Creek

Over the past twenty years concentrations of cadmium and zinc have been elevated well above the CWQG (i.e., mean values more than ten times the CWQG) in Flat Creek and Christal Creek (Minnow 2008). Since 2006, concentrations of both cadmium and zinc have been decreasing in both Flat Creek (KV-9) and Christal Creek (KV-7), and concentrations are now in the same range as those observed in the South McQuesten River. The improvement in water quality within the tributaries is likely associated with remedial measures implemented by EDRC starting in 2006 (e.g. clarifies at Galkeno 300 and water management and treatment at the Galkeno 900). While it is expected that concentrations will continue to decrease, they are not expected to achieve CWQG in the near future due to continued contributions from non-point sources. Therefore, alternative water quality goals need to be developed for Flat and Christal creeks.

Mean cadmium and zinc concentrations (2009) in both Flat and Christal Creeks have been greater than proposed long-term term exposure guidelines and also, in the case of zinc, maximum concentrations are above the proposed short-term exposure guideline, suggesting potential for effects to biota within these watercourses.

It is recommended that water quality goals for Flat and Christal creeks should aim to prevent further degradation of water quality such that the diversity and abundance of the existing resident biota is protected. Routine monitoring conducted as part of the Long-Term Aquatic Monitoring Program (Minnow 2011) should be assessed annually to ensure conditions are stable or improved relative to previous conditions and background and the effects of these goals should be verified through routine biological monitoring. It is expect that as conditions improve and stabilize the magnitude and frequency will be reduced.

TABLE OF CONTENTS

| | | |
|------------|-----------------------------------------------------------------------------------|-----------|
| 1.0 | INTRODUCTION | 1 |
| 1.1 | Background | 1 |
| 1.2 | Project Objectives and Approach | 3 |
| 1.3 | Document Organization | 3 |
| 2.0 | METHODS | 5 |
| 2.1 | Evaluation of Recent Water Quality | 5 |
| 2.2 | Assessment of Potential Impacts | 5 |
| 2.2.1 | Toxicity Data Sources and Organization | 5 |
| 2.2.2 | Assessing Relative Species Sensitivities | 6 |
| 2.3 | Assessment of Water Quality Impacts | 7 |
| 3.0 | GOALS AND OBJECTIVES | 8 |
| 3.1 | South McQuesten River | 8 |
| 3.1.1 | Current Conditions | 8 |
| 3.1.2 | Proposed Approach for Water Quality Assessment in the South McQuesten River | 9 |
| 3.2 | Tributaries | 9 |
| 3.2.1 | Current Water Quality | 9 |
| 3.2.2 | Review of Toxicity Information | 10 |
| 3.2.3 | Proposed Approach to Water Quality Goals | 12 |
| 4.0 | CONCLUSIONS | 14 |
| 5.0 | REFERENCES | 15 |

APPENDIX A: Water Quality Data

APPENDIX B: Cadmium Toxicity Data

APPENDIX C: Zinc Toxicity Data

APPENDIX D: Hardness- Toxicity Relationships for Zn and Cd

LIST OF TABLES

After Page:

| | | |
|------------|--------------------------------------------------------------------------------------------------------|----|
| Table 3.1: | Lowest effect endpoint reported for each freshwater species after a long-term exposure to cadmium..... | 11 |
| Table 3.2: | Lowest effect endpoint reported for each freshwater species after a long-term exposure to zinc..... | 11 |

LIST OF FIGURES

After Page:

| | | |
|-------------|--------------------------------------------------------------------------------------------------------|----|
| Figure 1.1: | Location of United Keno Hill Mine complex..... | 1 |
| Figure 1.2: | Mine workings and surrounding watercourses..... | 1 |
| Figure 2.1: | Water monitoring stations used in assessment..... | 5 |
| Figure 3.1 | Water concentration of total cadmium and zinc measured in the South McQuesten River 2006 to 2009 | 8 |
| Figure 3.2: | Comparison of water concentrations over time in the South McQuesten River. | 8 |
| Figure 3.3: | Concentrations of total cadmium and zinc measured at Flat and Christal Creek, 2006 to 2009 | 10 |
| Figure 3.4: | Mean concentrations of total cadmium, total zinc and dissolved hardness, 2009 | 10 |
| Figure 3.5: | Total cadmium and zinc concentrations at UKHM stations relative to CWQG and toxicity thresholds. | 10 |
| Figure 3.6: | Number of Heptageniidae identified in samples collected in the vicinity of UKHM in August 2009 | 13 |

1.0 INTRODUCTION

1.1 Background

United Keno Hill Mines Limited and UKH Minerals Ltd. were the previous owners of the properties located on and around Galena Hill, Keno Hill and Sourdough Hill, collectively known as the Keno Hill Mining Property. For the purposes of this report these mining areas are referred to as the United Keno Hill Mines (UKHM). The UKHM complex is located in north-central Yukon Territory (Figure 1.1) and is comprised of approximately 827 mineral claims that cover an area of approximately 15,000 ha (about 29 km long and 8 km wide). Although the mine has not operated since 1989, there are abandoned adits (more than 44), buildings/structures, and waste dumps associated with the UKHM Complex that represent sources of contaminants to the downstream watersheds. The most significant of these sources include the lime-treated discharge from the tailings pond system, Galkeno 900 Adit, Galkeno 300 Adit, Bellekeno 600 Adit, Silver King Adit and Valley Tailings (Figure 1.2; Burns 2008). The influence from these sources is largely limited to the tributaries that drain the properties (Christal, Flat and Lightning creeks), although some influence on water and sediment quality can be measured further downstream in the South McQuesten River (Minnow 2008, 2009). In addition to the historical mining activities, the area is currently host to a number of placer mining operations which cause extensive alteration of the watercourses and impacts to habitat and water quality downstream (Dan Cornett, Access Consulting, pers. comm.; Pentz and Kostaschuk, 1999).

In June 2005, Alexco Resources Corp. was selected as the preferred purchaser of the UKHM assets. As required in the purchase agreement, Alexco formed a subsidiary company – Elsa Reclamation and Development Company Ltd. (ERDC), to own and manage the site. Another requirement of the purchase agreement was for ERDC to prepare and implement, to the satisfaction of the Governments, a Closure Plan to address historical mining liabilities on the UHKM claims. Funding for the development and implementation of the Closure Plan is primarily from the Government of Canada (represented by Indian and Northern Affairs Canada (INAC) with some shared costs by ERDC.

Under the purchase agreement, ERDC is allowed to resume production at a historic mine by declaring it as a Production Unit. The terrestrial liability associated with historical mine operations within the Production Unit remains with the Government of Canada, however, ERDC becomes responsible for water related liabilities in addition to any new terrestrial



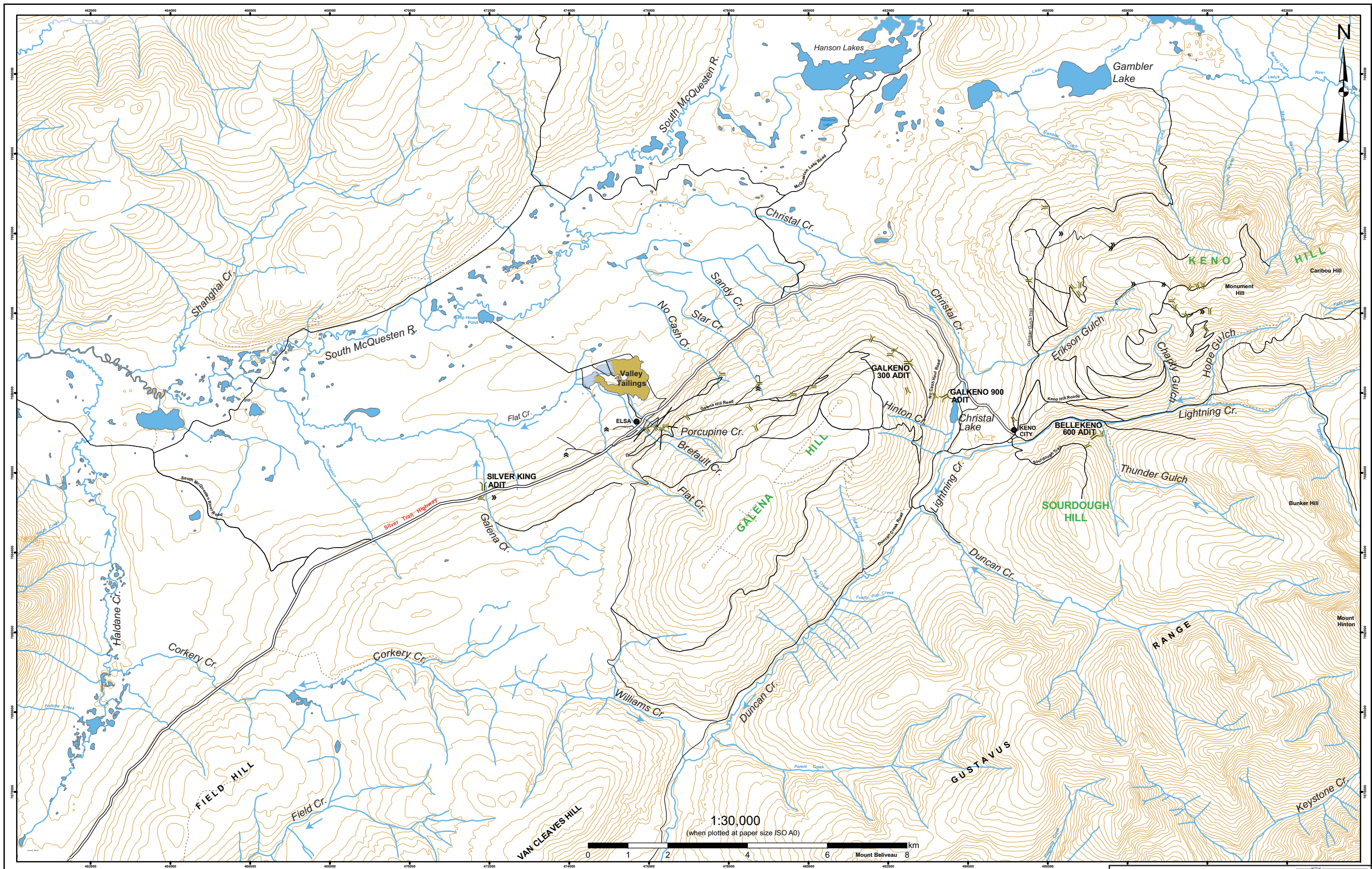
Figure 1.1

minnow
ENVIRONMENTAL INC.

Location of United Keno Hill Mines within the Yukon Territory.

Ref: 2272
Date: April 2011

Source: Access Consulting Group



Legend

Mine Working

- Adit
- Shaft (to surface - connection to underground not determined)
- Valley Tailings
- Pit

Topography

- Town
- Silver Trail
- Secondary and Limited-use roads

- Trail
- Watercourse
- Waterbody
- Flow Direction

Figure 1.2

United Keno Hill Mines, Mine Workings and Surrounding Watercourses

Ref: 2272
Date: April 2011

Source: Access Consulting Group

liabilities associated with the redevelopment of mine operations within the Production Unit. Alexco Resources operates the Bellekeno Mine and is responsible for the BK625 treatment facility and new terrestrial liabilities associated with Bellekeno operations. Regardless of the current and potential future production units, INAC has, and will continue to have, significant involvement in the development of the Closure Plan for the UKHM property.

As part of the development and implementation of a closure plan, EDRC, INAC, YG, First Nations and other interested groups will need to establish criteria on which to evaluate closure conditions and set expectations for environmental performance within the downstream receiving environment. A review of water quality at UKHM identified cadmium and zinc as the primary contaminants of concern (Minnow 2008). The concentrations of these substances were substantially elevated in the tributaries of the South McQuesten River (Flat Creek and Christal Creek) but were generally within Canadian Water Quality Guidelines (CWQG; CCME, 1999) within the South McQuesten River downstream of UKHM. Therefore it was expected that the CWQG could be used as assessment values for the South McQuesten River. However, water quality within the tributaries (Flat and Christal creeks) is not expected to achieve the CWQG in the near future. Furthermore, it is unlikely that concentrations of these elements (cadmium and zinc) would achieve a site specific water quality objective (SSWQO) developed through standard approaches (CCME 2003). Therefore, an alternative approach for setting water quality expectations should be considered for these historically affected areas.

EDRC retained Minnow Environmental Inc. to develop an approach to water quality goals and assessment in Christal and Flat creeks and the South McQuesten River¹. This report summarizes our assessment of water quality conditions in both the tributaries and South McQuesten River and provides a recommended approach for future water quality assessment downstream of UKHM. The approach provided herein is a conceptual level approach to water quality evaluation that needs to be considered by the various stakeholders associated with UKHM prior to proceeding with final objectives and goals for the various affected watercourses. This Framework will be reviewed by stakeholders (INAC, First Nations and YG) to arrive at final water quality objectives for the historic properties prior to the selection of closure options. Ultimately, specific details of an

¹ Lightning Creek was not considered in this assessment as the existing data base indicated that concentrations within Lightning Creek were close to the CWQG. However, the data base for the creek was limited with respect to the extent of data and appropriate method detection limits. As additional monitoring data is obtained, the status of Lightning Creek with respect to the need for water quality objectives should be reassessed.

approved approach to water quality evaluation will need to be incorporated into future water license requirements.

1.2 Project Objectives and Approach

As part of a previous water quality assessment (Minnow 2008), cadmium and zinc were identified as the primary contaminants of concern (COCs) related to the UKHM complex because they are the substances that are most elevated relative to guidelines and water quality in undisturbed areas. While a number of other substances were identified as possible COCs, these have yet to be confirmed². However, management of cadmium and zinc is expected to control other mine related substances. Thus, an approach to water quality goals is only recommended for cadmium and zinc at this time. Once sufficient information is compiled on the possible COCs, the need for water quality goals or objectives for these substances should be revisited.

Recognizing that the UKHM receiving environment has been effected over many years and decades by various mining operations, the objective of this study was to develop an approach to assessing water quality downstream of the historic UKHM complex that would:

- serve to protect the resident biota (no decline relative to current species diversity) and prevent further degradation of water quality with in the immediate receiving environment (Flat and Christal creeks) and
- provide for no further degradation in the South McQuesten River relative to upstream conditions.

In order to achieve this objective, recent water quality was reviewed relative to the toxicological thresholds in the proposed CWQG (Environment Canada 2009a, b), and upstream or regional background conditions.

1.3 Document Organization

Section 2.0 presents the methods used in the evaluation of data for this project. Section 3.0 summarizes the current water quality in the South McQuesten River and its tributaries, the toxicity data for cadmium and zinc presented in the scientific literature, and compares these values to recent water quality concentrations for aquatic environments downstream

² EDRC has initiated a more robust water quality monitoring program in order to compile the data necessary to assess possible COCs. When two years of monitoring data at appropriate method detection limits has been compiled, the final list of mine COCs and monitoring parameters can be established.

of UKHM. Conclusions are presented in Section 4.0. References cited throughout the document are presented in Section 5.0

2.0 METHODS

2.1 Evaluation of Recent Water Quality

Aqueous cadmium and zinc concentrations were assessed for routine monitoring stations in Christal Creek (KV-7) and Flat Creek (KV-9), as well as in the South McQuesten River both upstream (KV-1) and downstream of UKHM inputs (KV-4 and KV-5) (Figure 2.1; Appendix A). The mine exposed stations selected delimit mine influence within each water course assessed. Concentrations of these substances were plotted over time (2006 to 2009) and relative to water quality guidelines. The current guidelines (CCME 1999) were initially used to evaluate past water quality data. However, new guidelines are in development for cadmium and zinc that incorporate the large number of studies published in the scientific literature since the original guidelines were developed and also involve a different method for guideline derivation (Environment Canada 2009 a, b). The proposed guidelines are therefore a better reflection of the state-of-the-science regarding aquatic effects of cadmium and zinc; however, they are still undergoing provincial agency review and then will need to be circulated for public comment prior to being adopted. Although this process may result in modification of the guideline values, any adjustments are expected to be minor, so the proposed guidelines were considered in the water quality evaluation for UKHM.

The existing CWQG for cadmium (CCME 1999) and proposed revised guidelines for both cadmium and zinc (Environment Canada 2009 a, b) depend on water hardness, because the toxicity of these substances declines as hardness increases. A hardness of 100 mg/L as CaCO_3 was selected as the basis for comparison in this evaluation, because water hardness downstream of the UKHM is rarely lower than this level (Appendix A) and the mean background concentration is also higher (i.e., 162 mg/L; Minnow 2008). Thus the water quality guidelines applied in this assessment were conservative in terms of flagging concentrations that might be of concern with respect to protection of aquatic biota in waters downstream of UKHM.

For stations where water quality trends were observed over time, slopes were plotted and statistically compared using analysis of covariance (ANCOVA) (SPSS 2003).

2.2 Assessment of Potential Impacts

2.2.1 Toxicity Data Sources and Organization

Recently-prepared toxicity data summaries for zinc and cadmium, along with copies of much of the source literature were obtained from Environment Canada's Guidelines and

Standards Office in Gatineau Quebec, augmented by relevant scientific literature from Minnow's in-house files. The source literature was reviewed to verify the accuracy of the data sets provided and allow for clarification of test conditions. The reviews of source literature resulted in some minor modifications of the original data sets.

Raw data tables presented details of each toxicity test, including results of replicate exposures within studies, if available. The tabulated information included species common and scientific name, test duration, test endpoint, observed effect concentration, test conditions (e.g., hardness, pH, etc., if reported) and source (author, year). Toxicity test data were distinguished as short-term versus long-term exposures. Depending on the species and life-stage tested, short-term exposures were defined as less than one (algae), four (invertebrates and larval fish), or seven days (older fish), consistent with current Environment Canada protocols (Environment Canada 2008, 2009). Long-term exposure data were considered most relevant for assessing potential effects associated with chronic exposure to mine-related contaminants at UKHM so toxicity evaluations for cadmium and zinc focussed primarily on long-term exposure data.

2.2.2 Assessing Relative Species Sensitivities

The toxicities of cadmium and zinc are influenced by various water quality factors, of which the best characterized is hardness (Environment Canada 2009 a, b). In order to rank the relative sensitivities of different aquatic species to these contaminants, it was necessary to convert reported effect concentrations to common water hardness, thereby removing the influence of variable test conditions on relative species toxicity thresholds. A conservative hardness of 100 mg L⁻¹ as CaCO₃ was selected as the basis for comparison, because water hardness downstream of the UKHM is rarely lower than this level. However, effects at other hardnesses were also evaluated (see below).

Hardness-toxicity relationships were first defined for individual species for which adequate data were available. This was done by examining the raw data to identify toxicity tests for which the hardness of the exposure water was reported and identifying species for which there were data for the same or similar endpoints over a range of exposure water hardness levels (>2-fold range), including at least one exposure with water hardness of at least 100 mg/L. For zinc, only tests conducted at pH ≥ 7.4 were included to minimize the variability associated with this potentially confounding factor (more data were available for a basic than neutral-acidic range of exposure pH). If there were replicate test results within a study for the same endpoint at the same or similar water hardness, geometric mean hardness and effect concentrations were computed before using the data to define the hardness-toxicity relationship for the substance.

The resulting data were then plotted for each species based on a regression of natural logarithm (Ln) of toxicant concentration as the dependent variable against Ln hardness as the independent variable. This yielded the species-specific slope for the hardness-toxicity relationship. For both substances (Cd, Zn), there were sufficient data to derive a slope for at least one plant/alga, invertebrate, and fish species.

For each contaminant, the slopes for the hardness-toxicity relationship were tested for differences among species and, if not different, the slopes were combined to generate an average slope. After consideration of the data and potential implications of applying different slopes for different species or groups of species, a decision was made to generate a combined slope (including short- and long-term exposure data). This recognized that the range of slopes for short- versus long-term exposure data overlapped, there were no differences in slope that could be confidently ascribed to particular species or group, and this approach would ensure better consistency with respect to data handling for both contaminants.

To assess relative species sensitivity, the lowest toxic effect concentration corresponding to an exposure hardness of 100 mg/L was identified for each study and species. For studies not including an exposure hardness of 100 mg/L, the toxic effect concentration for each study-species combination was adjusted to an exposure hardness of 100 mg/L based on the combined slope for the hardness-toxicity relationship. The lowest value for each species (at hardness of 100 mg/L) was then identified and used to rank all species in terms of relative sensitivity. In cases where there were both effect (e.g., EC10, IC25, maximum acceptable toxicant concentration [MATC], etc.) and no-effect (NOEC) endpoints reported, the lowest effect concentration was selected.

For species for which the original test hardness was reported, the same slope factors described above were also used to estimate toxicity at other water hardness values (than 100 mg/L). If test hardness was not reported, an adjustment could not be made and the same reported effect concentration for the species was assumed for all water hardnesses.

2.3 Assessment of Water Quality Impacts

Recent water concentrations of cadmium and zinc measured at key locations downstream from UKHM were compared to toxic threshold concentrations reported in the literature for different aquatic species (at a water hardness of 100 mg/L). This was done to identify species that may be adversely affected by current water quality and take this into account in setting future water quality goals.

3.0 GOALS AND OBJECTIVES

3.1 South McQuesten River

3.1.1 Current Conditions

Prior to 2006, water concentrations of cadmium and zinc in the South McQuesten River downstream of UKHM were generally less than the existing CWQG (Minnow 2008). Therefore, it was expected that the CWQG could be used as benchmarks for assessment of future water quality and that site-specific water quality goals or objectives would not be required. However, in mid 2007, it was noted that concentrations of various metals including cadmium and zinc had increased upstream on the South McQuesten River at KV-1. Further examination of cadmium and zinc concentrations indicated that the concentrations have been above both the existing and proposed future CWQG both upstream and downstream of UKHM since mid-2006 (Figure 3.1). In addition, concentrations showed an increasing trend both upstream of UKHM at KV-1 and downstream at KV-4 and KV-5 between 2006 and 2009. Comparison of the slopes of these increases determined that the concentrations of cadmium and zinc are increasing at the same rate downstream (KV-4 and KV-5) as upstream (KV-1; Figure 3.2). Furthermore, mean zinc concentrations adjusted for date were significantly lower at stations downstream of the UKHM discharges (KV-4 and KV-5; $p < 0.001$) than at upstream station KV-1 over the same period. Similarly, cadmium concentrations at KV-5 were significantly lower than at KV-1 ($p = 0.001$), while mean cadmium concentrations at KV-4 and KV-1 were similar ($p = 0.208$; Appendix Table A.2c). These results indicate that a source upstream of KV-1 was responsible for the increase in water concentrations in the South McQuesten River (KV-1, KV-4 and KV-5) and that UKHM is not causing a measurable increase in concentrations downstream of the site at KV-4 and KV-5.

When background concentrations exceed a CWQG, a statistic describing the upper range of background concentrations (e.g. 95th percentile) can be used as a site specific water quality objective (SSQWO; CCME 2003). However, it is uncertain whether or for how long concentrations upstream of UKHM (KV-1) will continue to increase, so it is not currently possible to establish a single numerical value that represents the upper limit of upstream background conditions. This means that an alternative approach must be considered for evaluating the influence of UKHM in downstream areas of the South McQuesten (KV-4, KV-5).

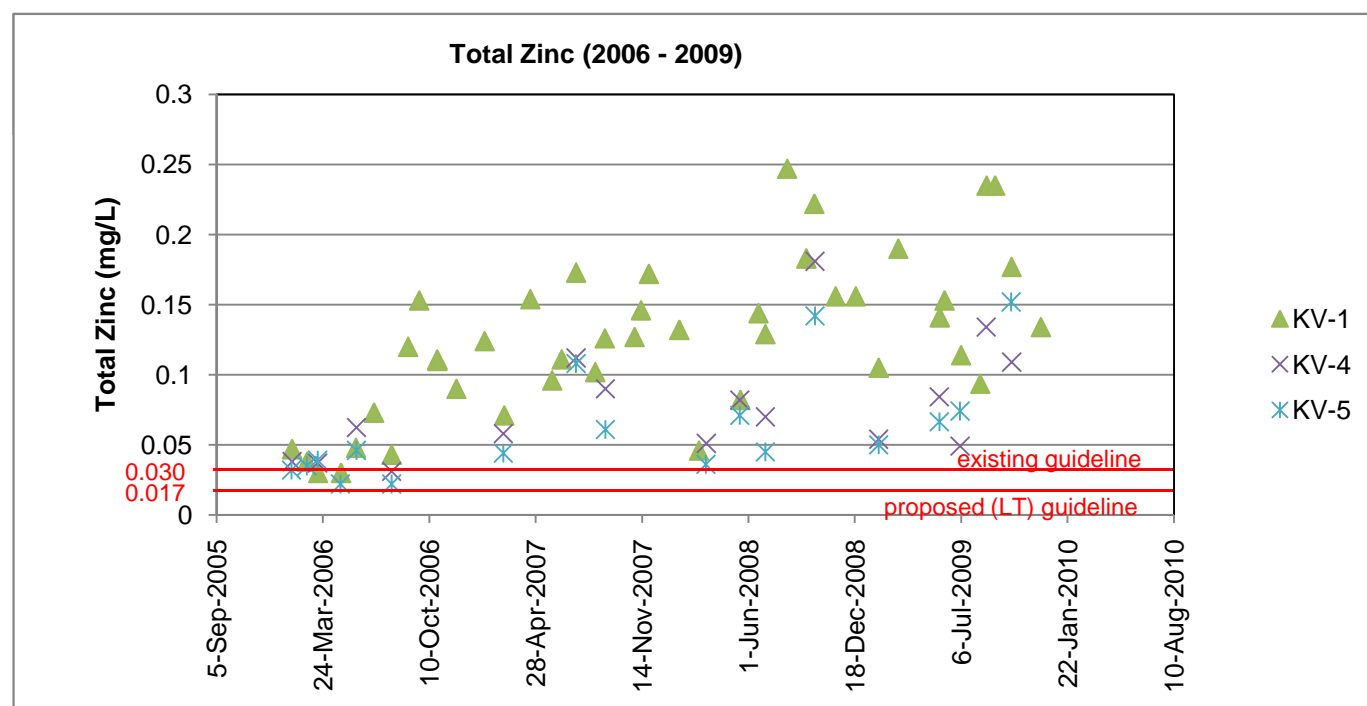
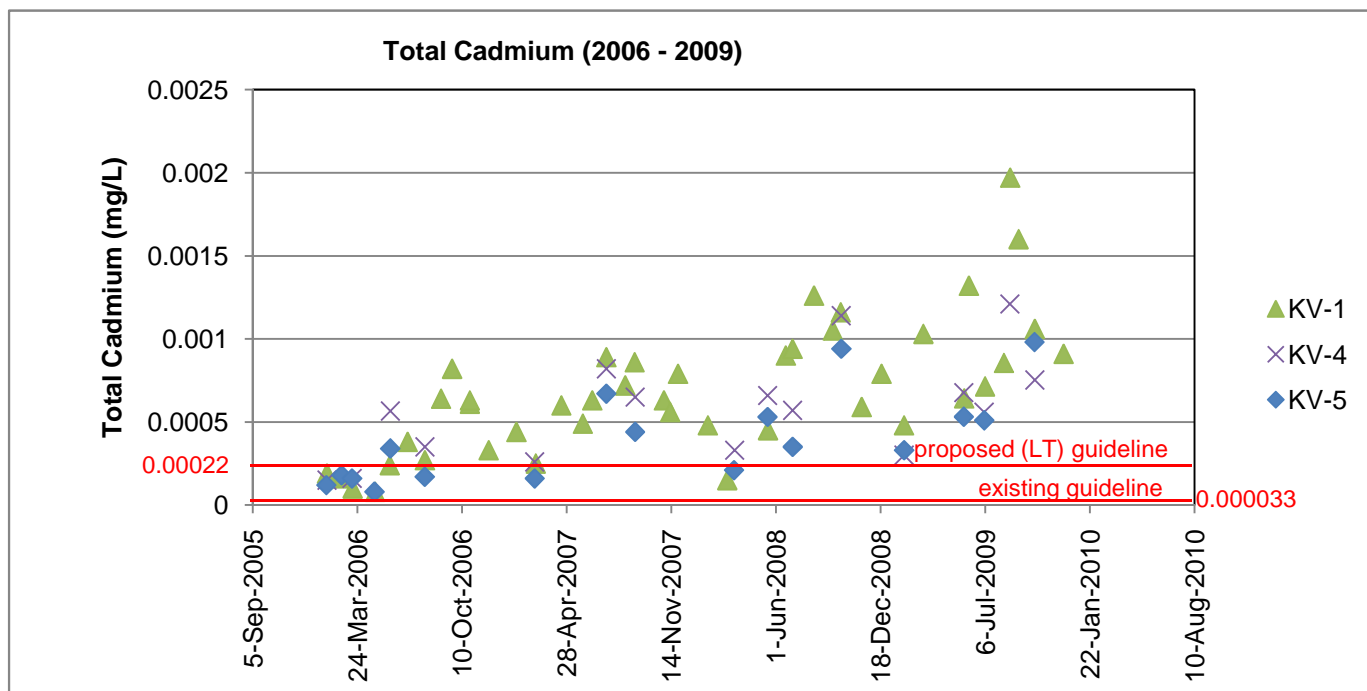


Figure 3.1: Water concentrations of total cadmium and zinc measured in the South McQuesten River at KV-1, KV-4 and KV-5 between 2006 and 2009 relative to existing (CCME 1999) and proposed long-term (LT) exposure guidelines based on a water hardness of 100 mg/L (Environment Canada 2009 a,b).

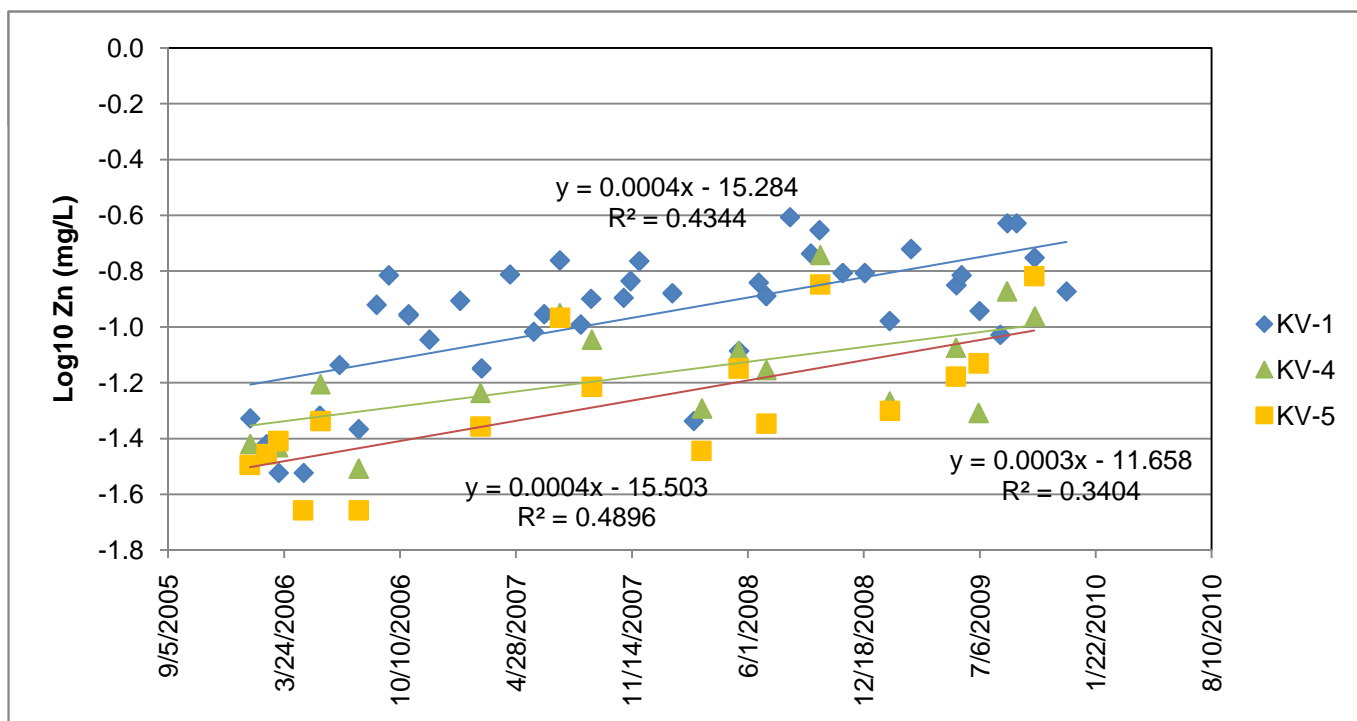
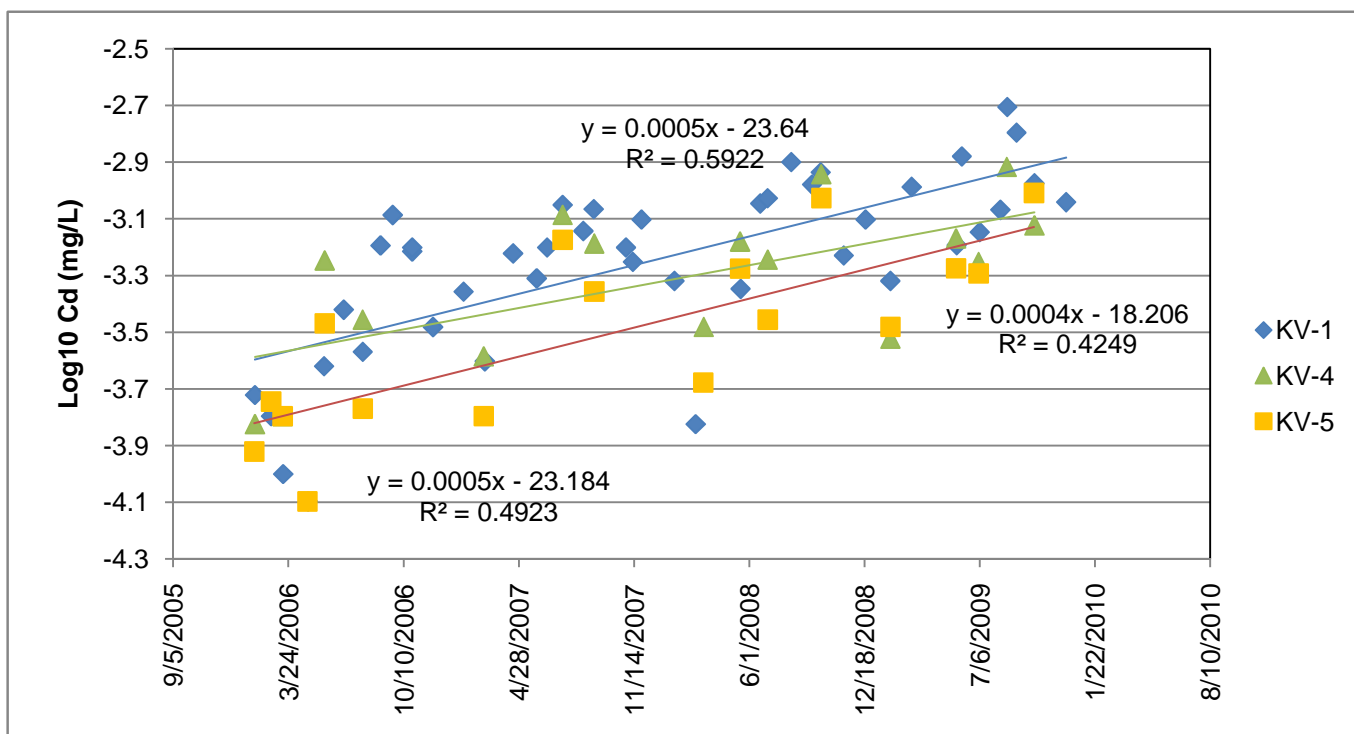


Figure 3.2: Comparison of water concentrations over time (slopes) at South McQuesten River stations KV-1, KV-4 and KV-5, 2006 to 2009. Slopes were not significantly different at $p=0.05$ (Appendix Table A.2).

3.1.2 Proposed Approach for Water Quality Assessment in the South McQuesten River

As noted above, concentrations of cadmium and zinc have been increasing downstream of UKHM as a result of increasing concentrations upstream. Until the concentrations upstream are reduced or stabilize, a single water quality criterion cannot reasonably be employed downstream in the South McQuesten River. Therefore, downstream water quality objectives for cadmium and zinc and possibly other contaminants should be linked to concentrations upstream of UKHM (i.e., KV-1) such that no further degradation of water quality occurs downstream of UKHM (KV-4 and KV-5) relative to upstream conditions. A possible approach to evaluating water quality within the South McQuesten River might include:

- An annual evaluation of monthly water samples collected at KV-1, KV-4 and KV-5, relative to all substances associated with UKHM, particularly cadmium and zinc.
- The mean annual concentrations of cadmium and zinc at KV-4 and KV-5 should not exceed those measured upstream at KV-1. Means should be statistically compared using ANOVA. An increase in mean cadmium and/or zinc that is not statistically significant should still be investigated to determine if it may be an early indicator of an increasing trend associated with historic UKHM inputs (e.g., perhaps examine trends in water quality monitoring data from upstream source areas based on both concentrations and loads).
- Water quality trends at all areas should be evaluated by a qualified professional at a frequency consistent with detailed watershed reporting as recommended in the Long-Term Aquatic Monitoring Program (LTAMP; Minnow 2011).
- The slope of downstream concentrations (KV-4 and KV-5) should be statistically compared to KV-1 (ANCOVA) to confirm that the rates of change downstream are equal to or less than that measured upstream.

3.2 Tributaries

3.2.1 Current Water Quality

Lightning Creek was not considered in this assessment as the existing data base indicated that concentrations within Lightning Creek were close to the CWQG (CCME 1999). However, the data base for the creek was limited with respect to the extent of data and appropriate method detection limits. The status of Lightning Creek with respect to the need for water quality objectives should be reassessed when additional monitoring data is

obtained and in light of proposed changes to the CCME cadmium and zinc guidelines (Environment Canada 2009a,b). In addition, there is extensive placer mining within the Lightning Creek and Duncan Creek watersheds and this will need to be taken into consideration as a potential contributor when an approach to water quality assessment is developed for Lightning Creek.

In Flat Creek and Christal Creek, concentrations of cadmium and zinc have historically been elevated well above the CWQG (i.e., mean values more than ten times the CWQG; Minnow 2008). Although closure and associated remediation measures are planned and/or underway, it is not possible to accurately predict future concentrations due to the number of sources and uncertainty of future loads from surface contamination within these catchments (i.e., remedial measures are aimed at addressing specific known sources but it is unclear to what extent total loadings can be expected to decrease).

Since 2006, concentrations of both cadmium and zinc have been decreasing in both Flat Creek (KV-9) and Christal Creek (KV-7; Figure 3.3), and concentrations are now in the same range of those observed in the South McQuesten River (Figure 3.4). The improvement in water quality within the tributaries is likely associated with remedial measures implemented by EDRC starting in 2006 (e.g. clarifies at Galkeno 300 and water management and treatment at the Galkeno 900). While it is expected that concentrations will continue to decrease, they are not expected to achieve CWQG in the near future due to continued contributions from non-point sources (e.g. disbursed tailings dust on surface soils). Therefore, alternative water quality goals need to be developed for these creeks. Mean cadmium and zinc concentrations (2009) in both Flat and Christal Creeks have been greater than proposed long-term term exposure guidelines and in the case of zinc, maximum concentrations are also above the proposed short-term exposure guideline, suggesting potential for effects to biota within these watercourses (Environment Canada 2009 a, b; Figure 3.5). Potential site-specific impacts are evaluated in more detail below.

3.2.2 Review of Toxicity Information

While concentrations are expected to remain above the long-term CWQG for the foreseeable future, the implications to resident and locally important biota is species-specific because effect concentrations vary widely between aquatic species and are dependent upon site specific water quality factors. Also, the CWQG presented are based on a conservative water hardness of 100 mg/L, which is lower than the hardnesses observed in Flat Creek and Christal Creek during the past three years (i.e., 80% of values > 200 mg/L and mean values \geq 300 mg/L) (Figure 3.4; Appendix Table A.1).

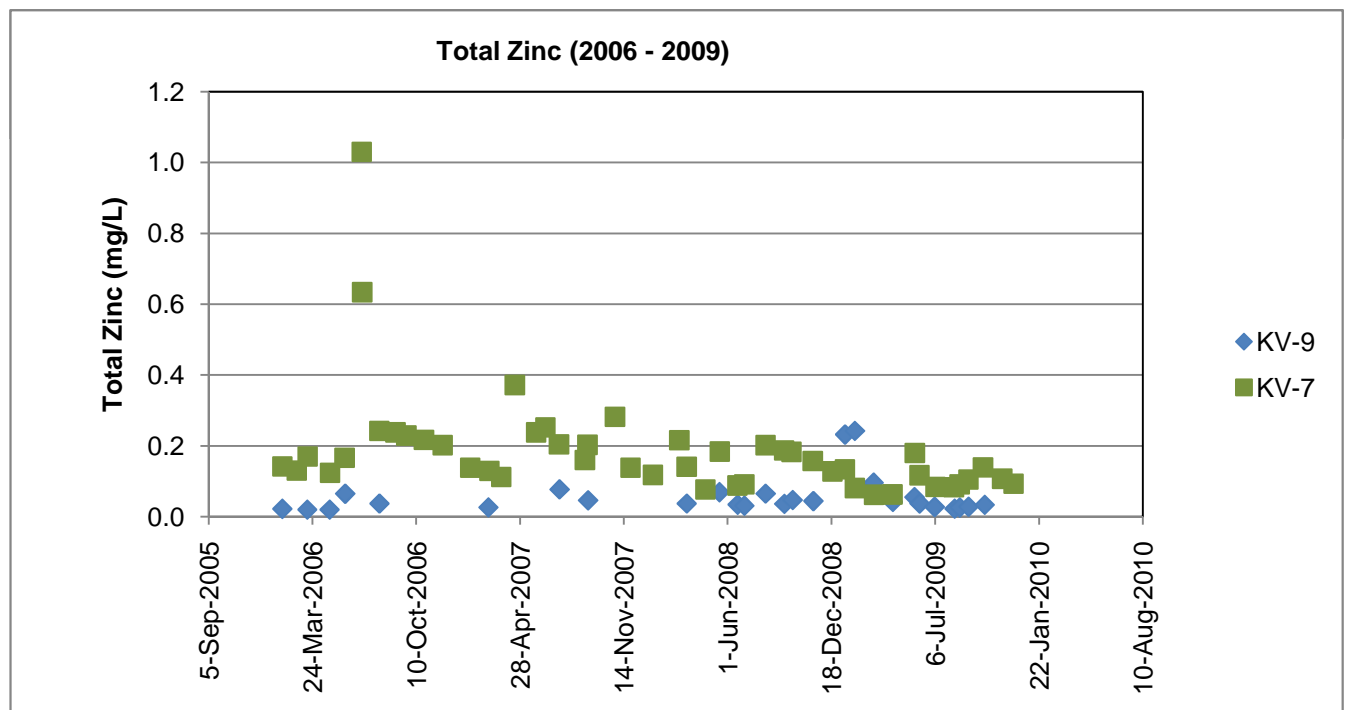
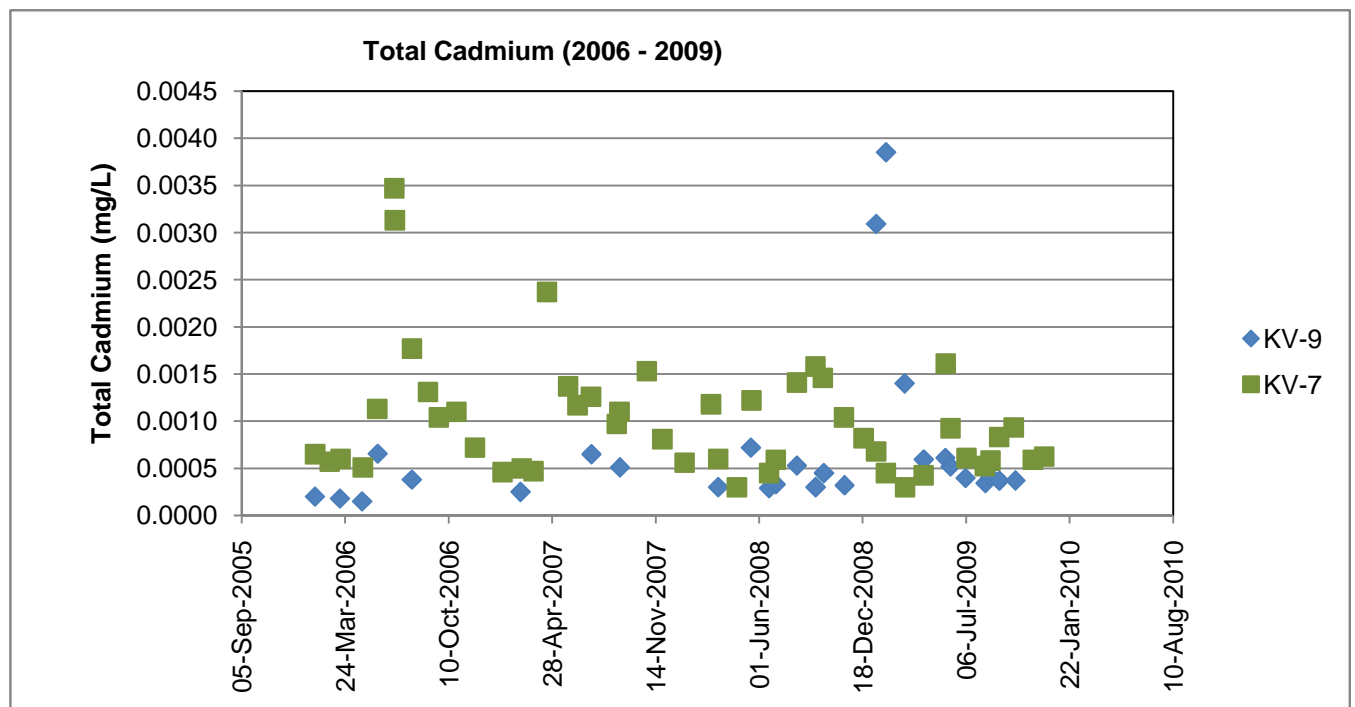


Figure 3.3: Concentrations of total cadmium and zinc measured at Flat Creek (KV-9) and Christal Creek (KV-7) between 2006 and 2009.

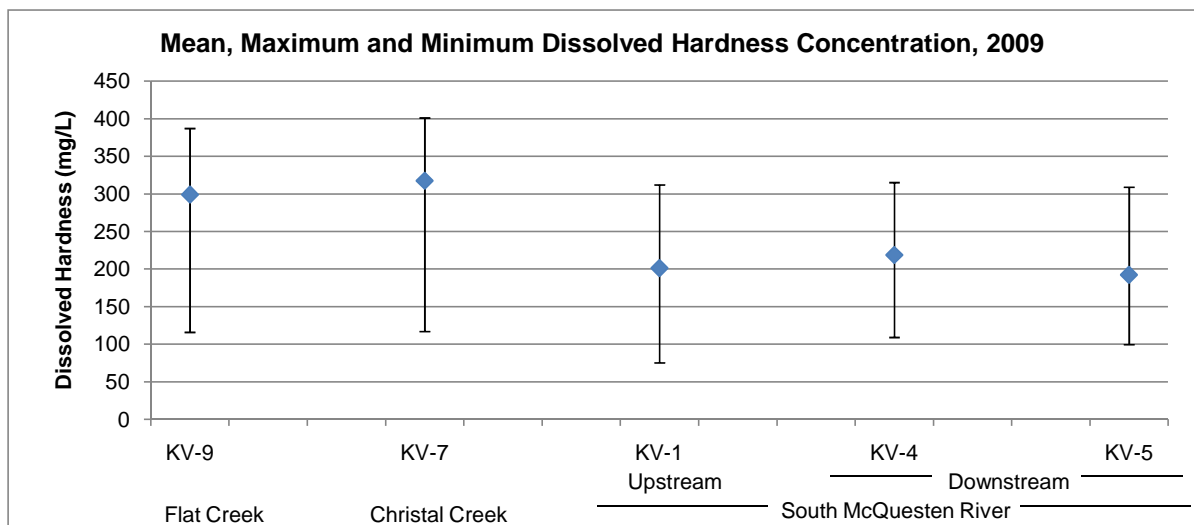
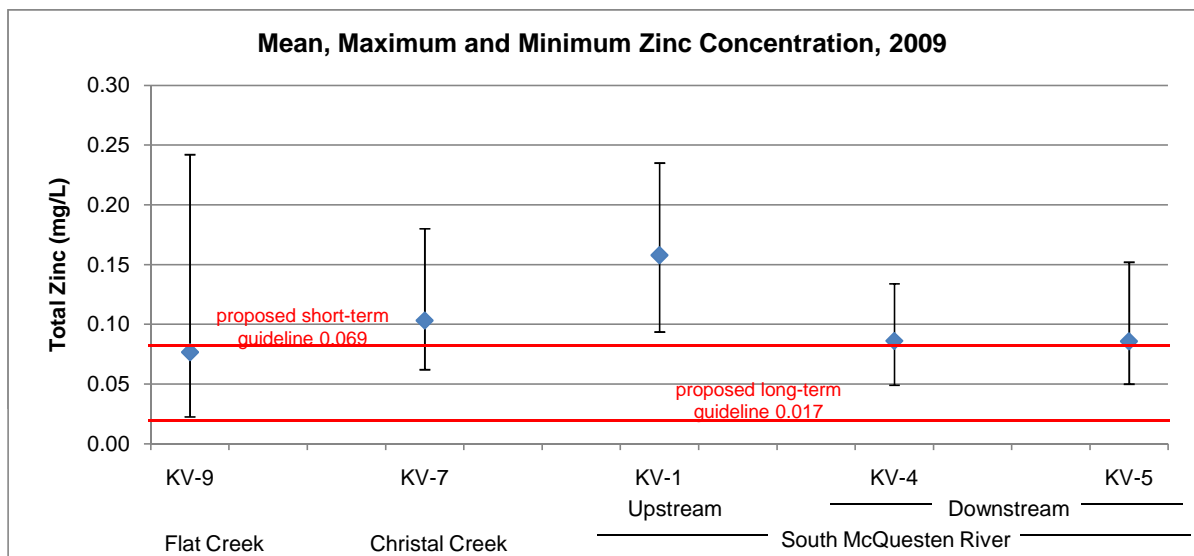
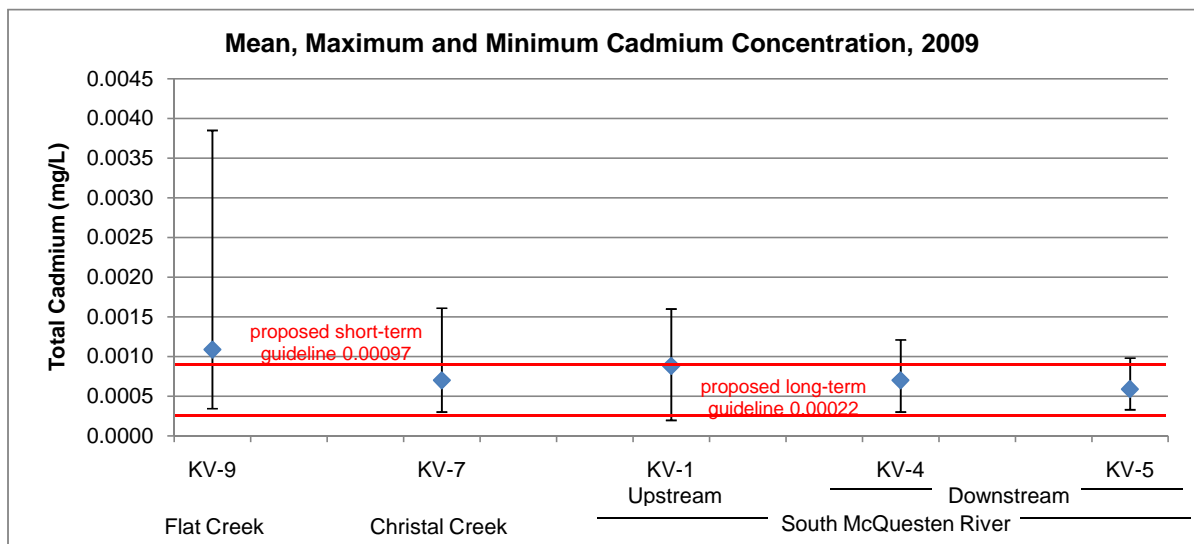


Figure 3.4: Mean water concentration (with maximum and minimum) of total cadmium, total zinc and dissolved hardness at key stations near UKHM in 2009. Guidelines are based on water hardness of 100 mg/L.

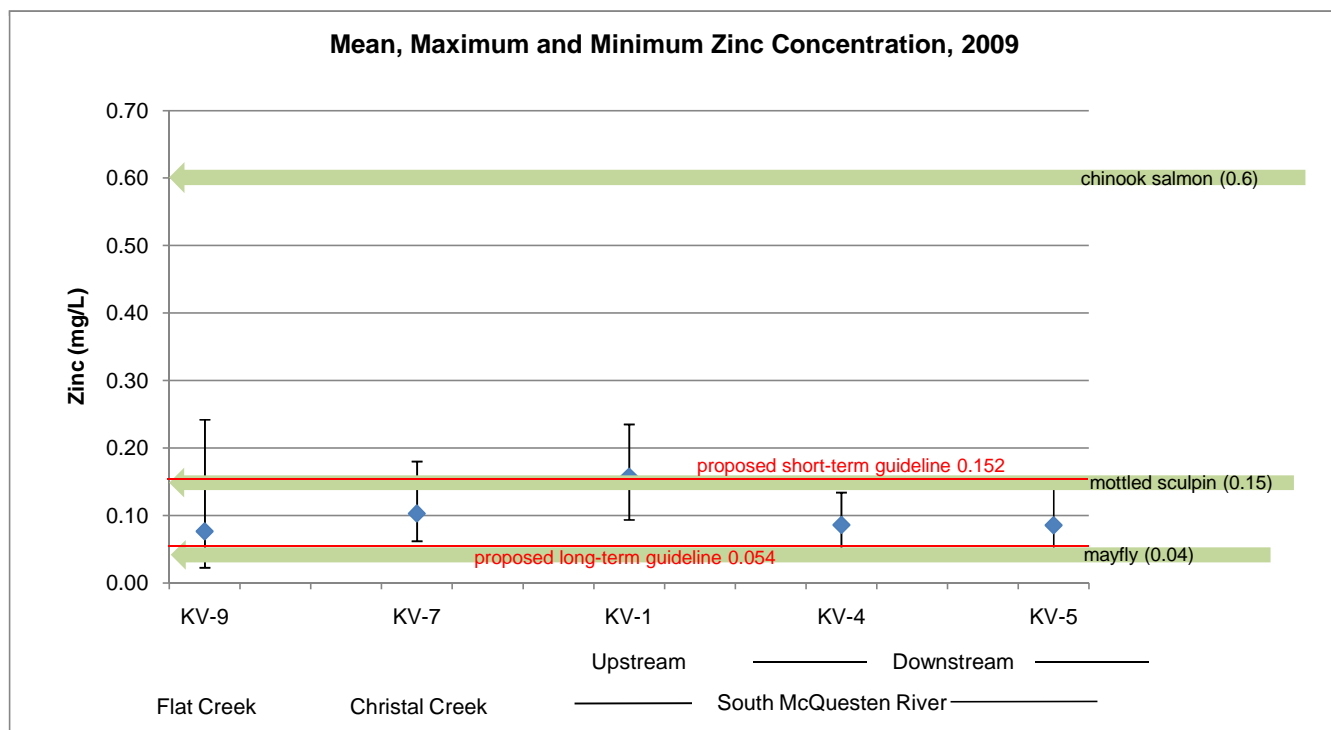
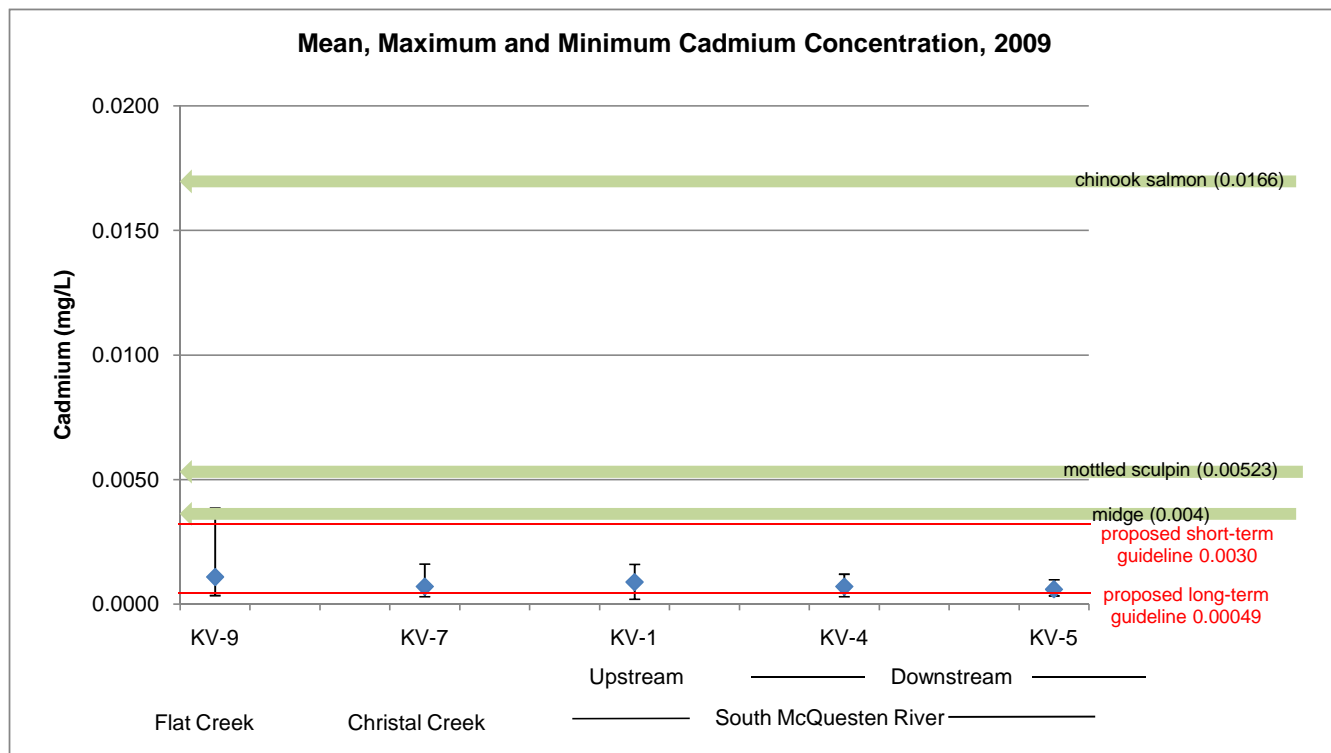


Figure 3.5: Total cadmium and zinc concentrations at selected UKHM stations relative to proposed CWQG and toxicity thresholds reported in the literature for selected species based on a water hardness of 300 mg/L.

The aquatic toxicities of zinc and cadmium are lower at higher water hardness (Appendix Figure D.1 and D.2), so effects to resident biota are likely less than would be predicted by CWQG established for a hardness of 100 mg/L. Therefore, potential impacts to aquatic biota near UKHM are discussed in more detail below relative to observed concentrations of cadmium, zinc and hardness and toxicity data presented in the literature.

In 2009, mean **cadmium** concentrations in Flat (KV-9) and Christal (KV-7) creeks were 0.0011 and 0.0007 mg/L, respectively (Figure 3.4). These concentrations were above the toxic thresholds of some aquatic biota at water hardnesses up to 300 mg/L (Table 3.1). Although this suggests that current water quality may affect some sensitive biota within these tributaries, the majority of species would not likely be affected, even during periods when water hardness is below average for each tributary (Table 3.1). In particular, fish species such as sculpin³ and Chinook salmon would not be affected at cadmium and hardness levels typically observed in each creek (Figure 3.5). While long-term exposure data were not available for arctic grayling, a short-term (4-day) test in very low water hardness (41 mg/L) resulted in mortality to 50% of exposed organisms (LC50) at 0.004 mg/L (Buhl and Hamilton 1991), suggesting arctic grayling are also not affected by current water quality downstream of UKHM. Overall, it is expected that if cadmium concentrations remain stable or decline over time that further biological impacts will be minimal and existing biological communities will be protected.

Mean **zinc** concentrations in 2009 were 0.077 and 0.103 mg/L in Flat Creek and Christal Creek respectively (Figure 3.4). The majority of species tested would not likely be affected by such concentrations, particularly at typical water hardnesses of 300 mg/L (Table 3.2). This includes locally important fish species such as sculpin and Chinook salmon. Long-term exposure data are lacking for Arctic grayling, but 96-h LC50s of 0.11 to 0.17 mg/L zinc were reported in exposures involving very low water hardness, so effect concentrations would be higher (lower toxicity) at water hardnesses typically observed in Flat and Crystal creeks. Generally, the effects associated with current zinc levels are probably low, although the upper concentrations reported in both creeks have the potential to affect locally important species if sustained. Nonetheless, as indicated for cadmium, current biological communities will be protected if future concentrations remain stable or decline relative to current levels.

The above results are consistent with continued observations of both sculpin and grayling in Flat and Christal creeks (Minnow 2009).

³ Toxicity data are available for mottled sculpin, whereas slimy sculpin is the species present near UKHM.

Table 3.1: Lowest effect endpoint reported for each freshwater species after a long-term exposure to cadmium (adapted from Minnow 2010).

| Species Common Name | Scientific Name | Duration | Endpoint | Observed Effect | Test Hardness (mg/L) | Reported Effect Concentration (mg/L) | Hardness-Adjusted Effect Concentration | | | |
|-------------------------|------------------------------------|----------|----------|------------------------------------------|----------------------|--------------------------------------|--------------------------------------------------------------|---------------------------------------------------------------|---------------------------------------------------------------|---------------------------------------------------------------|
| | | | | | | | Effect Concentration (mg/L) at 50 mg/L Hardness ^a | Effect Concentration (mg/L) at 100 mg/L Hardness ^a | Effect Concentration (mg/L) at 200 mg/L Hardness ^a | Effect Concentration (mg/L) at 300 mg/L Hardness ^a |
| Water flea | <i>Daphnia magna</i> | 7 d | EC10 | Reproduction - Brood size | 179 | 0.00014 | 0.00004 | 0.00008 | 0.00016 | 0.00024 |
| Water flea | <i>Ceriodaphnia reticulata</i> | 7 d | MATC | Reproduction - Number of young per adult | 240 | 0.00043 | 0.00009 | 0.00018 | 0.00036 | 0.00054 |
| Amphipod - scud | <i>Hyalella azteca</i> | 28 d | IC25 | Biomass, decrease in | 280 | 0.00051 | 0.00009 | 0.00018 | 0.00036 | 0.00055 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | 65 wks | MATC | Reproduction - delay in oogenesis | 250 | 0.00091 | 0.00018 | 0.00036 | 0.00072 | 0.00110 |
| Midge | <i>Chironomus tentans</i> | 60 d | IC25 | Hatching success | 280 | 0.004 | 0.001 | 0.001 | 0.003 | 0.004 |
| Mottled sculpin | <i>Cottus bairdi</i> | 21 d | EC50 | Biomass, decrease in | 104 | 0.00177 | 0.00084 | 0.00170 | 0.00346 | 0.00523 |
| Atlantic salmon | <i>Salmo salar</i> | 496 d | LOEC/L | Weight and Length | 28 | 0.00047 | 0.00085 | 0.00173 | 0.00351 | 0.00532 |
| Bull trout | <i>Salvelinus confluentus</i> | 55 d | MATC | Growth | 30.6 | 0.00055 | 0.00091 | 0.00184 | 0.00375 | 0.00567 |
| Green hydra | <i>Hydra viridissima</i> | 7 d | NOEC/L | Population growth inhibition | 19.5 | 0.0004 | 0.0010 | 0.0021 | 0.0043 | 0.0066 |
| Amphipod - gammarid | <i>Echinogammarus meridionalis</i> | 6 d | LOEC/L | Feeding inhibition | 263.4 | 0.0064 | 0.0012 | 0.0024 | 0.0048 | 0.0073 |
| Amphipod - gammarid | <i>Gammarus pulex</i> | 5 d | LOEC/L | Mortality | 269.2 | 0.0075 | 0.0013 | 0.0027 | 0.0055 | 0.0084 |
| Brown trout | <i>Salmo trutta</i> | 30 d | IC20 | Biomass, decrease in | 29.2 | 0.0009 | 0.0015 | 0.0031 | 0.0062 | 0.0094 |
| Brook Trout | <i>Salvelinus fontinalis</i> | 126 d | MATC | Biomass, decrease in | 45 | 0.002 | 0.002 | 0.005 | 0.009 | 0.014 |
| Coho salmon | <i>Oncorhynchus kisutch</i> | 27 d | MATC | Biomass, decrease in | 45 | 0.0021 | 0.0023 | 0.0048 | 0.0097 | 0.0146 |
| Chinook salmon | <i>Oncorhynchus tshawytscha</i> | 8 d | LC10 | Mortality | 23 | 0.0012 | 0.0027 | 0.0054 | 0.0110 | 0.0166 |
| Water flea | <i>Daphnia pulex</i> | 14 d | MATC | Reproduction - Number of young per adult | 240 | 0.0137 | 0.0028 | 0.0056 | 0.0114 | 0.0172 |
| Green algae | <i>Ankistrodesmus falcatus</i> | 96 h | NOEC/L | Growth | 118.0 | 0.01 | 0.00 | 0.01 | 0.02 | 0.03 |
| Water flea | <i>Ceriodaphnia dubia</i> | 14 d | MATC | Reproduction | 17.0 | 0.002 | 0.006 | 0.012 | 0.025 | 0.038 |
| White Sucker | <i>Catostomus commersoni</i> | 40 h | MATC | Biomass, decrease in | 45 | 0.0071 | 0.0079 | 0.0161 | 0.0327 | 0.0494 |
| Northern pike | <i>Esox lucius</i> | 35 d | MATC | Biomass, decrease in | 45 | 0.0074 | 0.0082 | 0.0167 | 0.0340 | 0.0515 |
| Lake Trout | <i>Salvelinus namaycush</i> | 41 d | MATC | Biomass, decrease in | 45 | 0.0074 | 0.0082 | 0.0167 | 0.0340 | 0.0515 |
| Marsh snail | <i>Lymnaea palustris</i> | 4 weeks | EC50 | Growth | 284 | 0.0582 | 0.0098 | 0.0200 | 0.0407 | 0.0616 |
| Great pond snail | <i>Lymnaea stagnalis</i> | 4 weeks | NOEC/L | Growth | 284 | 0.08 | 0.01 | 0.03 | 0.06 | 0.08 |
| Midge | <i>Chironomus riparius</i> | 17 d | MATC | Mortality | 98.0 | 0.0474 | 0.0238 | 0.0484 | 0.0983 | 0.1489 |
| Duckweed | <i>Lemna minor</i> | 7 d | EC50 | Growth rate | 166.0 | 0.214 | 0.063 | 0.127 | 0.259 | 0.392 |
| Northwestern salamander | <i>Ambystoma gracile</i> | 24 d | MATC | Weight | 45 | 0.0972 | 0.1083 | 0.2200 | 0.4471 | 0.6769 |

^a If reported toxicity applied to a different water hardness than that shown, a hardness adjusted toxicity value was calculated using the following equation: $\text{EXP}(\text{LN}(\text{effect conc}) - (1.023) * (\text{LN}(\text{measured water hardness}) - \text{LN}(\text{desired water hardness})))$

Table 3.2: Lowest effect endpoint reported for each freshwater species after a long-term exposure to zinc (adapted from Minnow 2010).

| Species Common Name | Scientific Name | Duration | Endpoint | Observed Effect | Test Reported | | Hardness-Adjusted Effect Concentration | | | |
|---------------------|----------------------------------------|----------|----------|------------------------------------------|-----------------|-----------------------------|--------------------------------------------------------------|---------------------------------------------------------------|---------------------------------------------------------------|---------------------------------------------------------------|
| | | | | | Hardness (mg/L) | Effect Concentration (mg/L) | Effect Concentration (mg/L) at 50 mg/L Hardness ^a | Effect Concentration (mg/L) at 100 mg/L Hardness ^a | Effect Concentration (mg/L) at 200 mg/L Hardness ^a | Effect Concentration (mg/L) at 300 mg/L Hardness ^a |
| Green algae | <i>Pseudokirchneriella subcapitata</i> | 7 d | EC10 | Growth | NR | 0.0011 | 0.0011 | 0.0011 | 0.0011 | 0.0011 |
| Mayfly | <i>Epeorus latifolium</i> | 4 weeks | IC10 | emergence | 83 | 0.0144 | 0.0094 | 0.0169 | 0.0303 | 0.0427 |
| Water flea | <i>Ceriodaphnia dubia</i> | 4 weeks | LOEC | Reproduction - Number of young per adult | 97.6 | 0.025 | 0.014 | 0.026 | 0.046 | 0.065 |
| Green alga | <i>Chlorella vulgaris</i> | 72 h | EC50 | biomass | NR | 0.034 | 0.034 | 0.034 | 0.034 | 0.034 |
| Water flea | <i>Daphnia magna</i> | 50 d | MATC | Reproduction - Brood size | 51.9 | 0.0217 | 0.0210 | 0.0378 | 0.0679 | 0.0957 |
| Snail | <i>Potamopyrgus jenkinsi</i> | 12 weeks | MATC | Growth | 159 | 0.091 | 0.034 | 0.061 | 0.110 | 0.156 |
| Chironomids | <i>Tanytarsus dissimilis</i> | 10 d | LC50 | Mortality | 46.8 | 0.0368 | 0.0389 | 0.0700 | 0.1257 | 0.1772 |
| Rotifer | <i>Brachionus havanaensis</i> | 18 d | EC10 | Population growth inhibition | NR | 0.0782 | 0.0782 | 0.0782 | 0.0782 | 0.0782 |
| Green alga | <i>Chlorella pyrenoidosa</i> | 24 h | MATC | Cell density | 25.51 | 0.0283 | 0.0500 | 0.0898 | 0.1615 | 0.2275 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | 30 d | LC10 | Mortality | 31.72 | 0.0345 | 0.0507 | 0.0911 | 0.1638 | 0.2309 |
| Green algae | <i>Scenedesmus quadricauda</i> | 15 d | IC10 | Growth | NR | 0.0961 | 0.0961 | 0.0961 | 0.0961 | 0.0961 |
| Chironomids | <i>Chironomus riparius</i> | 11 weeks | LOEC | Development | NR | 0.100 | 0.100 | 0.100 | 0.100 | 0.100 |
| Mottled sculpin | <i>Cottus bairdi</i> | 30 d | LC50 | Mortality | 48.6 | 0.032 | 0.033 | 0.059 | 0.106 | 0.149 |
| Green hydra | <i>Hydra viridissima</i> | 7 d | EC10 | Population growth inhibition | 20 | 0.0522 | 0.1134 | 0.2038 | 0.3664 | 0.5163 |
| Chinook salmon | <i>Oncorhynchus tshawytscha</i> | 200 h | LC10 | Mortality | 23 | 0.068 | 0.131 | 0.236 | 0.424 | 0.597 |
| Amphipod | <i>Hyalella azteca</i> | 7 d | LC50 | Mortality | 18 | 0.056 | 0.133 | 0.239 | 0.429 | 0.605 |
| Duckweed | <i>Lemna minor</i> | 7 d | IC10 | Growth | NR | 0.318 | 0.318 | 0.318 | 0.318 | 0.318 |
| Brook Trout | <i>Salvelinus fontinalis</i> | 24 w | MATC | egg fragility | 45.9 | 0.174 | 0.187 | 0.336 | 0.604 | 0.852 |
| Pink hydra | <i>Hydra vulgaris</i> | 7 d | EC10 | Population growth inhibition | 20 | 0.1779 | 0.3863 | 0.6943 | 1.2481 | 1.7588 |
| Common duckmeat | <i>Spirodela polyrrhiza</i> | 4 d | IC50 | Growth | NR | 0.935 | 0.935 | 0.935 | 0.935 | 0.935 |
| Cutthroat trout | <i>Oncorhynchus clarkii</i> | 14 d | LC10 | Mortality | 40 | 0.453 | 0.547 | 0.983 | 1.768 | 2.491 |
| Bryozoan | <i>Pectinatella magnifica</i> | 96 h | LC10 | Mortality | 205 | 2.286 | 0.693 | 1.245 | 2.239 | 3.155 |
| Star duckweed | <i>Lemna trisulca</i> | 14 d | EC50 | final yield (oven dry weight) | 20.37 | 0.327 | 0.699 | 1.256 | 2.258 | 3.183 |
| Atlantic salmon | <i>Salmo salar</i> | 14 d | LC50 | Mortality | 351 | 3.640 | 0.700 | 1.258 | 2.262 | 3.187 |
| Snail | <i>Physa gyrina</i> | 30 d | LC50 | Mortality | 36 | 0.771 | 1.018 | 1.830 | 3.289 | 4.635 |
| Bryozoan | <i>Plumatella emarginata</i> | 96 h | LC10 | Mortality | 205 | 3.474 | 1.053 | 1.893 | 3.402 | 4.794 |
| Bryozoan | <i>Lophopodella carteri</i> | 96 h | LC50 | Mortality | 205 | 4.093 | 1.241 | 2.230 | 4.008 | 5.649 |
| Diatom | <i>Cyclotella meneghiniana</i> | 5 d | LC10 | Growth rate | 121 | 2.803 | 1.327 | 2.386 | 4.288 | 6.043 |
| Mayfly | <i>Rhithrogena hageni</i> | 10 d | EC10 | Mortality | 44.4 | 2.069 | 2.288 | 4.113 | 7.392 | 10.417 |
| Green alga | <i>Chlamydomonas sp.</i> | 10 d | LC10 | Growth rate | 121 | 8.381 | 3.968 | 7.133 | 12.821 | 18.068 |
| Crayfish | <i>Orconectes virilis</i> | 14 d | LC10 | Mortality | 26 | 9.920 | 17.249 | 31.006 | 55.733 | 78.540 |

^a If reported toxicity applied to a different water hardness than that shown, a hardness-adjusted toxicity value was calculated using the following equation: $EXP(LN(effect\ conc)-(0.846)*(LN(measured\ water\ hardness)-LN(desired\ water\ hardness)))$

NR - not reported

3.2.3 Proposed Approach to Water Quality Goals

Based on the previous sections, water quality goals for Flat and Christal creeks should aim to prevent further degradation of water quality to protect the diversity and abundance of the existing resident biota. In order to accomplish this, a number of water quality goals are proposed:

- Monthly water samples should be collected in Flat Creek at KV-9 and in Christal Creek at KV-7 and concentrations of all substances associated with UKHM, particular cadmium and zinc, should be evaluated annually.
- Mean concentrations of cadmium and zinc should be equal to or less than the previous year based on statistical comparison of means by analysis of variance (ANOVA). An increase in mean concentrations, even if not statistically significant, should be investigated to determine if may be an early indicator of an increasing trend related to historic or possibly other sources within the watershed (e.g., possibly through examining trends in loads or concentrations from specific upstream source areas). Changes in background concentrations should also be considered in this assessment such that if an increase is observed in the background concentration of cadmium and/or zinc, the concentrations within the tributaries are normalized for this increase.
- Trends in the concentrations of cadmium and zinc at KV-9 and KV-7 should be evaluated by a qualified professional at a frequency consistent with detailed watershed reporting as recommended in the Long-Term Aquatic Monitoring Program (LTAMP; Minnow 2011).
- An increasing trend should trigger investigation and, if appropriate, remediation of the cause.

The effects of these goals should be verified through routine biological monitoring (i.e., LTAMP) the scope of which should include:

- The number of benthic invertebrate taxa (i.e., diversity) measured through routine environmental monitoring programs should not be less than previously observed at each area (KV-9 and KV-7) using appropriate statistical methods and assuming the use of standardized timing and methods of collection. Similar to water quality, this assessment will need to consider changes in reference locations such that changes associated with natural temporal variability are not attributed to historic UKHM influence.

- One of the most sensitive organisms to zinc is a mayfly (*Epeorus latifolium*) which is a member of the family Heptageniidae. This family of mayflies is present in the South McQuesten River watershed and several local streams (Minnow 2011, in preparation; Figure 3.6). The abundance of organisms within this family should be monitored over time as a potential indicator of zinc toxicity at KV-7. The objective would be to see an increase in Heptageniidae over time. It is not recommended that this indicator be used at KV-9 as the habitat at this station is generally not suitable to this family (slower flow and soft-bottom substrate).

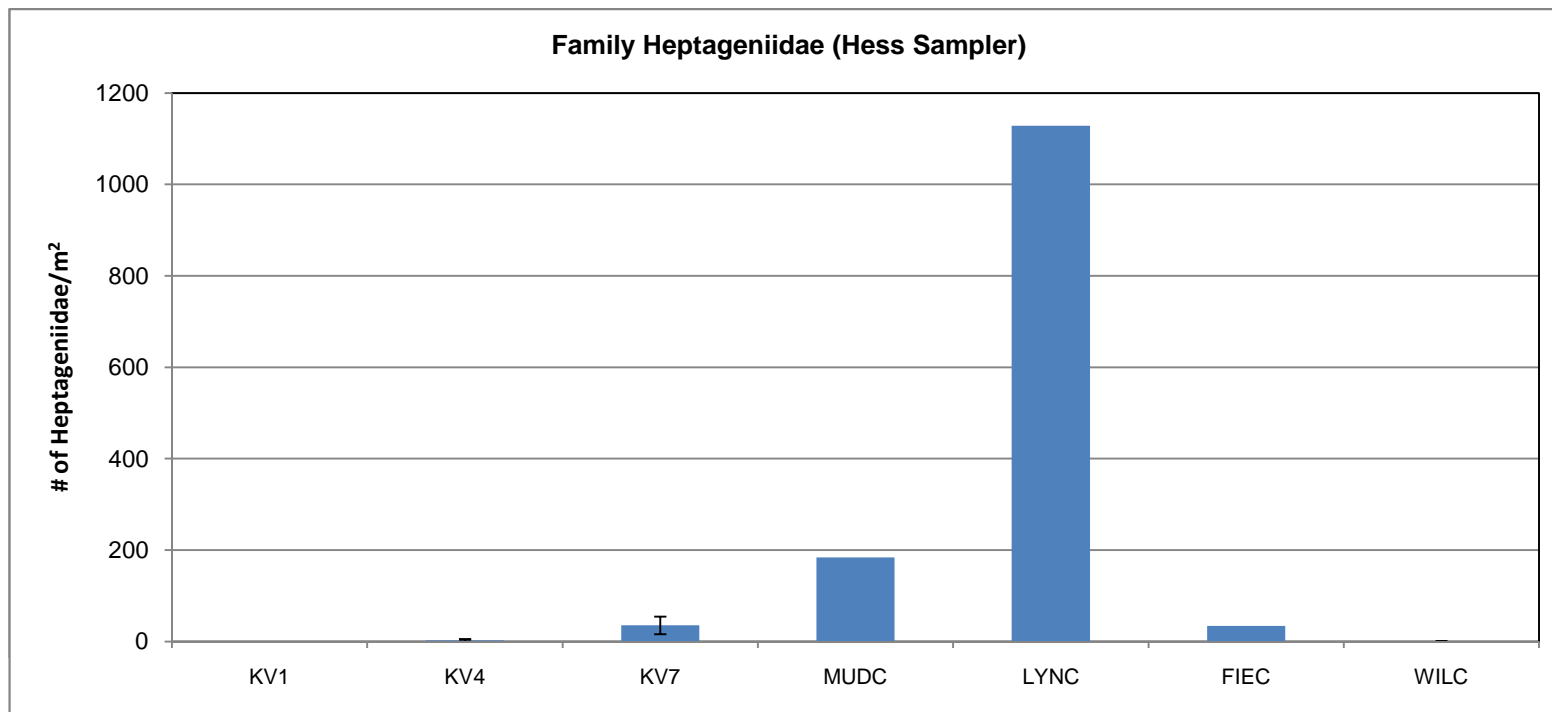


Figure 3.6: Number of heptageniidae identified in samples collected (by Hess) in August 2009 in the vicinity of United Keno Hill Mines

4.0 CONCLUSIONS

Based on the assessment of water quality presented in this report, the following conclusions are provided:

- Concentrations of cadmium and zinc have been increasing in the South McQuesten River downstream of UKHM (KV-4, KV-5) as a result of increasing concentrations upstream at KV-1. The concentrations and the rate of increase (slope) at downstream stations are equal to or lower than those upstream, indicating that the mine complex is not causing measurable increases in cadmium and zinc concentrations.
- Due to increasing concentrations upstream, a single water quality criterion can not reasonably be established for the South McQuesten River downstream of UKHM and an alternative approach is required.
- Water quality goals for cadmium and zinc should be linked to upstream concentrations and allow for no further degradation of water quality. The mean concentrations of cadmium and zinc should not exceed those measured upstream at KV-1 and the slope of concentrations relative to time at downstream stations should be equal to or less than that measured upstream at KV-1.
- Lightning Creek was not considered in this assessment as the existing data base indicated that concentrations within Lightning Creek were close to the CWQG. However, the data base for the creek was limited with respect to the extent of data and appropriate method detection limits. As additional monitoring data is obtained, the status of Lightning Creek should be reassessed with respect to the need for water quality objectives.
- Over the past twenty years concentrations of cadmium and zinc have been elevated well above the current CWQG (i.e., mean values more than ten times the CWQG) in Flat Creek and Christal Creek (Minnow 2008). It is expected that concentrations will decrease over time, but they are not expected to achieve the CWQG in the near future. Therefore, an alternative water quality goal is needed.
- Protection of current biological diversity and abundance in Flat and Christal creeks is predicated on prevention of further degradation of water quality. Therefore, annual mean concentrations of cadmium and zinc associated with the historic UKHM complex should not increase relative to the previous year and background conditions and trends over time should be stable or declining.

5.0 REFERENCES

- Anderson, R.L., C.T. Wallbridge, and J.T. Fiandt. 1980. Survival and growth of *tanytarsus dissimilis* (Chironomidae) exposed to copper, cadmium, zinc, and lead. Archives of Environmental Contamination and Toxicology 9:329-335.
- Baer, K.N., M.C. Ziegenfuss, D. Banks, and Z. Ling. 1999. Suitability of high-hardness combo medium for ecotoxicity testing using algae, daphnids, and fish. Bulletin of Environmental Contamination and Toxicology 63:289-296.
- Barata, C., and D.J. Baird. 2000. Determining the ecotoxicological mode of action of chemicals from measurements made on individuals: Results from instar-based tests with *Daphnia magna* Straus. Aquatic Toxicology 48:195-209.
- Belanger, S.E., and D.S. Cherry. 1990. Interacting effects of pH acclimatation, pH, and heavy metals on acute and chronic toxicity to *Ceriodaphnia dubia* (Cladocera). Journal of Crustacean Biology 10:225-235.
- Bengtsson, B.E. 1974. Effect of zinc on growth of the minnow *Phoxinus phoxinus*. OIKOS 25:370-373.
- Benhra, A., C.M. Radetski, and J.F. Férard. 1997. Cryoalgotox: Use of cryopreserved alga in a semistatic microplate test. Environmental Toxicology and Chemistry 16:505-508.
- Benoit, D.A. and G.W. Holcombe. 1978. Toxic effects of zinc on fathead minnows *Pimephales promelas* in soft water. Journal of Fish Biology 13:701-708.
- Besser, J.M., C.A. Mebane, D.R. Mount, C.D. Ivey, J.L. Kunz, I.E. Greer, T.W. May, and C.G. Ingersoll. 2007. Sensitivity of mottled sculpins (*Cottus bairdi*) and rainbow trout (*Oncorhynchus mykiss*) to acute and chronic toxicity of cadmium, copper, and zinc. Environmental Toxicology and Chemistry 26:1657-1665.
- Biesinger, K.E., and G.M. Christensen. 1972. Effects of various metals on survival, growth, reproduction and metabolism of *Daphnia magna*. Journal of the Fisheries Research Board of Canada 29:1691-1700.

- Borgmann, U., K.M. Ralph, and W.P. Norwood. 1989. Toxicity test procedures for *Hyalella azteca*, and chronic toxicity of cadmium and pentachlorophenol to *H. azteca*, *Gammarus fasciatus*, and *Daphnia magna*. Archives of Environmental Contamination and Toxicology 18:756-764.
- Borgmann, U., W.P. Norwood, and C. Clarke. 1993. Accumulation, regulation and toxicity of copper, zinc, lead and mercury in *Hyalella azteca*. Hydrobiologia 259:79-89.
- Borgmann, U., Y. Couillard, P. Doyle, and D.G. Dixons. 2005. Toxicity of sixty-three metals and metalloids to *Hyalella azteca* at two levels of water hardness. Environmental Toxicology and Chemistry 24(3):641-652.
- Brinkman, S., and J. Woodling. 2005. Zinc toxicity to the mottled sculpin (*Cottus bairdi*) in high-hardness water. Environmental Toxicology and Chemistry 24(6):1515-1517.
- Brinkman, S.F. and D.L. Hansen. 2007. Toxicity of cadmium to early life stages of brown trout (*Salmo trutta*) at multiple water hardnesses. Environmental Toxicology and Chemistry 26:1666-1671.
- Brinkman, S. F., and W.D. Johnston. 2008. Acute toxicity of aqueous copper, cadmium, and zinc to the mayfly *Rhithrogena hageni*. Archives of Environmental Contamination and Toxicology 54:466-472.
- Brown, V., D. Shurben, W. Miller, and M. Crane. 1994. Cadmium toxicity to rainbow trout *Oncorhynchus mykiss* Walbaum and brown trout *Salmo trutta* L. over extended exposure periods. Ecotoxicology and Environmental Safety 29:38-46.
- Buhl, K.J. and S.J. Hamilton. 1991. Relative sensitivity of early life stages of arctic grayling, coho salmon and rainbow trout to nine inorganics. Ecotoxicology and Environmental Safety 22: 184-197
- Burns, B., 2008. Keno Valley Stream Sediment and Benthic Invertebrate Monitoring Programs, 2007. Laberge Environmental Services, Whitehorse, YT. March 2008.
- Cairns, J., A.L. Buikema, A.G. Heath, and J.I. Parker. 1978. Effects of temperature on aquatic organisms to selected chemicals. Blacksburg, VA, USA, Virginia Water

Resources Research Center, Virginia Polytechnic Institute and State University. Bulletin 106.

Cairns, J., R.R. Garton, and R.A. Tubb. 1982. Use of fish ventilation frequency to estimate chronically safe toxicant concentrations. Transactions of the American Fisheries Society 111:70-77.

CCME (Canadian Council of Ministers of the Environment). 2003. Guidance on the Site-Specific Application of Water Quality Guidelines in Canada: Procedures for Deriving Numerical Water Quality Objectives. In: Canadian Environmental Quality Guidelines. 1999. Canadian Council of Ministers of the Environment, Winnipeg.

Canadian Council of the Ministers of the Environment (CCME). 1999. Canadian water quality guidelines for the protection of aquatic life for cadmium, published in Canadian Environmental Quality Guidelines. Winnipeg, Manitoba, Canadian Council of the Ministers of the Environment.

Chapman, G.A. 1978. Toxicities of cadmium, copper and zinc to four juvenile stages of Chinook salmon and steelhead. Transactions of the American Fisheries Society 107:841-847.

Chapman, G.A., S. Ota, and F. Recht. 1980. Effects of Water Hardness on the Toxicity of Metals to *Daphnia magna* (Status report - January 1980). Corvallis, Oregon, Corvallis Environmental Research Laboratory.

Chen, C.Y., K.C. Lin, and D.T. Yang. 1997. Comparison of the relative toxicity relationships based on batch and continuous algal toxicity tests. Chemosphere 35(9):1959-1965.

Chiaudani, G., and M. Vighi. 1978. The use of *Selenastrum capricornutum* batch cultures in toxicity studies. Mitt Internat Verein Limnol 21:316-329.

Coeurdassier, M., A. De Vaufleury, and P.M. Badot. 2003. Bioconcentration of cadmium and toxic effects on life-history traits of pond snails (*Lymnaea palustris* and *Lymnaea stagnalis*) in laboratory bioassays. Archives of Environmental Contamination and Toxicology 45:102-109.

- De Schamphelaere, K.A.C., and C.R. Janssen. 2004. Bioavailability and chronic toxicity of zinc to juvenile rainbow trout (*Oncorhynchus mykiss*): Comparison with other fish species and development of a Biotic Ligand Model. *Environmental Science and Technology* 38:6201-6209.
- De Schamphelaere, K.A.C., D.G. Heijerick, and C.R. Janssen. 2004. Comparison of the effect of different pH buffering techniques on the toxicity of copper and zinc to *Daphnia magna* and *Pseudokirchneriella subcapitata*. *Ecotoxicology* 13:697-705.
- De Schamphelaere, K.A.C., S. Lofts, and C.R. Janssen. 2005. Bioavailability models for predicting acute and chronic toxicity of zinc to algae, daphnids, and fish in natural surface waters. *Environmental Toxicology and Chemistry* 24(5):1190-1197.
- Dirilgen, N., and Y. Inel. 1994. Effects of zinc and copper on growth and metal accumulation in duckweed, *Lemna minor*. *Bulletin of Environmental Contamination and Toxicology* 53:442-449.
- Dorgelo, J., H. Meester, and C.V. Velzen. 1995. Effects of diet and heavy metals on growth rate and fertility in the deposit-feeding snail *Potamopyrgus jenkinsi* (Smith) (Gastropoda: Hydrobiidae). *Hydrobiologia* 316:199-210.
- Drost, W., M. Matzke, and T. Backhaus. 2007. Heavy metal toxicity to *Lemna minor*: studies on the time dependence of growth inhibition and the recovery after exposure. *Chemosphere* 67:36-43.
- Eaton, J.G., J.M. McKim, and G.W. Holcombe. 1978. Metal toxicity to embryos and larvae of seven freshwater fish species - I. Cadmium. *Bulletin of Environmental Contamination and Toxicology* 19:95-103.
- Elnabarawy, M.T., A.N. Welter, and R.R. Robideau. 1986. Relative sensitivity of three daphnid species to selected organic and inorganic chemicals. *Environmental Toxicology and Chemistry* 5(393):398.
- Enserink, E.L., J.L. Maas-Diepeveen, and C.J. Van Leeuwen. 1991. Combined effects of metals; an ecotoxicological evaluation. *Water Research* 25(6):679-687.

- Environment Canada. 2009a. Canadian water quality guideline: Zinc. Scientific Supporting Document (draft January 2009). Ecosystem Health: Science-based Solutions. National Guidelines and Standards Office, Environment Canada, Ottawa.
- Environment Canada. 2009b. Canadian water quality guideline: Cadmium. Scientific Supporting Document (draft January 2009). Ecosystem Health: Science-based Solutions. National Guidelines and Standards Office, Environment Canada, Ottawa.
- Errécalde, O., M. Seidl, and P.G.C. Campbell. 1998. Influence of a low molecular weight metabolite (citrate) on the toxicity of cadmium and zinc to the unicellular green alga *Selenastrum capricornutum*: An exception to the free-ion model. *Water Research* 32(2):419-429.
- Felten, V., G. Charmantier, R. Mons, A. Geffard, P. Rousselle, M. Coquery, J. Garric, and O. Geffard. 2007. Physiological and behavioural responses of *Gammarus pulex* (Crustacea Amphipoda) exposed to cadmium. *Aquatic Toxicology* 86(3):413-425.
- Gaur, J.P., N. Noraho, and Y.S. Chauhan. 1994. Relationship between heavy metal accumulation and toxicity in *Spirodela polyrhiza* (L.) Schleid. and *Azolla pinnata* R. Br. *Aquatic Botany* 49(2-3):183-192.
- Hansen, J.A., P.G. Welsh, J. Lipton, and M.J. Suedkamp. 2002. The effects of long-term cadmium exposure on the growth and survival of juvenile bull trout (*Salvelinus confluentus*). *Aquatic Toxicology* 58:165-174.
- Hatakeyama, S. 1989. Effect of copper and zinc on the growth and emergence of *Epeorus latifolium* (Ephemeroptera) in an indoor model stream. *Hydrobiologia* 174(1):17-27.
- Heijerick, D.G., K.A.C. De Schampelaere, and C.R. Janssen. 2002. Biotic ligand model development predicting Zn toxicity to the algae *Pseudokirchneriella subcapitata*: possibilities and limitations. *Comparative Biochemistry and Physiology - C Toxicology and Pharmacology* 133:207-218.
- Heijerick, D., C.R. Janssen, and W.M. De Coen. 2003. The combined effects of hardness, pH and dissolved organic carbon on the chronic toxicity of Zn to *D.*

- magna*: Development of a surface response model. Archives of Environmental Contamination and Toxicology 44:210-217.
- Heijerick, D.G., K.A.C. De Schamphelaere, P.A. Van Sprang, and C.R. Janssen. 2005. Development of a chronic zinc biotic ligand model for *Daphnia magna*. Ecotoxicology and Environmental Safety 62:1-10.
- Hodson, P.V., and J.B. Sprague. 1975. Temperature-induced changes in acute toxicity of zinc to Atlantic salmon (*Salmo salar*). Journal of the Fisheries Research Board of Canada 32:1-10.
- Holcombe, G.W., D.A. Benoit, and E.N. Leonard. 1979. Long-term effects of zinc exposures on brook trout (*Salvelinus fontinalis*). Transactions of the American Fisheries Society 108:76-87.
- Holdway, D.A., K. Lok, and M. Semaan. 2001. The acute and chronic toxicity of cadmium and zinc to two hydra species. Environmental Toxicology 16(6):557-565.
- Huang, W., L. Cao, X. Shan, Z. Xiao, Q. Wang, and S. Dou. 2009. Toxic effects of zinc on the development, growth, and survival of red sea bream *Pagrus major* embryos and larvae. Arch. Environ. Contam. Toxicol. 58(1):140-150.
- Huebert, D.B., and J.M. Shay. 1992. Zinc toxicity and its interaction with cadmium in the submerged aquatic macrophyte *Lemna trisulca*. Environmental Toxicology and Chemistry 11:715-720.
- Ince, N.H., N. Dirilgen, I.G. Apikyan, G. Tezcanli, and B. Ustün. 1999. Assessment of toxic interactions of heavy metals in binary mixtures: A statistical approach. Archives of Environmental Contamination and Toxicology 36:365-372.
- Ingersoll, C.G., and N. Kemble. 2001. Revised Description of Toxicity Data on Cadmium: Chronic Water-only Exposures with the Amphipod *Hyaella azteca* and the Midge *Chironomus tentans*. Roberts, C. Columbia, Missouri, United States Department of the Interior, U.S. Geological Survey.
- Juárez-Franco, M.F., S.S.S. Sarma, and S. Nandini. 2007. Effect of cadmium and zinc on the population growth of *Brachionus havanaensis* (Rotifera: Brachionidae).

Journal of Environmental Science and Health Part A-Environmental Science and Engineering & Toxic and Hazardous Substance Control 42:1489-1493.

Källqvist, T. 2007. Effect of water hardness on the toxicity of cadmium to the alga *Pseudokirchneriella subcapitata*. Oslo, Norwegian Institute for Water Research. SNO 5422-2007.

Kraak, M.H.S., Y.A. Wink, S.C. Stuijzand, M.C. Buckert-de Jong, C.J. De Groot, and W. Admiraal. 1994. Chronic ecotoxicity of Zn and Pb to the zebra mussel *Dreissena polymorpha*. Aquatic Toxicology 30:77-89.

Kühn, R., M. Pattard, K.D. Pernak, and A. Winter. 1989. Results of the harmful effects of water pollutants to *Daphnia magna* in the 21 day reproduction test. Water Research 23:501-510.

Lazorchak, J.M., and M.E. Smith. 2007. Rainbow trout (*Oncorhynchus mykiss*) and brook trout (*Salvelinus fontinalis*) 7-day survival and growth test method. Archives of Environmental Contamination and Toxicology 53:397-405.

Lin, K.C., Y.L. Lee, and C.Y. Chen. 2007. Metal toxicity to *Chlorella pyrenoidosa* assessed by a short-term continuous test. Journal of Hazardous Materials 142:236-241.

Magliette, R.J., F.G. Doherty, D. McKinney, and E.S. Venkataramani. 1995. Need for environmental quality guidelines based on ambient freshwater quality criteria in natural waters- case study "zinc". Bulletin of Environmental Contamination and Toxicology 54:626-632.

Marshall, J.S., J.I. Parker, D.L. Mellinger, and C. Lei. 1983. Bioaccumulation and effects of cadmium and zinc in a Lake Michigan plankton community. Canadian Journal of Fisheries and Aquatic Sciences 40(1469):1479.

Masters, J.A., M.A. Lewis, D. Davidson, and R.D. Bruce. 1991. Validation of a four-day *Ceriodaphnia* toxicity test and statistical considerations in data analysis. Environmental Toxicology and Chemistry 10:47-55.

- Mebane, C.A., D.P. Hennessy, and F.S. Dillon. 2008. Developing acute-to-chronic toxicity ratios for lead, cadmium and zinc using rainbow trout, a mayfly and a midge. *Water, Air, and Soil Pollution* 188:41-66.
- Minnow Environmental Inc., 2008. Water Quality Assessment Report for United Keno Hill Mines. Prepared for Elsa Reclamation and Development Company, Whitehorse, YT, July 2008.
- Minnow Environmental Inc., 2009. Aquatic Resource Assessment Report for United Keno Hill Mines. Prepared for Elsa Reclamation and Development Company, Whitehorse, YT, March 2009.
- Minnow Environmental Inc. 2010. Aquatic Impact Assessment Related to Current and Future Zinc, Cadmium and Sulphate Concentrations at Faro Mine, Yukon. Prepared for the Assessment and Abandoned Mines Branch, Energy Mines and Resources, Government of Yukon, Whitehorse, Yukon, February 2010.
- Minnow Environmental Inc., 2011. Interim Long-term Aquatic Monitoring Program for United Keno Hill Mines. Prepared for Elsa Reclamation and Development Company, Whitehorse, YT, April 2011.
- Mirenda, R.J. 1986. Acute toxicity and accumulation of zinc in the crayfish, *Orconectes virilis* (Hagen). *Bulletin of Environmental Contamination and Toxicology* 37(3):387-394.
- Münzinger, A., and F. Monicelli. 1991. A comparison of the sensitivity of three *Daphnia magna* populations under chronic heavy metal stress. *Ecotoxicology and Environmental Safety* 22:24-31.
- Muyssen, B.T.A., and C.R. Janssen. 2001. Zinc acclimation and its effect on the zinc tolerance of *Raphidocelis subcapitata* and *Chlorella vulgaris* in laboratory experiments. *Chemosphere* 45:507-514.
- Muyssen, B.T.A., and C.R. Janssen. 2002. Tolerance and acclimation to zinc of *Ceriodaphnia dubia*. *Environmental Pollution* 117:301-306.
- Nebeker, A.V., A. Stinchfield, C. Savonen, and G.A. Chapman. 1986. Effects of copper, nickel and zinc on three species of Oregon freshwater snails. *Environmental Toxicology and Chemistry* 5:807-811.

- Nebeker, A.V., G.S. Schuytema, and S.L. Ott. 1995. Effects of cadmium on growth and bioaccumulation in the Northwestern salamander *Ambystoma gracile*. Archives of Environmental Contamination and Toxicology 29:492-499.
- Nehring, R. B., and J.P. Goettl. 1974. Acute toxicity of a zinc-polluted stream to four species of salmonids. Bulletin of Environmental Contamination and Toxicology 12(4): 464-469.
- Norberg, T.J. and D.I. Mount. 1985. A new fathead minnow (*Pimephales promelas*) subchronic toxicity test. Environmental Toxicology and Chemistry 4:711-718.
- Norberg-King, T.J. 1989. An evaluation of the fathead minnow seven-day subchronic test for estimating chronic toxicity. Environmental Toxicology and Chemistry 8:1075-1089.
- Pardue, W.J., and T.S. Wood. 1980. Baseline toxicity data for freshwater bryozoa exposed to copper, cadmium, chromium, and zinc. Journal of the Tennessee Academy of Science 55 (1): 27-31.
- Pardos, M., C. Benninghoff, and R.L. Thomas. 1998. Photosynthetic and population growth response of the test alga *Selenastrum capricornutum* Printz to zinc, cadmium and suspended sediment elutriates. Journal of Applied Phycology 10:145-151.
- Pascoe, D., K.A. Williams, and D.W.J. Green. 1989. Chronic toxicity of cadmium to *Chironomus riparius* Meigen - Effects upon larval development and adult emergence. Hydrobiologia 175:109-115.
- Paulauskis, J.D. and R.W. Winner. 1988. Effects of water hardness and humic acid on zinc toxicity to *Daphnia magna* Straus. Aquatic Toxicology 12:273-290.
- Pentz, S.B. and Kostaschuk, R.A. 1999. Effect of placer mining on suspended sediments in reaches of sensitive fish habitat. Environmental Geology, 37:78-89.
- Pestana, J.L.T., A. Ré, A.J.A. Nogueira, and A.M.V.M. Soares. 2007. Effects of cadmium and zinc on the feeding behaviour of two freshwater crustaceans: *Atyaephyra desmarestii* (Decapoda) and *Echinogammarus meridionalis* (Amphipoda). Chemosphere 68:1556-1562.

- Pickering, Q.H., and W.N. Vigor. 1965. The acute toxicity of zinc to eggs and fry of the fathead minnow. *The Progressive Fish-Culturist* 27:153-157.
- Pickering, Q.H. 1968. Some effects of dissolved oxygen concentration upon the toxicity of zinc to the bluegill, *Lepomis macrochirus*, Raf. *Water Research* 2:187-194.
- Rombough, P.J. and E.T. Garside. 1982. Cadmium toxicity and accumulation in eggs and alevins of Atlantic salmon *Salmo salar*. *Canadian Journal of Zoology* 60:2006-2014.
- Sinley, J.R., J.P. Goettl, and P.H. Davies. 1974. The effect of zinc on rainbow trout (*Salmo gairdneri*) in hard and soft water. *Bulletin of Environmental Contamination and Toxicology* 12(2):193-201.
- Spehar, R.L., and A.R. Carlson. 1984. Derivation of site-specific water quality criteria for cadmium and the St. Louis River basin, Duluth, Minnesota. *Environmental Toxicology and Chemistry* 3:651-665.
- SPSS Inc. 2003. SPSS Version 12. 233 S. Wacker Drive, 11th floor Chicago, Illinois, U.S.A.
- Stanley, J.K., B.W. Brooks, and T.W. La Point. 2005. A comparison of chronic cadmium effects on *Hyalella azteca* in effluent-dominated stream mesocosms to similar laboratory exposures in effluent and reconstituted hard water. *Environmental Toxicology and Chemistry* 24:902-908.
- Starodub, M.E., P.T.S Wong, and C.I. Mayfield. 1987. Short-term and long-term studies on individual and combined toxicities of copper, zinc and lead to *Scenedesmus quadricauda*. *The Science of the Total Environment* 63:101-110.
- Suedel, B.C., J.H. Rodgers, and E. Deaver. 1997. Experimental factors that may affect toxicity of cadmium to freshwater organisms. *Archives of Environmental Contamination and Toxicology* 33:188-193.
- Timmermans, K.R., W. Peeters, and M. Tonkes. 1992. Cadmium, zinc, lead and copper in *Chironomus riparius* (Meigen) larvae (Diptera, Chironomidae): uptake and effects. *Hydrobiologia* 241(2):119-134.

- Williams, N.D. and D.A. Holdway. 2000. The effects of pulse-exposed cadmium and zinc on embryo hatchability, larval development, and survival of Australian crimson spotted rainbow fish (*Melanotaenia fluviatilis*). *Environ. Toxicol.* 15(3):165-173.
- Winner, R.W. 1981. A comparison of body length, brood size and longevity as indices of chronic copper and zinc stresses in *Daphnia magna*. *Environmental Pollution Series A, Ecological and Biological* 26:33-37.
- Winner, R.W. 1988. Evaluation of the relative sensitivities of 7-d *Daphnia magna* and *Ceriodaphnia dubia* toxicity tests for cadmium and sodium pentachlorophenate. *Environmental Toxicology and Chemistry* 7:153-159.
- Woodling, J., S. Brinkman, and S. Albeke. 2002. Acute and chronic toxicity of zinc to the mottled sculpin *Cottus bairdi*. *Environmental Toxicology and Chemistry* 21(9):1922-1926.

APPENDIX A

Raw Water Quality Data From Selected UKHM Stations 2006 to 2009

Table A.2: Statistical comparison of cadmium and zinc concentrations over time between KV-1, KV-4 and KV-5.

a) Linear regression of concentration versus time

| Substance | Station | P value | r ² |
|-----------|---------|---------|----------------|
| Cadmium | KV-1 | 0.000 | 0.492 |
| | KV-4 | 0.006 | 0.425 |
| | KV-5 | 0.000 | 0.592 |
| Zinc | KV-1 | 0.000 | 0.434 |
| | KV-4 | 0.018 | 0.340 |
| | KV-5 | 0.002 | 0.490 |

b) Comparison of slopes of cadmium and zinc concentrations over time

| Substance | Comparison | Significantly Different | P value |
|-----------|--------------|-------------------------|---------|
| Cadmium | KV-1 vs KV-4 | N | 0.381 |
| | KV-1 vs KV-5 | N | 0.966 |
| | | | |
| Zinc | KV-1 vs KV-4 | N | 0.415 |
| | KV-1 vs KV-5 | N | 0.986 |

c) ANCOVA comparisons of means adjusted for date

| Substance | Comparison | Log Adjusted Mean | Significantly Different | Difference Relative to KV-1 | P value |
|-----------|--------------|-------------------|-------------------------|-----------------------------|---------|
| Cadmium | KV-1 vs KV-4 | KV-1 -3.244 | N | - | 0.208 |
| | | KV-4 -3.327 | | | |
| | KV-1 vs KV-5 | KV-1 -3.260 | Y | ↓ | 0.001 |
| | | KV-5 -3.480 | | | |
| Zinc | KV-1 vs KV-4 | KV-1 -0.954 | Y | ↓ | 0.000 |
| | | KV-4 -1.170 | | | |
| | KV-1 vs KV-5 | KV-1 -0.965 | Y | ↓ | 0.000 |
| | | KV-5 -1.261 | | | |

APPENDIX B

Cadmium Toxicity Data

Table B.1: Summary of long-term effects of cadmium on a variety of aquatic organisms

| Species Common Name | Scientific Name | Life stage | Duration | Endpoint | Observed Effect | Test Hardness (mg/L) | Effect Concentration (ug/L) | Effect Concentration (ug/L) at 100 mg/L Hardness ¹ | pH | Authors | Year |
|---------------------|-----------------------------|--------------------|----------|----------|--------------------------------------------|----------------------|-----------------------------|---------------------------------------------------------------|---------------|----------------------|------|
| Amphipod - gammarid | Echinogammarus meridionalis | Adult | 6 d | LOEC/L | Feeding inhibition | 263.4 | 6.35 | 2.4 | 7.92 (+-0.02) | Pestana et al. | 2007 |
| Amphipod - gammarid | Echinogammarus meridionalis | Adult | 6 d | NOEC/L | Feeding inhibition | 263.4 | 4.2 | 1.6 | 7.92 (+-0.02) | Pestana et al. | 2007 |
| Amphipod - gammarid | Gammarus pulex | Adult | 5 d | LOEC/L | Behaviour - Inhibition of swimming ability | 269.2 | 7.5 | 2.7 | 7.19 +- 0.02 | Felten et al. | 2007 |
| Amphipod - gammarid | Gammarus pulex | Adult | 7 d | LOEC/L | Behaviour - Inhibition of swimming ability | 269.2 | 7.5 | 2.7 | 7.19 +- 0.02 | Felten et al. | 2007 |
| Amphipod - gammarid | Gammarus pulex | Adult | 7 d | LOEC/L | Feeding inhibition | 269.2 | 15 | 5.4 | 7.19 +- 0.02 | Felten et al. | 2007 |
| Amphipod - gammarid | Gammarus pulex | Adult | 5 d | LOEC/L | Mortality | 269.2 | 7.5 | 2.7 | 7.19 +- 0.02 | Felten et al. | 2007 |
| Amphipod - gammarid | Gammarus pulex | Adult | 7 d | LOEC/L | Mortality | 269.2 | 15 | 5.4 | 7.19 +- 0.02 | Felten et al. | 2007 |
| Amphipod - gammarid | Gammarus pulex | Adult | 5 d | LOEC/L | Respiration | 269.2 | 15 | 5.4 | 7.19 +- 0.02 | Felten et al. | 2007 |
| Amphipod - gammarid | Gammarus pulex | Adult | 7 d | LOEC/L | Respiration | 269.2 | 7.5 | 2.7 | 7.19 +- 0.02 | Felten et al. | 2007 |
| Amphipod - gammarid | Gammarus pulex | Adult | 7 d | MATC | Feeding inhibition | 269.2 | 10.6 | 3.8 | 7.19 +- 0.02 | Felten et al. | 2007 |
| Amphipod - gammarid | Gammarus pulex | Adult | 7 d | MATC | Mortality | 269.2 | 10.6 | 3.8 | 7.19 +- 0.02 | Felten et al. | 2007 |
| Amphipod - gammarid | Gammarus pulex | Adult | 5 d | MATC | Respiration | 269.2 | 10.6 | 3.8 | 7.19 +- 0.02 | Felten et al. | 2007 |
| Amphipod - gammarid | Gammarus pulex | Adult | 7 d | NOEC/L | Feeding inhibition | 269.2 | 7.5 | 2.7 | 7.19 +- 0.02 | Felten et al. | 2007 |
| Amphipod - gammarid | Gammarus pulex | Adult | 7 d | NOEC/L | Mortality | 269.2 | 7.5 | 2.7 | 7.19 +- 0.02 | Felten et al. | 2007 |
| Amphipod - gammarid | Gammarus pulex | Adult | 5 d | NOEC/L | Respiration | 269.2 | 7.5 | 2.7 | 7.19 +- 0.02 | Felten et al. | 2007 |
| Amphipod - scud | Hyalella azteca | 7-8 d old | 28 d | IC25 | Biomass, decrease in | 280 | 0.51 | 0.2 | 7.80 | Ingersoll and Kemble | 2001 |
| Amphipod - scud | Hyalella azteca | 7-8 d old | 28 d | IC25 | Length | 280 | 2.6 | 0.9 | 7.80 | Ingersoll and Kemble | 2001 |
| Amphipod - scud | Hyalella azteca | 7-8 d old | 42 d | IC25 | Mortality | 280 | 1.9 | 0.7 | 7.80 | Ingersoll and Kemble | 2001 |
| Amphipod - scud | Hyalella azteca | 7-8 d old | 42 d | IC25 | Reproduction | 280 | 1.4 | 0.5 | 7.80 | Ingersoll and Kemble | 2001 |
| Amphipod - scud | Hyalella azteca | 7-8 d old | 28 d | IC25 | Weight | 280 | 0.74 | 0.3 | 7.80 | Ingersoll and Kemble | 2001 |
| Amphipod - scud | Hyalella azteca | 7-8 d old | 28 d | LOEC/L | Mortality | 139.6 | 22.97 | 16.3 | 7.0 (0.3) | Stanley et al. | 2005 |
| Amphipod - scud | Hyalella azteca | 7-8 d old | 42 d | LOEC/L | Mortality | 139.6 | 22.97 | 16.3 | 7.0 (0.3) | Stanley et al. | 2005 |
| Amphipod - scud | Hyalella azteca | 7-8 d old | 28 d | LOEC/L | Mortality | 162.7 | 5.09 | 3.1 | 7.9 (0.1) | Stanley et al. | 2005 |
| Amphipod - scud | Hyalella azteca | 7-8 d old | 28 d | MATC | Mortality | 139.6 | 12.52 | 8.9 | 7.0 (0.3) | Stanley et al. | 2005 |
| Amphipod - scud | Hyalella azteca | 7-8 d old | 42 d | MATC | Mortality | 139.6 | 12.52 | 8.9 | 7.0 (0.3) | Stanley et al. | 2005 |
| Amphipod - scud | Hyalella azteca | 7-8 d old | 28 d | MATC | Mortality | 162.7 | 3.56 | 2.2 | 7.9 (0.1) | Stanley et al. | 2005 |
| Amphipod - scud | Hyalella azteca | 7-8 d old | 28 d | NOEC/L | Mortality | 139.6 | 6.82 | 4.8 | 7.0 (0.3) | Stanley et al. | 2005 |
| Amphipod - scud | Hyalella azteca | 7-8 d old | 42 d | NOEC/L | Mortality | 139.6 | 6.82 | 4.8 | 7.0 (0.3) | Stanley et al. | 2005 |
| Amphipod - scud | Hyalella azteca | 7-8 d old | 28 d | NOEC/L | Mortality | 162.7 | 2.49 | 1.5 | 7.9 (0.1) | Stanley et al. | 2005 |
| Amphipod - scud | Hyalella azteca | Juvenile | 14 d | MATC | Mortality | 17.0 | 0.16 | 1.0 | 5.5-7.7 | Suedel et al | 1997 |
| Amphipod - scud | Hyalella azteca | Juvenile | 7 d | MATC | Mortality | 17.0 | 1.4 | 8.6 | 5.5-7.7 | Suedel et al | 1997 |
| Amphipod - scud | Hyalella azteca | Juvenile | 10 d | MATC | Mortality | 17.0 | 1.4 | 8.6 | 5.5-7.7 | Suedel et al | 1997 |
| Amphipod - scud | Hyalella azteca | Juvenile | 14 d | NOEC/L | Growth | 17.0 | 2 | 12.3 | 5.5-7.7 | Suedel et al | 1997 |
| Atlantic salmon | Salmo salar | Egg | 470 d | LOEC/L | Biomass, decrease in | 28 | 2.5 | 9.2 | 7.3 (6.8-7.5) | Rombough and Garside | 1982 |
| Atlantic salmon | Salmo salar | Egg | 470 d | LOEC/L | Weight | 28 | 2.5 | 9.2 | 7.3 (6.8-7.5) | Rombough and Garside | 1982 |
| Atlantic salmon | Salmo salar | Egg | 496 d | LOEC/L | Weight and Length | 28 | 0.47 | 1.7 | 7.3 (6.8-7.5) | Rombough and Garside | 1982 |
| Atlantic salmon | Salmo salar | Egg | 402 d | MATC | Biomass, decrease in | 19 | 5.5 | 30.1 | 6.5 (6.3-6.8) | Rombough and Garside | 1982 |
| Atlantic salmon | Salmo salar | Egg | 496 d | MATC | Biomass, decrease in | 28 | 0.61 | 2.2 | 7.3 (6.8-7.5) | Rombough and Garside | 1982 |
| Atlantic salmon | Salmo salar | Early gastrulation | 78 d | MATC | Hatching success | 19 | 88 | 481 | 6.5 (6.3-6.8) | Rombough and Garside | 1982 |
| Atlantic salmon | Salmo salar | Eyed egg stage | 45 d | MATC | Hatching success | 19 | 156 | 853 | 6.5 (6.3-6.8) | Rombough and Garside | 1982 |
| Atlantic salmon | Salmo salar | Egg | 96 d | MATC | Hatching success | 19 | 156 | 853 | 6.5 (6.3-6.8) | Rombough and Garside | 1982 |
| Atlantic salmon | Salmo salar | Egg | 45 d | MATC | Hatching success | 28 | 490 | 1802 | 7.3 (6.8-7.5) | Rombough and Garside | 1982 |
| Atlantic salmon | Salmo salar | Egg | 48 d | MATC | Hatching success | 28 | 490 | 1802 | 7.3 (6.8-7.5) | Rombough and Garside | 1982 |
| Atlantic salmon | Salmo salar | Egg | 158 d | MATC | Mortality after hatch | 19 | 156 | 853 | 6.5 (6.3-6.8) | Rombough and Garside | 1982 |
| Atlantic salmon | Salmo salar | Egg | 92 d | MATC | Mortality after hatch | 28 | 4.5 | 16.5 | 7.3 (6.8-7.5) | Rombough and Garside | 1982 |
| Atlantic salmon | Salmo salar | Egg | 92 d | MATC | Mortality after hatch | 28 | 490 | 1802 | 7.3 (6.8-7.5) | Rombough and Garside | 1982 |
| Atlantic salmon | Salmo salar | Egg | 402 d | MATC | Weight | 19 | 5.5 | 30.1 | 6.5 (6.3-6.8) | Rombough and Garside | 1982 |

Table B.1: Summary of long-term effects of cadmium on a variety of aquatic organisms

| Species Common Name | Scientific Name | Life stage | Duration | Endpoint | Observed Effect | Test Hardness (mg/L) | Effect Concentration (ug/L) | Effect Concentration (ug/L) at 100 mg/L Hardness ¹ | pH | Authors | Year |
|---------------------|--------------------------|-------------|----------|----------|----------------------|----------------------|-----------------------------|---------------------------------------------------------------|------------------|---------------------|------|
| Brook Trout | Salvelinus fontinalis | Larva | 126 d | LOEC/L | Biomass, decrease in | 45 | 3.8 | 8.6 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Brook Trout | Salvelinus fontinalis | Larva | 126 d | MATC | Biomass, decrease in | 45 | 2 | 4.5 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Brook Trout | Salvelinus fontinalis | Larva | 126 d | NOEC/L | Biomass, decrease in | 45 | 1.1 | 2.5 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Brown trout | Salmo trutta | Swim-up fry | 30 d | IC20 | Biomass, decrease in | 29.2 | 0.87 | 3.1 | 7.54 (0.13) | Brinkman and Hansen | 2007 |
| Brown trout | Salmo trutta | Egg | 55 d | IC20 | Biomass, decrease in | 30.6 | 2.22 | 7.5 | 7.72 (0.12) | Brinkman and Hansen | 2007 |
| Brown trout | Salmo trutta | Swim-up fry | 30 d | IC20 | Biomass, decrease in | 67.6 | 2.18 | 3.3 | 7.60 (0.10) | Brinkman and Hansen | 2007 |
| Brown trout | Salmo trutta | Egg | 55 d | IC20 | Biomass, decrease in | 71.3 | 4.71 | 6.7 | 7.75 (0.14) | Brinkman and Hansen | 2007 |
| Brown trout | Salmo trutta | Swim-up fry | 30 d | IC20 | Biomass, decrease in | 151.0 | 6.62 | 4.3 | 7.51 (0.12) | Brinkman and Hansen | 2007 |
| Brown trout | Salmo trutta | Egg | 55 d | IC20 | Mortality | 149.0 | 13.6 | 9.0 | 7.83 (0.14) | Brinkman and Hansen | 2007 |
| Brown trout | Salmo trutta | Swim-up fry | 30 d | LOEC/L | Mortality | 29.2 | 1.4 | 4.9 | 7.54 (0.13) | Brinkman and Hansen | 2007 |
| Brown trout | Salmo trutta | Egg | 55 d | LOEC/L | Mortality | 30.6 | 4.87 | 16.4 | 7.72 (0.12) | Brinkman and Hansen | 2007 |
| Brown trout | Salmo trutta | Swim-up fry | 30 d | LOEC/L | Mortality | 67.6 | 2.58 | 3.9 | 7.60 (0.10) | Brinkman and Hansen | 2007 |
| Brown trout | Salmo trutta | Egg | 55 d | LOEC/L | Mortality | 71.3 | 8.64 | 12.2 | 7.75 (0.14) | Brinkman and Hansen | 2007 |
| Brown trout | Salmo trutta | Egg | 55 d | LOEC/L | Mortality | 149.0 | 19.1 | 12.7 | 7.83 (0.14) | Brinkman and Hansen | 2007 |
| Brown trout | Salmo trutta | Swim-up fry | 30 d | LOEC/L | Mortality | 151.0 | 8.88 | 5.8 | 7.51 (0.12) | Brinkman and Hansen | 2007 |
| Brown trout | Salmo trutta | Swim-up fry | 30 d | LOEC/L | Weight | 29.2 | 2.72 | 9.6 | 7.54 (0.13) | Brinkman and Hansen | 2007 |
| Brown trout | Salmo trutta | Swim-up fry | 30 d | LOEC/L | Weight | 67.6 | 4.49 | 6.7 | 7.60 (0.10) | Brinkman and Hansen | 2007 |
| Brown trout | Salmo trutta | Swim-up fry | 30 d | NOEC/L | Mortality | 29.2 | 0.74 | 2.6 | 7.54 (0.13) | Brinkman and Hansen | 2007 |
| Brown trout | Salmo trutta | Egg | 55 d | NOEC/L | Mortality | 30.6 | 2.54 | 8.5 | 7.72 (0.12) | Brinkman and Hansen | 2007 |
| Brown trout | Salmo trutta | Swim-up fry | 30 d | NOEC/L | Mortality | 67.6 | 1.3 | 1.9 | 7.60 (0.10) | Brinkman and Hansen | 2007 |
| Brown trout | Salmo trutta | Egg | 55 d | NOEC/L | Mortality | 71.3 | 4.68 | 6.6 | 7.75 (0.14) | Brinkman and Hansen | 2007 |
| Brown trout | Salmo trutta | Egg | 55 d | NOEC/L | Mortality | 149.0 | 9.62 | 6.4 | 7.83 (0.14) | Brinkman and Hansen | 2007 |
| Brown trout | Salmo trutta | Swim-up fry | 30 d | NOEC/L | Mortality | 151.0 | 4.81 | 3.2 | 7.51 (0.12) | Brinkman and Hansen | 2007 |
| Brown trout | Salmo trutta | Swim-up fry | 30 d | NOEC/L | Weight | 29.2 | 1.4 | 4.9 | 7.54 (0.13) | Brinkman and Hansen | 2007 |
| Brown trout | Salmo trutta | Swim-up fry | 30 d | NOEC/L | Weight | 67.6 | 2.58 | 3.9 | 7.60 (0.10) | Brinkman and Hansen | 2007 |
| Brown trout | Salmo trutta | Embryo | 83 d | LOEC/L | Biomass, decrease in | 45 | 11.7 | 26.5 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Brown trout | Salmo trutta | Embryo | 31 d | LOEC/L | Biomass, decrease in | 45 | 11.2 | 25.4 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Brown trout | Salmo trutta | Larva | 60 d | LOEC/L | Biomass, decrease in | 45 | 11.7 | 26.5 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Brown trout | Salmo trutta | Larva | 61 d | LOEC/L | Biomass, decrease in | 45 | 3.7 | 8.4 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Brown trout | Salmo trutta | Larva | 61 d | MATC | Biomass, decrease in | 45 | 2 | 4.5 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Brown trout | Salmo trutta | Embryo | 31 d | MATC | Biomass, decrease in | 45 | 6.4 | 14.5 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Brown trout | Salmo trutta | Larva | 60 d | MATC | Biomass, decrease in | 45 | 6.7 | 15.2 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Brown trout | Salmo trutta | Embryo | 83 d | MATC | Biomass, decrease in | 45 | 6.7 | 15.2 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Brown trout | Salmo trutta | Larva | 61 d | NOEC/L | Biomass, decrease in | 45 | 1.1 | 2.5 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Brown trout | Salmo trutta | Embryo | 31 d | NOEC/L | Biomass, decrease in | 45 | 3.7 | 8.4 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Brown trout | Salmo trutta | Larva | 60 d | NOEC/L | Biomass, decrease in | 45 | 3.8 | 8.6 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Brown trout | Salmo trutta | Embryo | 83 d | NOEC/L | Biomass, decrease in | 45 | 3.8 | 8.6 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Bull trout | Salvelinus confluentus | Juvenile | 55 d | LOEC/L | Growth | 30.6 | 0.786 | 2.6 | 7.55 (SD = 0.12) | Hansen et al. | 2002 |
| Bull trout | Salvelinus confluentus | Juvenile | 55 d | LOEC/L | Mortality | 30.6 | 0.786 | 2.6 | 7.55 (SD = 0.12) | Hansen et al. | 2002 |
| Bull trout | Salvelinus confluentus | Juvenile | 55 d | MATC | Growth | 30.6 | 0.549 | 1.8 | 7.55 (SD = 0.12) | Hansen et al. | 2002 |
| Bull trout | Salvelinus confluentus | Juvenile | 55 d | MATC | Mortality | 30.6 | 0.549 | 1.8 | 7.55 (SD = 0.12) | Hansen et al. | 2002 |
| Bull trout | Salvelinus confluentus | Juvenile | 55 d | NOEC/L | Growth | 30.6 | 0.383 | 1.3 | 7.55 (SD = 0.12) | Hansen et al. | 2002 |
| Bull trout | Salvelinus confluentus | Juvenile | 55 d | NOEC/L | Mortality | 30.6 | 0.383 | 1.3 | 7.55 (SD = 0.12) | Hansen et al. | 2002 |
| Chinook salmon | Oncorhynchus tshawytscha | Swim-up fry | 8 d | LC10 | Mortality | 23 | 1.2 | 5.4 | 7.1-7.5 | Chapman | 1978 |
| Chinook salmon | Oncorhynchus tshawytscha | Alevin | 8 d | LC10 | Mortality | 23 | >6 | >27 | 7.1-7.5 | Chapman | 1978 |

Table B.1: Summary of long-term effects of cadmium on a variety of aquatic organisms

| Species Common Name | Scientific Name | Life stage | Duration | Endpoint | Observed Effect | Test Hardness (mg/L) | Effect Concentration (ug/L) | Effect Concentration (ug/L) at 100 mg/L Hardness ¹ | pH | Authors | Year |
|---------------------|---------------------------------|-----------------|----------|-----------|------------------------------------------|----------------------|-----------------------------|---------------------------------------------------------------|---------------|---------------------|------|
| Cladocerans | Ceriodaphnia reticulata | Less than 24hrs | 7 d | MATC | Reproduction - Number of young per adult | 240 | 0.43 | 0.2 | 8.0 +- 0.3 | Elnabarawy et al | 1986 |
| Cladocerans | Ceriodaphnia reticulata | Less than 24hrs | 9 d | LOEC/L | Mortality | 67.0 | 15.2 | 22.9 | 7.2-7.8 | Spehar and Carlson | 1984 |
| Cladocerans | Ceriodaphnia reticulata | Less than 24hrs | 9 d | LOEC/L | Reproduction | 67.0 | 7.2 | 10.8 | 7.2-7.8 | Spehar and Carlson | 1984 |
| Cladocerans | Ceriodaphnia reticulata | Less than 24hrs | 9 d | MATC | Mortality | 67.0 | 10.5 | 15.8 | 7.2-7.8 | Spehar and Carlson | 1984 |
| Cladocerans | Ceriodaphnia reticulata | Less than 24hrs | 9 d | MATC | Reproduction | 67.0 | 4.9 | 7.4 | 7.2-7.8 | Spehar and Carlson | 1984 |
| Cladocerans | Ceriodaphnia reticulata | Less than 24hrs | 9 d | NOEC/L | Mortality | 67.0 | 7.2 | 10.8 | 7.2-7.8 | Spehar and Carlson | 1984 |
| Cladocerans | Ceriodaphnia reticulata | Less than 24hrs | 9 d | NOEC/L | Reproduction | 67.0 | 3.4 | 5.1 | 7.2-7.8 | Spehar and Carlson | 1984 |
| Coho salmon | Oncorhynchus kisutch | Embryo | 27 d | LOEC/L | Biomass, decrease in | 45 | 3.4 | 8 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Coho salmon | Oncorhynchus kisutch | Embryo | 47 d | LOEC/L | Biomass, decrease in | 45 | 12.5 | 28 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Coho salmon | Oncorhynchus kisutch | Larva | 62 d | LOEC/L | Biomass, decrease in | 45 | 12.5 | 28 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Coho salmon | Oncorhynchus kisutch | Embryo | 27 d | MATC | Biomass, decrease in | 45 | 2.1 | 5 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Coho salmon | Oncorhynchus kisutch | Embryo | 47 d | MATC | Biomass, decrease in | 45 | 7.2 | 16 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Coho salmon | Oncorhynchus kisutch | Larva | 62 d | MATC | Biomass, decrease in | 45 | 7.2 | 16 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Coho salmon | Oncorhynchus kisutch | Embryo | 27 d | NOEC/L | Biomass, decrease in | 45 | 1.3 | 3 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Coho salmon | Oncorhynchus kisutch | Embryo | 47 d | NOEC/L | Biomass, decrease in | 45 | 4.1 | 9 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Coho salmon | Oncorhynchus kisutch | Larva | 62 d | NOEC/L | Biomass, decrease in | 45 | 4.1 | 9 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Duckweed | Lemna minor | Not reported | 7 d | EC50 | Growth rate | 166.0 | 214 | 127 | 5.5 +- 0.2 | Drost et al. | 2007 |
| Duckweed | Lemna minor | Not reported | 6 d | EC50 | Growth rate | 166.0 | 214 | 127 | 5.5 +- 0.2 | Drost et al. | 2007 |
| Duckweed | Lemna minor | Not reported | 5 d | EC50 | Growth rate | 166.0 | 315 | 188 | 5.5 +- 0.2 | Drost et al. | 2007 |
| Duckweed | Lemna minor | Not reported | 3 d | EC50 | Growth rate | 166.0 | 393 | 234 | 5.5 +- 0.2 | Drost et al. | 2007 |
| Duckweed | Lemna minor | Not reported | 4 d | EC50 | Growth rate | 166.0 | 337 | 201 | 5.5 +- 0.2 | Drost et al. | 2007 |
| European shrimp | Atyaephyra desmarestii | Adult | 6 d | LOEC/L | Feeding inhibition | 263.4 | 6.53 | 2.4 | 7.92 (+0.02) | Pestana et al. | 2007 |
| European shrimp | Atyaephyra desmarestii | Adult | 6 d | NOEC/L | Feeding inhibition | 263.4 | 4.2 | 1.6 | 7.92 (+0.02) | Pestana et al. | 2007 |
| Fathead minnow | Pimephales promelas | Juvenile | 32 d | LOEC/L | Mortality | 67.0 | 26.7 | 40.2 | 7.2-7.8 | Spehar and Carlson | 1984 |
| Fathead minnow | Pimephales promelas | Juvenile | 32 d | MATC | Mortality | 67.0 | 18.9 | 28.5 | 7.2-7.8 | Spehar and Carlson | 1984 |
| Fathead minnow | Pimephales promelas | Juvenile | 32 d | NOEC/L | Mortality | 67.0 | 13.4 | 20.2 | 7.2-7.8 | Spehar and Carlson | 1984 |
| Fathead minnow | Pimephales promelas | Larva | 10 d | MATC | Mortality | 17.0 | 1.4 | 8.6 | 5.5-7.7 | Suedel et al | 1997 |
| Fathead minnow | Pimephales promelas | Larva | 7 d | MATC | Mortality | 17.0 | 4.9 | 30.0 | 5.5-7.7 | Suedel et al | 1997 |
| Fathead minnow | Pimephales promelas | Larva | 14 d | MATC | Mortality | 17.0 | 2.4 | 14.7 | 5.5-7.7 | Suedel et al | 1997 |
| Fathead minnow | Pimephales promelas | Larva | 10 d | NOEC/L | Growth | 17.0 | 2 | 12.3 | 5.5-7.7 | Suedel et al | 1997 |
| Fathead minnow | Pimephales promelas | Larva | 14 d | NOEC/L | Growth | 17.0 | 3 | 18.4 | 5.5-7.7 | Suedel et al | 1997 |
| Great pond snail | Lymnaea stagnalis | Adult | 4 weeks | EC50 | Growth | 284 | 142.2 | 48.9 | 6.65-8.14 | Coeurdassier et al. | 2003 |
| Great pond snail | Lymnaea stagnalis | Adult | 4 weeks | NOEC/L | Growth | 284 | 80 | 27.5 | 6.65-8.14 | Coeurdassier et al. | 2003 |
| Green alga | Ankistrodesmus falcatus | Population | 96 h | NOEC/L | Growth | 118.0 | 10 | 8 | 7.7 (7.2-8.2) | Baer et al. | 1999 |
| Green alga | Pseudokirchneriella subcapitata | Population | 96 h | NOEC/L | Growth | 118.0 | 5 | 4 | 7.7 (7.2-8.2) | Baer et al. | 1999 |
| Green alga | Pseudokirchneriella subcapitata | Population | 72 h | EC50 | Growth | 250 | 43.5 | 17 | 8.1 | Benhra et al. | 1997 |
| Green alga | Pseudokirchneriella subcapitata | Population | 72 h | EC10 | Growth rate | 3.42 | 2.8 | 88 | 6.71 | Kallqvist | 2007 |
| Green alga | Pseudokirchneriella subcapitata | Population | 72 h | EC10 | Growth rate | 6.21 | 7.5 | 129 | 6.85 | Kallqvist | 2007 |
| Green alga | Pseudokirchneriella subcapitata | Population | 72 h | EC10 | Growth rate | 16.21 | 8.5 | 55 | 6.74 | Kallqvist | 2007 |
| Green alga | Pseudokirchneriella subcapitata | Population | 72 h | EC10 | Growth rate | 46.21 | 6 | 13 | 6.65 | Kallqvist | 2007 |
| Green alga | Pseudokirchneriella subcapitata | Population | 72 h | Mean EC10 | Growth rate | | 5.7 | 53.6 | | Mean | |
| Green hydra | Hydra viridissima | | 7 d | NOEC/L | Population growth inhibition | 19.5 | 0.4 | 2.1 | 7.25-7.53 | Holdway et al. | 2001 |
| Lake Trout | Salvelinus namaycush | Embryo | 41 d | LOEC/L | Biomass, decrease in | 45 | 12.3 | 27.8 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Lake Trout | Salvelinus namaycush | Larva | 64 d | LOEC/L | Biomass, decrease in | 45 | 12.3 | 27.8 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Lake Trout | Salvelinus namaycush | Embryo | 41 d | MATC | Biomass, decrease in | 45 | 7.4 | 16.7 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Lake Trout | Salvelinus namaycush | Larva | 64 d | MATC | Biomass, decrease in | 45 | 7.4 | 16.7 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Lake Trout | Salvelinus namaycush | Embryo | 41 d | NOEC/L | Biomass, decrease in | 45 | 4.4 | 10.0 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Lake Trout | Salvelinus namaycush | Larva | 64 d | NOEC/L | Biomass, decrease in | 45 | 4.4 | 10.0 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |

Table B.1: Summary of long-term effects of cadmium on a variety of aquatic organisms

| Species Common Name | Scientific Name | Life stage | Duration | Endpoint | Observed Effect | Test Hardness (mg/L) | Effect Concentration (ug/L) | Effect Concentration (ug/L) at 100 mg/L Hardness ¹ | pH | Authors | Year |
|-------------------------|---------------------|------------------|----------|----------|---------------------------------------|----------------------|-----------------------------|---------------------------------------------------------------|------------------|----------------------|------|
| Marsh snail | Lymnaea palustris | Adult | 4 weeks | EC50 | Growth | 284 | 58.2 | 20.0 | 6.65-8.14 | Coeurdassier et al. | 2003 |
| Marsh snail | Lymnaea palustris | Adult | 4 weeks | EC50 | Repro - No. egg masses per individual | 284 | 60.9 | 20.9 | 6.65-8.14 | Coeurdassier et al. | 2003 |
| Marsh snail | Lymnaea palustris | Adult | 4 weeks | EC50 | Repro - No. eggs per egg mass | 284 | 124 | 42.6 | 6.65-8.14 | Coeurdassier et al. | 2003 |
| Marsh snail | Lymnaea palustris | Adult | 4 weeks | EC50 | Repro - No. eggs per individual | 284 | 64.7 | 22.2 | 6.65-8.14 | Coeurdassier et al. | 2003 |
| Marsh snail | Lymnaea palustris | Adult | 4 weeks | NOEC/L | Growth | 284 | 40 | 13.8 | 6.65-8.14 | Coeurdassier et al. | 2003 |
| Marsh snail | Lymnaea palustris | Adult | 4 weeks | NOEC/L | Repro - No. egg masses per individual | 284 | 40 | 13.8 | 6.65-8.14 | Coeurdassier et al. | 2003 |
| Marsh snail | Lymnaea palustris | Adult | 4 weeks | NOEC/L | Repro - No. eggs per individual | 284 | 40 | 13.8 | 6.65-8.14 | Coeurdassier et al. | 2003 |
| Midge | Chironomus riparius | 1st instar | 17 d | LOEC/L | Mortality | 98.0 | 150 | 153.1 | 7.6 | Pascoe et al. | 1989 |
| Midge | Chironomus riparius | 1st instar | 17 d | MATC | Mortality | 98.0 | 47.4 | 48.4 | 7.6 | Pascoe et al. | 1989 |
| Midge | Chironomus riparius | 1st instar | 17 d | NOEC/L | Mortality | 98.0 | 15 | 15.3 | 7.6 | Pascoe et al. | 1989 |
| Midge | Chironomus tentans | Less than 24hrs | 20 d | IC25 | Biomass, decrease in | 280 | 10.3 | 3.6 | 7.80 | Ingersoll and Kemble | 2001 |
| Midge | Chironomus tentans | Less than 24hrs | 60 d | IC25 | Hatching success | 280 | 4 | 1.4 | 7.80 | Ingersoll and Kemble | 2001 |
| Midge | Chironomus tentans | Less than 24hrs | 20 d | IC25 | Mortality | 280 | 16.4 | >5.7 | 7.80 | Ingersoll and Kemble | 2001 |
| Midge | Chironomus tentans | Less than 24hrs | 60 d | IC25 | Percent emergence | 280 | 8.1 | 2.8 | 7.80 | Ingersoll and Kemble | 2001 |
| Midge | Chironomus tentans | Less than 24hrs | 60 d | IC25 | Repro - No. eggs per individual | 280 | >16.4 | >5.7 | 7.80 | Ingersoll and Kemble | 2001 |
| Midge | Chironomus tentans | Less than 24hrs | 20 d | IC25 | Weight | 280 | 9.9 | 3.5 | 7.80 | Ingersoll and Kemble | 2001 |
| Midge | Chironomus tentans | 2nd instar | 7 d | LOEC/L | Growth | 17.0 | 500 | 3063.5 | 5.5-7.7 | Suedel et al | 1997 |
| Midge | Chironomus tentans | 2nd instar | 10 d | LOEC/L | Growth | 17.0 | 500 | 3063.5 | 5.5-7.7 | Suedel et al | 1997 |
| Midge | Chironomus tentans | 2nd instar | 14 d | LOEC/L | Growth | 17.0 | 100 | 612.7 | 5.5-7.7 | Suedel et al | 1997 |
| Midge | Chironomus tentans | 2nd instar | 7 d | MATC | Mortality | 17.0 | 707 | 4331.8 | 5.5-7.7 | Suedel et al | 1997 |
| Midge | Chironomus tentans | 2nd instar | 10 d | MATC | Mortality | 17.0 | 707 | 4331.8 | 5.5-7.7 | Suedel et al | 1997 |
| Midge | Chironomus tentans | 2nd instar | 14 d | MATC | Mortality | 17.0 | 707 | 4331.8 | 5.5-7.7 | Suedel et al | 1997 |
| Mottled sculpin | Cottus bairdi | Swim-up fry | 28 d | EC50 | Biomass, decrease in | 102 | 2.4 | 2 | 8.21 | Besser et al. | 2007 |
| Mottled sculpin | Cottus bairdi | Swim-up fry | 21 d | EC50 | Biomass, decrease in | 104 | 1.77 | 2 | 8.23 | Besser et al. | 2007 |
| Northern pike | Esox lucius | Embryo | 35 d | LOEC/L | Biomass, decrease in | 45 | 12.9 | 29 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Northern pike | Esox lucius | Embryo | 35 d | MATC | Biomass, decrease in | 45 | 7.4 | 17 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Northern pike | Esox lucius | Embryo | 35 d | NOEC/L | Biomass, decrease in | 45 | 4.2 | 10 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| Northwestern salamander | Ambystoma gracile | Larva | 24 d | LOEC/L | Weight | 45 | 193.1 | 437 | 6.8 | Nebeker et al | 1995 |
| Northwestern salamander | Ambystoma gracile | Larva | 10 d | LOEC/L | Weight | 45 | 227.3 | 514 | 6.8 | Nebeker et al | 1995 |
| Northwestern salamander | Ambystoma gracile | Larva | 24 d | MATC | Weight | 45 | 97.2 | 220 | 6.8 | Nebeker et al | 1995 |
| Northwestern salamander | Ambystoma gracile | Larva | 10 d | MATC | Weight | 45 | 155.4 | 352 | 6.8 | Nebeker et al | 1995 |
| Northwestern salamander | Ambystoma gracile | Larva | 24 d | NOEC/L | Weight | 45 | 48.9 | 111 | 6.8 | Nebeker et al | 1995 |
| Northwestern salamander | Ambystoma gracile | Larva | 10 d | NOEC/L | Weight | 45 | 106.3 | 241 | 6.8 | Nebeker et al | 1995 |
| Rainbow trout | Oncorhynchus mykiss | Adult | 65 wks | LOEC/L | Reproduction - delay in oogenesis | 250 | 1.77 | 0.7 | 7.4-8.0 | Brown et al | 1994 |
| Rainbow trout | Oncorhynchus mykiss | Adult | 65 wks | MATC | Reproduction - delay in oogenesis | 250 | 0.91 | 0.4 | 7.4-8.0 | Brown et al | 1994 |
| Rainbow trout | Oncorhynchus mykiss | Adult | 65 wks | NOEC/L | Reproduction - delay in oogenesis | 250 | 0.47 | 0.2 | 7.4-8.0 | Brown et al | 1994 |
| Rainbow trout | Oncorhynchus mykiss | Alevin | 8 d | LC10 | Mortality | 23 | >6 | >27 | 7.1-7.5 | Chapman | 1978 |
| Rainbow trout | Oncorhynchus mykiss | Swim-up fry | 8 d | LC10 | Mortality | 23 | 1 | 4.5 | 7.1-7.5 | Chapman | 1978 |
| Rainbow trout | Oncorhynchus mykiss | Early life stage | 62 d | EC10 | Length | 29.4 | >2.5 | >8.7 | 7.19 (SD = 0.30) | Mebane et al. | 2007 |
| Rainbow trout | Oncorhynchus mykiss | Early life stage | 53 d | EC10 | Mortality | 19.7 | 0.82 | 4.3 | 6.75 (5.0-7.7) | Mebane et al. | 2007 |
| Rainbow trout | Oncorhynchus mykiss | Early life stage | 62 d | EC10 | Mortality | 29.4 | 1.6 | 5.6 | 7.19 (SD = 0.30) | Mebane et al. | 2007 |
| Rainbow trout | Oncorhynchus mykiss | Early life stage | 62 d | EC10 | Weight | 29.4 | 0.15 | 0.5 | 7.19 (SD = 0.30) | Mebane et al. | 2007 |
| Rainbow trout | Oncorhynchus mykiss | Early life stage | 62 d | LOEC/L | Length | 29.4 | 0.16 | 0.6 | 7.19 (SD = 0.30) | Mebane et al. | 2007 |
| Rainbow trout | Oncorhynchus mykiss | Early life stage | 53 d | LOEC/L | Mortality | 19.7 | 1.3 | 6.9 | 6.75 (5.0-7.7) | Mebane et al. | 2007 |
| Rainbow trout | Oncorhynchus mykiss | Early life stage | 62 d | LOEC/L | Mortality | 29.4 | 2.5 | 8.7 | 7.19 (SD = 0.30) | Mebane et al. | 2007 |
| Rainbow trout | Oncorhynchus mykiss | Early life stage | 62 d | LOEC/L | Weight | 29.4 | 0.16 | 0.6 | 7.19 (SD = 0.30) | Mebane et al. | 2007 |
| Rainbow trout | Oncorhynchus mykiss | Early life stage | 53 d | MATC | Mortality | 19.7 | 0.88 | 4.6 | 6.75 (5.0-7.7) | Mebane et al. | 2007 |
| Rainbow trout | Oncorhynchus mykiss | Early life stage | 62 d | MATC | Mortality | 29.4 | 1.6 | 5.6 | 7.19 (SD = 0.30) | Mebane et al. | 2007 |
| Rainbow trout | Oncorhynchus mykiss | Early life stage | 53 d | NOEC/L | Mortality | 19.7 | 0.6 | 3.2 | 6.75 (5.0-7.7) | Mebane et al. | 2007 |
| Rainbow trout | Oncorhynchus mykiss | Early life stage | 62 d | NOEC/L | Mortality | 29.4 | 1 | 3.5 | 7.19 (SD = 0.30) | Mebane et al. | 2007 |

Table B.1: Summary of long-term effects of cadmium on a variety of aquatic organisms

| Species Common Name | Scientific Name | Life stage | Duration | Endpoint | Observed Effect | Test Hardness (mg/L) | Effect Concentration (ug/L) | Effect Concentration (ug/L) at 100 mg/L Hardness ¹ | pH | Authors | Year |
|---------------------|-----------------------|-----------------|----------|----------|------------------------------------------|----------------------|-----------------------------|---------------------------------------------------------------|----------------|---------------------------|------|
| Water flea | Ceriodaphnia dubia | Not reported | 14 d | MATC | Mortality | 17.0 | 11.4 | 69.8 | 5.5-7.7 | Suedel et al | 1997 |
| Water flea | Ceriodaphnia dubia | Not reported | 10 d | MATC | Mortality | 17.0 | 11.4 | 69.8 | 5.5-7.7 | Suedel et al | 1997 |
| Water flea | Ceriodaphnia dubia | Not reported | 7 d | MATC | Mortality | 17.0 | 11.4 | 69.8 | 5.5-7.7 | Suedel et al | 1997 |
| Water flea | Ceriodaphnia dubia | Not reported | 14 d | MATC | Reproduction | 17.0 | 2 | 12.3 | 5.5-7.7 | Suedel et al | 1997 |
| Water flea | Ceriodaphnia dubia | Not reported | 10 d | MATC | Reproduction | 17.0 | 2 | 12.3 | 5.5-7.7 | Suedel et al | 1997 |
| Water flea | Ceriodaphnia dubia | Not reported | 7 d | MATC | Reproduction | 17.0 | 2 | 12.3 | 5.5-7.7 | Suedel et al | 1997 |
| Water flea | Daphnia magna | Adult | 7 d | EC10 | Feeding inhibition | 179 | 0.13 | 0.1 | 8.07 +- 0.07 | Barata and Baird | 2000 |
| Water flea | Daphnia magna | Adult | 7 d | EC10 | Repro - brood mass | 179 | 0.13 | 0.1 | 8.07 +- 0.07 | Barata and Baird | 2000 |
| Water flea | Daphnia magna | Adult | 7 d | EC10 | Reproduction - Brood size | 179 | 0.14 | 0.1 | 8.07 +- 0.07 | Barata and Baird | 2000 |
| Water flea | Daphnia magna | Adult | 7 d | EC10 | Weight | 179 | 1.65 | 0.9 | 8.07 +- 0.07 | Barata and Baird | 2000 |
| Water flea | Daphnia magna | Adult | 7 d | LC10 | Mortality | 179 | 1.15 | 0.6 | 8.07 +- 0.07 | Barata and Baird | 2000 |
| Water flea | Daphnia magna | Less than 24hrs | 21 d | EC16 | Reproduction | 45.3 | 0.17 | 0.4 | 7.74 (7.4-8.2) | Biesinger and Christensen | 1972 |
| Water flea | Daphnia magna | Not reported | 21 d | LOEC/L | Reproduction - Number of young per adult | 130 | 1.86 | 1.4 | | Borgmann et al | 1989 |
| Water flea | Daphnia magna | Not reported | 21 d | MATC | Reproduction - Number of young per adult | 130 | 0.64 | 0.5 | | Borgmann et al | 1989 |
| Water flea | Daphnia magna | Not reported | 21 d | NOEC/L | Reproduction - Number of young per adult | 130 | 0.22 | 0.2 | | Borgmann et al | 1989 |
| Water flea | Daphnia magna | Less than 24hrs | 21 d | MATC | Repro - Number of young per survivor | 53 | 1.52 | 2.9 | 7.5 +- 0.2 | Chapman et al | 1980 |
| Water flea | Daphnia magna | Less than 24hrs | 21 d | MATC | Repro - Number of young per survivor | 103 | 0.21 | 0.2 | 7.9 +- 0.3 | Chapman et al | 1980 |
| Water flea | Daphnia magna | Less than 24hrs | 21 d | MATC | Reproduction - Number of young per adult | 53 | 0.15 | 0.3 | 7.5 +- 0.2 | Chapman et al | 1980 |
| Water flea | Daphnia magna | Less than 24hrs | 21 d | MATC | Reproduction - Number of young per adult | 103 | 0.38 | 0.4 | 7.9 +- 0.3 | Chapman et al | 1980 |
| Water flea | Daphnia magna | Less than 24hrs | 14 d | MATC | Reproduction - Number of young per adult | 240 | 4.3 | 1.8 | 8.0 +- 0.3 | Elnabarawy et al | 1986 |
| Water flea | Daphnia magna | 24h | 21 d | LOEC/L | Reproduction | 249.8 | 1.94 | 0.8 | 8.0 +- 0.2 | Kuhn et al | 1989 |
| Water flea | Daphnia magna | 24h | 21 d | MATC | Reproduction | 249.8 | 1.09 | 0.4 | 8.0 +- 0.2 | Kuhn et al | 1989 |
| Water flea | Daphnia magna | 24h | 21 d | NOEC/L | Reproduction | 249.8 | 0.6 | 0.2 | 8.0 +- 0.2 | Kuhn et al | 1989 |
| Water flea | Daphnia magna | Not reported | 7 d | MATC | Mortality | 78.0 | 7.1 | 9.2 | 6.9-8.3 | Suedel et al | 1997 |
| Water flea | Daphnia magna | Not reported | 10 d | MATC | Mortality | 78.0 | 7.1 | 9.2 | 6.9-8.3 | Suedel et al | 1997 |
| Water flea | Daphnia magna | Not reported | 14 d | MATC | Mortality | 78.0 | 7.1 | 9.2 | 6.9-8.3 | Suedel et al | 1997 |
| Water flea | Daphnia magna | Neonate | 7 d | MATC | Growth | 90 | 1.2 | 1.3 | | Winner | 1988 |
| Water flea | Daphnia pulex | Less than 24hrs | 14 d | MATC | Reproduction - Number of young per adult | 240 | 13.7 | 5.6 | 8.0 +- 0.3 | Elnabarawy et al | 1986 |
| White Sucker | Catostomus commersoni | Embryo | 40 h | LOEC/L | Biomass, decrease in | 45 | 12 | 27 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| White Sucker | Catostomus commersoni | Embryo | 40 h | MATC | Biomass, decrease in | 45 | 7.1 | 16 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| White Sucker | Catostomus commersoni | Embryo | 40 h | NOEC/L | Biomass, decrease in | 45 | 4.2 | 10 | 7.6 (7.2-7.8) | Eaton et al. | 1978 |

¹ If adjusted from another hardness, value was calculated using the following equation: EXP(LN(EFFECT conc)-(1.023)*(LN(measured water hardness)-LN(desired water hardness)))

most sensitive effect end-point for species

APPENDIX C

Zinc Toxicity Data

Table C.1: Summary of long-term effects of zinc on a variety of aquatic organisms

| Species Common Name | Scientific Name | Life Stage | Duration | Endpoint | Observed Effect | Effect Concentration (ug/L) | Test Hardness (mg/L) | Effect Concentration (ug/L) at 100 mg/L Hardness ¹ | pH | Authors | Year |
|---------------------|---------------------------------|-----------------|-----------|----------|----------------------------------------|-----------------------------|----------------------|---------------------------------------------------------------|---------------|---------------------|------|
| Amphipod | <i>Hyalella azteca</i> | 1 week | 10 weeks | LOEC | Mortality | 180 | 130 | 144 | 7.9-8.6 | Borgmann et al | 1993 |
| Amphipod | <i>Hyalella azteca</i> | 1 week | 10 weeks | MATC | Mortality | 134.16 | 130 | 107 | 7.9-8.6 | Borgmann et al | 1993 |
| Amphipod | <i>Hyalella azteca</i> | 1 week | 10 weeks | NOEC | Mortality | 100 | 130 | 80 | 7.9-8.6 | Borgmann et al | 1993 |
| Amphipod | <i>Hyalella azteca</i> | 1- 11 days | 7 d | LC50 | Mortality | 56 | 18 | 239 | 6.44-8.68 | Borgmann et al | 2005 |
| Amphipod | <i>Hyalella azteca</i> | 1- 11 days | 7 d | LC50 | Mortality | 70 | 18 | 299 | 6.44-8.68 | Borgmann et al | 2005 |
| Amphipod | <i>Hyalella azteca</i> | 1- 11 days | 7 d | LC50 | Mortality | 222 | 124 | 185 | 7.23-8.98 | Borgmann et al | 2005 |
| Atlantic salmon | <i>Salmo salar</i> | parr | 14 d | LC50 | Mortality | 3640 | 351 | 1258 | 7.6 | Hodson and Sprague | 1975 |
| Atlantic salmon | <i>Salmo salar</i> | parr | 14 d | LC50 | Mortality | 5046 | 351 | 1744 | 7.6 | Hodson and Sprague | 1975 |
| Atlantic salmon | <i>Salmo salar</i> | parr | 14 d | LC50 | Mortality | 5198 | 351 | 1797 | 7.6 | Hodson and Sprague | 1975 |
| Bluegill | <i>Lepomis macrochirus</i> | Not reported | 20 d | TLM | Mortality | 11300 | 370 | 3736 | 7.8 (7.7-8.0) | Pickering | 1968 |
| Brook Trout | <i>Salvelinus fontinalis</i> | Egg | 100 weeks | EC10 | Hatching success | 418 | 45.9 | 808 | 7.2-7.9 | Holcombe et al. | 1979 |
| Brook Trout | <i>Salvelinus fontinalis</i> | Egg | 24 w | IC10 | egg fragility | 200 | 45.9 | 386 | 7.2-7.9 | Holcombe et al. | 1979 |
| Brook Trout | <i>Salvelinus fontinalis</i> | Egg | 8 weeks | LC10 | Embryo (6 hours old) survival to hatch | 1114 | 45.9 | 2153 | 7.2-7.9 | Holcombe et al. | 1979 |
| Brook Trout | <i>Salvelinus fontinalis</i> | Embryo | 12 weeks | LC10 | Mortality | 1215 | 45.9 | 2348 | 7.2-7.9 | Holcombe et al. | 1979 |
| Brook Trout | <i>Salvelinus fontinalis</i> | Egg | 24 w | LOEC | egg fragility | 266 | 45.9 | 514 | 7.2-7.9 | Holcombe et al. | 1979 |
| Brook Trout | <i>Salvelinus fontinalis</i> | Egg | 100 weeks | LOEC | Hatching success | 534 | 45.9 | 1032 | 7.2-7.9 | Holcombe et al. | 1979 |
| Brook Trout | <i>Salvelinus fontinalis</i> | Embryo | 12 weeks | LOEC | Mortality | 1382 | 45.9 | 2671 | 7.2-7.9 | Holcombe et al. | 1979 |
| Brook Trout | <i>Salvelinus fontinalis</i> | Embryo | 12 weeks | LOEC | Mortality | 2099 | 45.9 | 4056 | 7.2-7.9 | Holcombe et al. | 1979 |
| Brook Trout | <i>Salvelinus fontinalis</i> | Embryo | 8 weeks | LOEC | Mortality | 1382 | 45.9 | 2671 | 7.2-7.9 | Holcombe et al. | 1979 |
| Brook Trout | <i>Salvelinus fontinalis</i> | Egg | 24 w | MATC | egg fragility | 174 | 45.9 | 336 | 7.2-7.9 | Holcombe et al. | 1979 |
| Brook Trout | <i>Salvelinus fontinalis</i> | Egg | 8 weeks | MATC | Embryo (6 hours old) survival to hatch | 990 | 45.9 | 1913 | 7.2-7.9 | Holcombe et al. | 1979 |
| Brook Trout | <i>Salvelinus fontinalis</i> | Egg | 100 weeks | MATC | Hatching success | 377 | 45.9 | 729 | 7.2-7.9 | Holcombe et al. | 1979 |
| Brook Trout | <i>Salvelinus fontinalis</i> | Embryo | 12 weeks | MATC | Mortality | 990 | 45.9 | 1913 | 7.2-7.9 | Holcombe et al. | 1979 |
| Brook Trout | <i>Salvelinus fontinalis</i> | Embryo | 8 weeks | MATC | Mortality | 1000 | 45.9 | 1932 | 7.2-7.9 | Holcombe et al. | 1979 |
| Brook Trout | <i>Salvelinus fontinalis</i> | Embryo | 8 weeks | MATC | Mortality | 1685 | 45.9 | 3256 | 7.2-7.9 | Holcombe et al. | 1979 |
| Brook Trout | <i>Salvelinus fontinalis</i> | Egg | 24 w | NOEC | egg fragility | 114 | 45.9 | 220 | 7.2-7.9 | Holcombe et al. | 1979 |
| Brook Trout | <i>Salvelinus fontinalis</i> | Egg | 100 weeks | NOEC | Hatching success | 266 | 45.9 | 514 | 7.2-7.9 | Holcombe et al. | 1979 |
| Brook Trout | <i>Salvelinus fontinalis</i> | Embryo | 12 weeks | NOEC | Mortality | 709 | 45.9 | 1370 | 7.2-7.9 | Holcombe et al. | 1979 |
| Brook Trout | <i>Salvelinus fontinalis</i> | Embryo | 12 weeks | NOEC | Mortality | 1353 | 45.9 | 2615 | 7.2-7.9 | Holcombe et al. | 1979 |
| Brook Trout | <i>Salvelinus fontinalis</i> | Embryo | 8 weeks | NOEC | Mortality | 724 | 45.9 | 1399 | 7.2-7.9 | Holcombe et al. | 1979 |
| Brook Trout | <i>Salvelinus fontinalis</i> | 30-45-d old fry | 7 d | IC25 | Growth | 486.61 | | 487 | | Lazorchak and Smith | 2007 |
| Brook Trout | <i>Salvelinus fontinalis</i> | 30-45-d old fry | 7 d | LC50 | survival | 798.91 | | 799 | | Lazorchak and Smith | 2007 |
| Brook Trout | <i>Salvelinus fontinalis</i> | 30-45-d old fry | 7 d | LOEC | Growth | 500 | | 500 | | Lazorchak and Smith | 2007 |
| Brook Trout | <i>Salvelinus fontinalis</i> | 30-45-d old fry | 7 d | LOEC | survival | 594.6 | | 595 | | Lazorchak and Smith | 2007 |
| Brook Trout | <i>Salvelinus fontinalis</i> | 30-45-d old fry | 7 d | MATC | survival | 443 | | 443 | | Lazorchak and Smith | 2007 |
| Brook Trout | <i>Salvelinus fontinalis</i> | 30-45-d old fry | 7 d | NOEC | Growth | 287.17 | | 287 | | Lazorchak and Smith | 2007 |
| Brook Trout | <i>Salvelinus fontinalis</i> | 30-45-d old fry | 7 d | NOEC | survival | 329.87 | | 330 | | Lazorchak and Smith | 2007 |
| Brook Trout | <i>Salvelinus fontinalis</i> | Juvenile | 14 d | LC10 | Mortality | 445 | | 445 | 7.3 | Nehring and Goettl | 1974 |
| Brook Trout | <i>Salvelinus fontinalis</i> | Juvenile | 14 d | LC50 | Mortality | 960 | | 960 | 7.3 | Nehring and Goettl | 1974 |
| Bryozoan | <i>Lophopodella carteri</i> | 2-3 days | 96 h | LC50 | Mortality | 4093 | 205 | 2230 | 6.7-7.0 | Pardue and Wood | 1980 |
| Bryozoan | <i>Lophopodella carteri</i> | 2-3 days | 96 h | LC50 | Mortality | 5630 | 205 | 3067 | 6.7-7.0 | Pardue and Wood | 1980 |
| Bryozoan | <i>Pectinatella magnifica</i> | 2-3 days | 96 h | LC10 | Mortality | 2286 | 205 | 1245 | 6.7-7.0 | Pardue and Wood | 1980 |
| Bryozoan | <i>Pectinatella magnifica</i> | 2-3 days | 96 h | LC50 | Mortality | 4310 | 205 | 2348 | 6.7-7.0 | Pardue and Wood | 1980 |
| Bryozoan | <i>Plumatella emarginata</i> | 2-3 days | 96 h | LC10 | Mortality | 3474 | 205 | 1893 | 6.7-7.0 | Pardue and Wood | 1980 |
| Bryozoan | <i>Plumatella emarginata</i> | 2-3 days | 96 h | LC50 | Mortality | 5300 | 205 | 2888 | 6.7-7.0 | Pardue and Wood | 1980 |
| Chinook salmon | <i>Oncorhynchus tshawytscha</i> | Alevin | 200 h | LC10 | Mortality | 364-661 | 23 | 1262-2292 | 7.1 | Chapman | 1978 |
| Chinook salmon | <i>Oncorhynchus tshawytscha</i> | Fry | 200 h | LC10 | Mortality | 68 | 23 | 236 | 7.1 | Chapman | 1978 |
| Chinook salmon | <i>Oncorhynchus tshawytscha</i> | Fry | 200 h | LC50 | Mortality | 97 | 23 | 336 | 7.1 | Chapman | 1978 |
| Chironomid | <i>Chironomus riparius</i> | 1st instar | 11 weeks | LOEC | Development | 100 | | 100 | 7.3-7.7 | Timmermans et al. | 1992 |
| Chironomid | <i>Tanytarsus dissimilis</i> | Larva | 10 d | LC50 | Mortality | 36.8 | 46.8 | 70 | 7.3- 7.7 | Anderson et al | 1980 |
| Common duckmeat | <i>Spirodela polyrrhiza</i> | Adult | 4 d | IC50 | Growth | 935 | ND | 935 | | Gaur et al. | 1994 |
| crayfish | <i>Orconectes virilis</i> | Adult | 14 d | LC10 | Mortality | 9920 | 26 | 31006 | 7.1 | Mirenda | 1986 |

Table C.1: Summary of long-term effects of zinc on a variety of aquatic organisms

| Species Common Name | Scientific Name | Life Stage | Duration | Endpoint | Observed Effect | Effect Concentration (ug/L) | Test Hardness (mg/L) | Effect Concentration (ug/L) at 100 mg/L Hardness ¹ | pH | Authors | Year |
|---------------------|--------------------------------|--------------|----------|----------|-------------------|-----------------------------|----------------------|---------------------------------------------------------------|-------------|---------------------|------|
| crayfish | <i>Orconectes virilis</i> | Adult | 14 d | LC50 | Mortality | 84000 | 26 | 262550 | 7.1 | Mirenda | 1986 |
| Cutthroat trout | <i>Oncorhynchus clarkii</i> | Juvenile | 14 d | LC10 | Mortality | 453 | 40 | 983 | 7.2 | Nehring and Goettl | 1974 |
| Cutthroat trout | <i>Oncorhynchus clarkii</i> | Juvenile | 14 d | LC50 | Mortality | 670 | 40 | 1455 | 7.2 | Nehring and Goettl | 1974 |
| Diatom | <i>Cyclotella meneghiniana</i> | Population | 5 d | LC10 | Growth rate | 10689 | 121 | 9097 | 6.8 | Cairns et al. | 1978 |
| Diatom | <i>Cyclotella meneghiniana</i> | Population | 5 d | LC10 | Growth rate | 2803 | 121 | 2386 | 6.8 | Cairns et al. | 1978 |
| Diatom | <i>Cyclotella meneghiniana</i> | Population | 5 d | LC10 | Growth rate | 5716 | 121 | 4865 | 6.8 | Cairns et al. | 1978 |
| Duckweed | <i>Lemna minor</i> | Adult | 7 d | IC10 | Growth | 318 | | 318 | 6.30-6.40 | Dirilgen and Inel | 1994 |
| Duckweed | <i>Lemna minor</i> | Not reported | 7 d | EC50 | Growth | 3014.48 | | 3014 | 5.5 +/- 0.2 | Drost et al | 2007 |
| Duckweed | <i>Lemna minor</i> | Not reported | 7 d | EC10 | Growth | 1379.05 | | 1379 | 6 | Ince et al | 1999 |
| Duckweed | <i>Lemna minor</i> | Not reported | 7 d | EC50 | Growth | 9600 | | 9600 | 6 | Ince et al | 1999 |
| Eurasian minnow | <i>Phoxinus phoxinus</i> | Yearling | 150 d | LC10 | Mortality | 102 | 71 | 136 | 7.1-8.2 | Bengtsson | 1974 |
| Eurasian minnow | <i>Phoxinus phoxinus</i> | Adult | 30 d | LOEC | Growth | 200 | 71 | 267 | 7.1-8.2 | Bengtsson | 1974 |
| Eurasian minnow | <i>Phoxinus phoxinus</i> | Adult | 150 d | LOEC | Growth | 200 | 71 | 267 | 7.1-8.2 | Bengtsson | 1974 |
| Eurasian minnow | <i>Phoxinus phoxinus</i> | Yearling | 30 d | LOEC | Growth | 130 | 71 | 174 | 7.1-8.2 | Bengtsson | 1974 |
| Eurasian minnow | <i>Phoxinus phoxinus</i> | Yearling | 150 d | LOEC | Growth | 50 | 71 | 67 | 7.1-8.2 | Bengtsson | 1974 |
| Eurasian minnow | <i>Phoxinus phoxinus</i> | Adult | 30 d | MATC | Growth | 161 | 71 | 215 | 7.1-8.2 | Bengtsson | 1974 |
| Eurasian minnow | <i>Phoxinus phoxinus</i> | Adult | 150 d | MATC | Growth | 161 | 71 | 215 | 7.1-8.2 | Bengtsson | 1974 |
| Eurasian minnow | <i>Phoxinus phoxinus</i> | Yearling | 30 d | MATC | Growth | 81 | 71 | 108 | 7.1-8.2 | Bengtsson | 1974 |
| Eurasian minnow | <i>Phoxinus phoxinus</i> | Yearling | 150 d | MATC | Growth | 32 | 71 | 43 | 7.1-8.2 | Bengtsson | 1974 |
| Eurasian minnow | <i>Phoxinus phoxinus</i> | Adult | 30 d | NOEC | Growth | 130 | 71 | 174 | 7.1-8.2 | Bengtsson | 1974 |
| Eurasian minnow | <i>Phoxinus phoxinus</i> | Adult | 150 d | NOEC | Growth | 130 | 71 | 174 | 7.1-8.2 | Bengtsson | 1974 |
| Eurasian minnow | <i>Phoxinus phoxinus</i> | Yearling | 30 d | NOEC | Growth | 50 | 71 | 67 | 7.1-8.2 | Bengtsson | 1974 |
| Eurasian minnow | <i>Phoxinus phoxinus</i> | Yearling | 150 d | NOEC | Growth | 20 | 71 | 27 | 7.1-8.2 | Bengtsson | 1974 |
| Fathead minnow | <i>Pimephales promelas</i> | Egg | NR | EC10 | Eggs adhesiveness | 46.9 | 46 | 90 | 7-8 | Benoit and Holcombe | 1978 |
| Fathead minnow | <i>Pimephales promelas</i> | Egg | NR | LOEC/L | Eggs adhesiveness | 145 | 46 | 280 | 7-8 | Benoit and Holcombe | 1978 |
| Fathead minnow | <i>Pimephales promelas</i> | Egg | NR | LOEC/L | Hatching success | 295 | 46 | 569 | 7-8 | Benoit and Holcombe | 1978 |
| Fathead minnow | <i>Pimephales promelas</i> | Egg | NR | LOEC/L | Mortality | 295 | 46 | 569 | 7-8 | Benoit and Holcombe | 1978 |
| Fathead minnow | <i>Pimephales promelas</i> | Egg | NR | MATC | Eggs adhesiveness | 106 | 46 | 204 | 7-8 | Benoit and Holcombe | 1978 |
| Fathead minnow | <i>Pimephales promelas</i> | Egg | NR | MATC | Hatching success | 207 | 46 | 399 | 7-8 | Benoit and Holcombe | 1978 |
| Fathead minnow | <i>Pimephales promelas</i> | Egg | NR | MATC | Mortality | 207 | 46 | 399 | 7-8 | Benoit and Holcombe | 1978 |
| Fathead minnow | <i>Pimephales promelas</i> | Egg | NR | NOEC/L | Eggs adhesiveness | 78 | 46 | 150 | 7-8 | Benoit and Holcombe | 1978 |
| Fathead minnow | <i>Pimephales promelas</i> | Egg | NR | NOEC/L | Hatching success | 145 | 46 | 280 | 7-8 | Benoit and Holcombe | 1978 |
| Fathead minnow | <i>Pimephales promelas</i> | Egg | NR | NOEC/L | Mortality | 145 | 46 | 280 | 7-8 | Benoit and Holcombe | 1978 |
| Fathead minnow | <i>Pimephales promelas</i> | Larva | 7 d | ChV | Growth | 430 | 190 | 250 | 8.3-8.7 | Magliette et al. | 1995 |
| Fathead minnow | <i>Pimephales promelas</i> | Larva | 7 d | LC50 | Mortality | 780 | 190 | 453 | 8.3-8.7 | Magliette et al. | 1995 |
| Fathead minnow | <i>Pimephales promelas</i> | Larva | 7 d | LOEC/L | Growth | 630 | 190 | 366 | 8.3-8.7 | Magliette et al. | 1995 |
| Fathead minnow | <i>Pimephales promelas</i> | Larva | 7 d | IC10 | Growth | 83.9 | 48 | 156 | | Norberg and Mount | 1985 |
| Fathead minnow | <i>Pimephales promelas</i> | Larva | 7 d | LOEC | Growth | 374 | 48 | 696 | | Norberg and Mount | 1985 |
| Fathead minnow | <i>Pimephales promelas</i> | Larva | 7 d | LOEC | Mortality | 184 | 48 | 342 | | Norberg and Mount | 1985 |
| Fathead minnow | <i>Pimephales promelas</i> | Larva | 7 d | MATC | Growth | 262 | 48 | 487 | | Norberg and Mount | 1985 |
| Fathead minnow | <i>Pimephales promelas</i> | Larva | 7 d | MATC | Mortality | 125 | 48 | 233 | | Norberg and Mount | 1985 |
| Fathead minnow | <i>Pimephales promelas</i> | Larva | 7 d | NOEC | Growth | 184 | 48 | 342 | | Norberg and Mount | 1985 |
| Fathead minnow | <i>Pimephales promelas</i> | Larva | 7 d | NOEC | Mortality | 84.6 | 48 | 157 | | Norberg and Mount | 1985 |
| Fathead minnow | <i>Pimephales promelas</i> | Larva | 7 d | ChV | Growth | 184 | 47 | 349 | 7.4-8.2 | Norberg-King | 1989 |
| Fathead minnow | <i>Pimephales promelas</i> | Larva | 7 d | ChV | Growth | 315 | 47 | 597 | 7.4-8.2 | Norberg-King | 1989 |
| Fathead minnow | <i>Pimephales promelas</i> | Larva | 32 d | ChV | Mortality | 188 | 47 | 356 | 7.4-8.2 | Norberg-King | 1989 |
| Fathead minnow | <i>Pimephales promelas</i> | Larva | 7 d | LC50 | Mortality | 250 | 47 | 474 | 7.4-8.2 | Norberg-King | 1989 |
| Fathead minnow | <i>Pimephales promelas</i> | Larva | 7 d | LC50 | Mortality | 283 | 47 | 536 | 7.4-8.2 | Norberg-King | 1989 |
| Fathead minnow | <i>Pimephales promelas</i> | Larva | 7 d | LOEC | Growth | 278 | 47 | 527 | 7.4-8.2 | Norberg-King | 1989 |
| Fathead minnow | <i>Pimephales promelas</i> | Larva | 7 d | LOEC | Growth | 454 | 47 | 860 | 7.4-8.2 | Norberg-King | 1989 |
| Fathead minnow | <i>Pimephales promelas</i> | Larva | 32 d | LOEC | Mortality | 275 | 47 | 521 | 7.4-8.2 | Norberg-King | 1989 |
| Fathead minnow | <i>Pimephales promelas</i> | Larva | 7 d | NOEC | Growth | 122 | 47 | 231 | 7.4-8.2 | Norberg-King | 1989 |

Table C.1: Summary of long-term effects of zinc on a variety of aquatic organisms

| Species Common Name | Scientific Name | Life Stage | Duration | Endpoint | Observed Effect | Effect Concentration (ug/L) | Test Hardness (mg/L) | Effect Concentration (ug/L) at 100 mg/L Hardness ¹ | pH | Authors | Year |
|---------------------|----------------------------------------|--------------------------|----------|----------|-----------------|-----------------------------|----------------------|---------------------------------------------------------------|---------|------------------------|------|
| Fathead minnow | <i>Pimephales promelas</i> | Larva | 7 d | NOEC | Growth | 218 | 47 | 413 | 7.4-8.2 | Norberg-King | 1989 |
| Fathead minnow | <i>Pimephales promelas</i> | Larva | 32 d | NOEC | Mortality | 129 | 47 | 244 | 7.4-8.2 | Norberg-King | 1989 |
| Fathead minnow | <i>Pimephales promelas</i> | Egg | 12 d | NOEC | Mortality | 1050 | 186 | 621 | 7.5-7.6 | Pickering and Vigor | 1965 |
| Fathead minnow | <i>Pimephales promelas</i> | Fry | 7 d | NOEC | Mortality | 560 | 186 | 331 | 7.5-7.6 | Pickering and Vigor | 1965 |
| Fathead minnow | <i>Pimephales promelas</i> | Egg | 7 d | TLM | Mortality | 1690 | 186 | 1000 | 7.5-7.6 | Pickering and Vigor | 1965 |
| Fathead minnow | <i>Pimephales promelas</i> | Egg | 12 d | TLM | Mortality | 1630 | 186 | 964 | 7.5-7.6 | Pickering and Vigor | 1965 |
| Fathead minnow | <i>Pimephales promelas</i> | Fry | 7 d | TLM | Mortality | 870 | 186 | 515 | 7.5-7.6 | Pickering and Vigor | 1965 |
| Green alga | <i>Chlamydomonas sp.</i> | Population | 10 d | LC10 | Growth rate | 8381 | 121 | 7133 | 6.8 | Cairns et al. | 1978 |
| Green alga | <i>Chlamydomonas sp.</i> | Population | 10 d | LC10 | Growth rate | 9398 | 121 | 7998 | 6.8 | Cairns et al. | 1978 |
| Green alga | <i>Chlorella pyrenoidosa</i> | Not reported | 24 h | EC50 | Growth | 57 | 25.51 | 181 | | Lin et al | 2007 |
| Green alga | <i>Chlorella pyrenoidosa</i> | Not reported | 24 h | LOEC | Cell density | 40 | 25.51 | 127 | | Lin et al | 2007 |
| Green alga | <i>Chlorella pyrenoidosa</i> | Not reported | 24 h | MATC | Cell density | 28.28 | 25.51 | 90 | | Lin et al | 2007 |
| Green alga | <i>Chlorella pyrenoidosa</i> | Not reported | 24 h | NOEC | Cell density | 20 | 25.51 | 64 | | Lin et al | 2007 |
| Green alga | <i>Chlorella vulgaris</i> | exponential growth phase | 72 h | EC50 | biomass | 34 | | 34 | | Muyssen and Janssen | 2001 |
| Green alga | <i>Chlorella vulgaris</i> | exponential growth phase | 72 h | EC50 | Growth | 153 | | 153 | | Muyssen and Janssen | 2001 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Not reported | 24 h | EC50 | Growth | 15 | | 15 | | Chen et al | 1997 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Not reported | 96 h | EC50 | Growth | 178 | | 178 | | Chen et al | 1997 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Not reported | 7 d | EC10 | Growth | 1.05 | | 1.1 | 6.0-6.3 | Chiaaudani and Vighi | 1978 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Not reported | 96 h | EC10 | Growth | 1.32 | | 1.3 | 6.0-6.3 | Chiaaudani and Vighi | 1978 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Not reported | 96 h | EC10 | Growth | 11.74 | | 12 | 6.0-6.3 | Chiaaudani and Vighi | 1978 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Not reported | 7 d | EC10 | Growth | 13.48 | | 13 | 6.0-6.3 | Chiaaudani and Vighi | 1978 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Not reported | 7 d | EC50 | Growth | 4.1 | | 4.1 | 6.0-6.3 | Chiaaudani and Vighi | 1978 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Not reported | 96 h | EC50 | Growth | 4.4 | | 4.4 | 6.0-6.3 | Chiaaudani and Vighi | 1978 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Not reported | 96 h | EC50 | Growth | 27 | | 27 | 6.0-6.3 | Chiaaudani and Vighi | 1978 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Not reported | 7 d | EC50 | Growth | 32 | | 32 | 6.0-6.3 | Chiaaudani and Vighi | 1978 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 58.1 | 19.6 | 231 | 8 | De Schampelaere et al. | 2004 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 62.3 | 19.6 | 247 | 8 | De Schampelaere et al. | 2004 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 71.2 | 19.6 | 283 | 8 | De Schampelaere et al. | 2004 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 85 | 19.6 | 337 | 7 | De Schampelaere et al. | 2004 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 131 | 19.6 | 520 | 7 | De Schampelaere et al. | 2004 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 142 | 19.6 | 564 | 6 | De Schampelaere et al. | 2004 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 142 | 19.6 | 564 | 7 | De Schampelaere et al. | 2004 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 191 | 19.6 | 758 | 6 | De Schampelaere et al. | 2004 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 215 | 19.6 | 853 | 6 | De Schampelaere et al. | 2004 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | less than 72h | 72 h | IC50 | biomass | 4.12 | 40 | 8.9 | 7.3 | Errécalde et al. | 1998 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | less than 72h | 72 h | IC50 | biomass | 32.7 | 40 | 71 | 7.3 | Errécalde et al. | 1998 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | less than 72h | 72 h | IC50 | biomass | 39.24 | 40 | 85 | 7.3 | Errécalde et al. | 1998 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | less than 72h | 72 h | IC50 | Growth rate | 11.12 | 40 | 24 | 7.3 | Errécalde et al. | 1998 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | less than 72h | 72 h | IC50 | Growth rate | 45.13 | 40 | 98 | 7.3 | Errécalde et al. | 1998 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | less than 72h | 72 h | IC50 | Growth rate | 68.68 | 40 | 149 | 7.3 | Errécalde et al. | 1998 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 10.5 | 24.4 | 35 | 7.8 | Heijerick et al. | 2002 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 15.8 | 24.4 | 52 | 7.7 | Heijerick et al. | 2002 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 21.1 | 24.4 | 70 | 7.65 | Heijerick et al. | 2002 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 26.4 | 24.4 | 87 | 7.3 | Heijerick et al. | 2002 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 26.4 | 24.4 | 87 | 7.4 | Heijerick et al. | 2002 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 29 | 24.4 | 96 | 7.6 | Heijerick et al. | 2002 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 47.5 | 24.4 | 157 | 7.1 | Heijerick et al. | 2002 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 94.8 | 24.4 | 313 | 6.8 | Heijerick et al. | 2002 |

Table C.1: Summary of long-term effects of zinc on a variety of aquatic organisms

| Species Common Name | Scientific Name | Life Stage | Duration | Endpoint | Observed Effect | Effect Concentration (ug/L) | Test Hardness (mg/L) | Effect Concentration (ug/L) at 100 mg/L Hardness ¹ | pH | Authors | Year |
|---------------------|----------------------------------------|--------------------------|----------|----------|---------------------------------|-----------------------------|----------------------|---------------------------------------------------------------|----------------|-----------------------|------|
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 119 | 24.4 | 392 | 6.2 | Heijerick et al. | 2002 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 224 | 24.4 | 739 | 5.6 | Heijerick et al. | 2002 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 8.44 | 37.2 | 19 | 7.5 | Heijerick et al. | 2002 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 10.5 | 37.2 | 24 | 7.5 | Heijerick et al. | 2002 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 10.9 | 62.3 | 16 | 7.5 | Heijerick et al. | 2002 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 11.1 | 62.3 | 17 | 7.5 | Heijerick et al. | 2002 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 14.9 | 112.3 | 14 | 7.5 | Heijerick et al. | 2002 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 27.9 | 112.3 | 25 | 7.5 | Heijerick et al. | 2002 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 15.4 | 162.3 | 10 | 7.5 | Heijerick et al. | 2002 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 33.2 | 162.3 | 22 | 7.5 | Heijerick et al. | 2002 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 17.5 | 212.4 | 9 | 7.5 | Heijerick et al. | 2002 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 34.4 | 212.4 | 18 | 7.5 | Heijerick et al. | 2002 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Population | 72 h | EC50 | biomass | 54.9 | 262.6 | 24 | 7.5 | Heijerick et al. | 2002 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | exponential growth phase | 72 h | EC50 | biomass | 39 | | 39 | | Muyssen and Janssen | 2001 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | exponential growth phase | 72 h | EC50 | Growth | 138 | | 138 | | Muyssen and Janssen | 2001 |
| Green alga | <i>Pseudokirchneriella subcapitata</i> | Not reported | 48 h | EC50 | Growth | 96 | 130 | 77 | 7.8-8.8 | Pardos et al | 1998 |
| Green alga | <i>Scenedesmus quadricauda</i> | Population | 5 d | LC10 | Growth rate | 10451 | 121 | 8894 | 6.8 | Cairns et al. | 1978 |
| Green alga | <i>Scenedesmus quadricauda</i> | Population | 5 d | LC10 | Growth rate | 9559 | 121 | 8135 | 6.8 | Cairns et al. | 1978 |
| Green alga | <i>Scenedesmus quadricauda</i> | Population | 15 d | IC10 | Growth | 96.1 | | 96 | 4.5 | Starodub et al. | 1987 |
| Green alga | <i>Scenedesmus quadricauda</i> | Population | 14 d | LOEC | Growth | 225 | | 225 | 6.5 | Starodub et al. | 1987 |
| Green alga | <i>Scenedesmus quadricauda</i> | Population | 14 d | LOEC | Growth | 500 | | 500 | 8.5 | Starodub et al. | 1987 |
| Green alga | <i>Scenedesmus quadricauda</i> | Population | 15 d | LOEC | Growth | 100 | | 100 | 4.5 | Starodub et al. | 1987 |
| Green alga | <i>Scenedesmus quadricauda</i> | Population | 14 d | MATC | Growth | 150 | | 150 | 6.5 | Starodub et al. | 1987 |
| Green alga | <i>Scenedesmus quadricauda</i> | Population | 14 d | MATC | Growth | 335 | | 335 | 8.5 | Starodub et al. | 1987 |
| Green alga | <i>Scenedesmus quadricauda</i> | Population | 14 d | NOEC | Growth | 100 | | 100 | 6.5 | Starodub et al. | 1987 |
| Green alga | <i>Scenedesmus quadricauda</i> | Population | 14 d | NOEC | Growth | 225 | | 225 | 8.5 | Starodub et al. | 1987 |
| Green hydra | <i>Hydra viridissima</i> | Not reported | 7 d | EC10 | Population growth inhibition | 52.23 | 20 | 204 | 7.25-7.53 | Holdway et al | 2001 |
| Green hydra | <i>Hydra viridissima</i> | Not reported | 7 d | LOEC | Population growth inhibition | 75 | 20 | 293 | 7.25-7.53 | Holdway et al | 2001 |
| Green hydra | <i>Hydra viridissima</i> | Not reported | 7 d | MATC | Population growth inhibition | 53.4 | 20 | 208 | 7.25-7.53 | Holdway et al | 2001 |
| Green hydra | <i>Hydra viridissima</i> | Not reported | 7 d | NOEC | Population growth inhibition | 38 | 20 | 148 | 7.25-7.53 | Holdway et al | 2001 |
| Mayfly | <i>Epeorus latifolium</i> | Larva | 4 weeks | IC10 | emergence | 14.4 | 83 | 17 | 7.9-8.0 | Hatakeyama | 1989 |
| Mayfly | <i>Epeorus latifolium</i> | Larva | 4 weeks | LC10 | Mortality | 15 | 83 | 18 | 7.9-8.0 | Hatakeyama | 1989 |
| Mayfly | <i>Rhithrogena hageni</i> | nymph | 10 d | EC10 | Mortality | 2069.2 | 44.4 | 4113 | 7.77 | Brinkman and Johnston | 2008 |
| Mayfly | <i>Rhithrogena hageni</i> | nymph | 10 d | LOEC | Mortality | 10800 | 44.4 | 21465 | 7.77 | Brinkman and Johnston | 2008 |
| Mayfly | <i>Rhithrogena hageni</i> | nymph | 10 d | MATC | Mortality | 7565.71 | 44.4 | 15037 | 7.77 | Brinkman and Johnston | 2008 |
| Mayfly | <i>Rhithrogena hageni</i> | nymph | 10 d | NOEC | Mortality | 5300 | 44.4 | 10534 | 7.77 | Brinkman and Johnston | 2008 |
| Mixed invertebrates | N/A | Population | 14 d | LOEC | Community similarity | 17.1 | 135 | 13 | | Marshall et al. | 1983 |
| Mixed invertebrates | N/A | Population | 14 d | LOEC | Primary productivity | 17.1 | 135 | 13 | | Marshall et al. | 1983 |
| Mixed invertebrates | N/A | Population | 14 d | LOEC | Specific zooplanton populations | 17.1 | 135 | 13 | | Marshall et al. | 1983 |
| Mixed invertebrates | N/A | Population | 14 d | LOEC | Zooplancion species diversity | 17.1 | 135 | 13 | | Marshall et al. | 1983 |
| Mottled sculpin | <i>Cottus bairdi</i> | less than 2 months old | 30 d | EC10 | Mortality | 155.7 | 154 | 108 | 7.5 (7.4-7.7) | Brinkman and Woodling | 2005 |
| Mottled sculpin | <i>Cottus bairdi</i> | less than 2 months old | 30 d | LOEC | Mortality | 379 | 154 | 263 | 7.5 (7.4-7.7) | Brinkman and Woodling | 2005 |
| Mottled sculpin | <i>Cottus bairdi</i> | less than 2 months old | 30 d | MATC | Mortality | 255 | 154 | 177 | 7.5 (7.4-7.7) | Brinkman and Woodling | 2005 |
| Mottled sculpin | <i>Cottus bairdi</i> | less than 2 months old | 30 d | NOEC | Mortality | 172 | 154 | 119 | 7.5 (7.4-7.7) | Brinkman and Woodling | 2005 |
| Mottled sculpin | <i>Cottus bairdi</i> | newly emerged | 30 d | LC50 | Mortality | 32 | 48.6 | 59 | 7.38 (7.2-7.6) | Woodling et al | 2002 |
| Pink hydra | <i>Hydra vulgaris</i> | Not reported | 7 d | EC10 | Population growth inhibition | 177.93 | 20 | 694 | 7.25-7.53 | Holdway et al | 2001 |
| Pink hydra | <i>Hydra vulgaris</i> | Not reported | 7 d | LOEC | Population growth inhibition | 250 | 20 | 976 | 7.25-7.53 | Holdway et al | 2001 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Embryo | 72 d | LC10 | Mortality | 458 | 25 | 1480 | 6.9-7.1 | Cairns and Garton | 1982 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Embryo | 72 d | LOEC | Mortality | 819 | 25 | 2646 | 6.9-7.1 | Cairns and Garton | 1982 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Embryo | 72 d | MATC | Mortality | 603 | 25 | 1948 | 6.9-7.1 | Cairns and Garton | 1982 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Embryo | 72 d | NOEC | Mortality | 444 | 25 | 1435 | 6.9-7.1 | Cairns and Garton | 1982 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Alevin | 186 h | LC10 | Mortality | 256 | 23 | 888 | 7.1 | Chapman | 1978 |

Table C.1: Summary of long-term effects of zinc on a variety of aquatic organisms

| Species Common Name | Scientific Name | Life Stage | Duration | Endpoint | Observed Effect | Effect Concentration (ug/L) | Test Hardness (mg/L) | Effect Concentration (ug/L) at 100 mg/L Hardness ¹ | pH | Authors | Year |
|---------------------|----------------------------|-----------------|----------|----------|-----------------|-----------------------------|----------------------|---------------------------------------------------------------|--------------|------------------------------|------|
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Swim-up fry | 200 h | LC10 | Mortality | 54 | 23 | 187 | 7.1 | Chapman | 1978 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Alevin | 186 h | LC50 | Mortality | 555 | 23 | 1924 | 7.1 | Chapman | 1978 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Swim-up fry | 200 h | LC50 | Mortality | 93 | 23 | 322 | 7.1 | Chapman | 1978 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LC10 | Mortality | 312 | 29.1 | 887 | 5.68 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LC10 | Mortality | 46.1 | 29.1 | 131 | 7.65 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LC10 | Mortality | 73.6 | 29.1 | 209 | 7.58 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LC10 | Mortality | 99.1 | 29.1 | 282 | 6.78 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LC10 | Mortality | 38.4 | 29.6 | 108 | 7.45 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LC10 | Mortality | 34.5 | 31.72 | 91 | 7.58-7.87 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LC10 | Mortality | 171 | 104.99 | 164 | 7.58-7.87 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LC10 | Mortality | 290 | 190.35 | 168 | 7.58-7.87 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LC10 | Mortality | 337 | 398.68 | 105 | 7.58-7.87 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LOEC | Growth | >1280 | 29.1 | >3637 | 6.7 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LOEC | Growth | >1280 | 29.1 | >3637 | 7.74 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LOEC | Growth | >1740 | 29.1 | >4944 | 7.58 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LOEC | Growth | >800 | 29.1 | >2273 | 5.68 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LOEC | Growth | 345 | 29.1 | 980 | 7.61 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LOEC | Growth | >375 | 29.6 | >1050 | 7.45 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LOEC | Growth | 2310 | 29.6 | 6470 | 7.87 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LOEC | survival | 162 | 29.1 | 460 | 7.65 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LOEC | survival | 166 | 29.1 | 472 | 7.61 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LOEC | survival | 117 | 29.6 | 328 | 7.45 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LOEC | survival | 2310 | 29.6 | 6470 | 7.87 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | NOEC | Growth | 166 | 29.1 | 472 | 7.61 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | NOEC | Growth | 974 | 29.6 | 2728 | 7.87 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | NOEC | survival | 45.4 | 29.1 | 129 | 7.73 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | NOEC | survival | 78.9 | 29.1 | 224 | 7.61 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | NOEC | survival | 31.5 | 29.6 | 88 | 7.45 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | NOEC | survival | 974 | 29.6 | 2728 | 7.87 | De Schamphelaere and Janssen | 2004 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LC10 | Mortality | 259 | 23.4 | 885 | 6.15 | De Schamphelaere et al. | 2005 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LC10 | Mortality | 185 | 28.2 | 540 | 6.8 | De Schamphelaere et al. | 2005 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LC10 | Mortality | 219 | 31.5 | 582 | 7.08 | De Schamphelaere et al. | 2005 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LC10 | Mortality | 902 | 103.7 | 875 | 7.76 | De Schamphelaere et al. | 2005 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LC10 | Mortality | 578 | 176.3 | 358 | 8.13 | De Schamphelaere et al. | 2005 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LC50 | Mortality | 582 | 23.4 | 1989 | 6.15 | De Schamphelaere et al. | 2005 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LC50 | Mortality | 406 | 28.2 | 1185 | 6.8 | De Schamphelaere et al. | 2005 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LC50 | Mortality | 337 | 31.5 | 895 | 7.08 | De Schamphelaere et al. | 2005 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LC50 | Mortality | 1970 | 103.7 | 1910 | 7.76 | De Schamphelaere et al. | 2005 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | LC50 | Mortality | 1850 | 176.3 | 1145 | 8.13 | De Schamphelaere et al. | 2005 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | NOEC | Mortality | 370 | 23.4 | 1264 | 6.15 | De Schamphelaere et al. | 2005 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | NOEC | Mortality | 324 | 28.2 | 945 | 6.8 | De Schamphelaere et al. | 2005 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | NOEC | Mortality | 199 | 31.5 | 529 | 7.08 | De Schamphelaere et al. | 2005 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | NOEC | Mortality | 771 | 103.7 | 748 | 7.76 | De Schamphelaere et al. | 2005 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 30 d | NOEC | Mortality | 696 | 176.3 | 431 | 8.13 | De Schamphelaere et al. | 2005 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | 15-25-d old fry | 7 d | IC25 | Growth | 148.03 | | 148 | | Lazorchak and Smith | 2007 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | 15-25-d old fry | 7 d | LC50 | survival | 195.38 | | 195 | | Lazorchak and Smith | 2007 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | 15-25-d old fry | 7 d | LOEC | Growth | 250 | | 250 | | Lazorchak and Smith | 2007 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | 15-25-d old fry | 7 d | LOEC | survival | 250 | | 250 | | Lazorchak and Smith | 2007 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | 15-25-d old fry | 7 d | MATC | survival | 177 | | 177 | | Lazorchak and Smith | 2007 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | 15-25-d old fry | 7 d | NOEC | Growth | 114.63 | | 115 | | Lazorchak and Smith | 2007 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | 15-25-d old fry | 7 d | NOEC | survival | 125 | | 125 | | Lazorchak and Smith | 2007 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Fry | 69 days | EC10 | Length | 300 | 19.7 | 1186 | 6.75 +/- 0.4 | Mebane et al | 2008 |

Table C.1: Summary of long-term effects of zinc on a variety of aquatic organisms

| Species Common Name | Scientific Name | Life Stage | Duration | Endpoint | Observed Effect | Effect Concentration (ug/L) | Test Hardness (mg/L) | Effect Concentration (ug/L) at 100 mg/L Hardness ¹ | pH | Authors | Year |
|---------------------|-------------------------------|----------------------|-----------|----------|------------------------------------------|-----------------------------|----------------------|---------------------------------------------------------------|-----------------|---------------------------|------|
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Fry | 69 days | EC10 | Mortality | 88 | 19.7 | 348 | 6.75 +/- 0.4 | Mebane et al | 2008 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Fry | 69 days | EC10 | Weight | 199 | 19.7 | 787 | 6.75 +/- 0.4 | Mebane et al | 2008 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Fry | 69 days | EC20 | Mortality | 147 | 19.7 | 581 | 6.75 +/- 0.4 | Mebane et al | 2008 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Fry | 69 days | EC20 | Weight | 387 | 19.7 | 1530 | 6.75 +/- 0.4 | Mebane et al | 2008 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Fry | 69 days | LOEC | Length | 365 | 19.7 | 1443 | 6.75 +/- 0.4 | Mebane et al | 2008 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Fry | 69 days | LOEC | Mortality | 117 | 19.7 | 462 | 6.75 +/- 0.4 | Mebane et al | 2008 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Fry | 69 days | LOEC | Weight | 365 | 19.7 | 1443 | 6.75 +/- 0.4 | Mebane et al | 2008 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Fry | 69 days | MATC | Length | 279 | 19.7 | 1103 | 6.75 +/- 0.4 | Mebane et al | 2008 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Fry | 69 days | MATC | Weight | 279 | 19.7 | 1103 | 6.75 +/- 0.4 | Mebane et al | 2008 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Fry | 69 days | NOEC | Length | 214 | 19.7 | 846 | 6.75 +/- 0.4 | Mebane et al | 2008 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Fry | 69 days | NOEC | Weight | 214 | 19.7 | 846 | 6.75 +/- 0.4 | Mebane et al | 2008 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 14 d | LC10 | Mortality | 318 | 37 | 737 | 7.3 | Nehring and Goettl | 1974 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Juvenile | 14 d | LC50 | Mortality | 410 | 37 | 951 | 7.3 | Nehring and Goettl | 1974 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Fingerling | 22 months | LOEC/L | Mortality | 640 | 333 | 231 | 7.81 | Sinley et al. | 1974 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Fingerling | 22 months | MATC | Mortality | 453 | 333 | 164 | 7.81 | Sinley et al. | 1974 |
| Rainbow trout | <i>Oncorhynchus mykiss</i> | Fingerling | 22 months | NOEC/L | Mortality | 320 | 333 | 116 | 7.81 | Sinley et al. | 1974 |
| Rotifer | <i>Brachionus havanaensis</i> | adults and juveniles | 18 d | EC10 | Population growth inhibition | 78.2 | | 78 | 7.1-7.3 | Juarez-Franco et al | 2007 |
| Rotifer | <i>Brachionus havanaensis</i> | adults and juveniles | 18 d | LOEC | Population growth inhibition | 141.94 | | 142 | 7.1-7.3 | Juarez-Franco et al | 2007 |
| Rotifer | <i>Brachionus havanaensis</i> | adults and juveniles | 18 d | MATC | Population growth inhibition | 100.36 | | 100 | 7.1-7.3 | Juarez-Franco et al | 2007 |
| Rotifer | <i>Brachionus havanaensis</i> | adults and juveniles | 18 d | NOEC | Population growth inhibition | 70.96 | | 71 | 7.1-7.3 | Juarez-Franco et al | 2007 |
| Sea trout | <i>Salmo trutta</i> | Juvenile | 14 d | LC10 | Mortality | 504 | 39 | 1118 | 7.2 | Nehring and Goettl | 1974 |
| Sea trout | <i>Salmo trutta</i> | Juvenile | 14 d | LC50 | Mortality | 640 | 39 | 1420 | 7.2 | Nehring and Goettl | 1974 |
| Snail | <i>Physa gyrina</i> | Adult | 30 d | LC50 | Mortality | 771 | 36 | 1830 | 6.9 | Nebeker et al. | 1986 |
| Snail | <i>Physa gyrina</i> | Adult | 30 d | NOEC/L | Mortality | 570 | 36 | 1353 | 6.9 | Nebeker et al. | 1986 |
| Snail | <i>Potamopyrgus jenkinsi</i> | Juvenile | 12 weeks | EC50 | Growth | 103 | 159 | 70 | 7.8-8.2 | Dorgelo et al. | 1995 |
| Snail | <i>Potamopyrgus jenkinsi</i> | Juvenile | 12 weeks | LOEC | Growth | 115 | 159 | 78 | 7.8-8.2 | Dorgelo et al. | 1995 |
| Snail | <i>Potamopyrgus jenkinsi</i> | Juvenile | 12 weeks | MATC | Growth | 91 | 159 | 61 | 7.8-8.2 | Dorgelo et al. | 1995 |
| Snail | <i>Potamopyrgus jenkinsi</i> | Juvenile | 12 weeks | NOEC | Growth | 72 | 159 | 49 | 7.8-8.2 | Dorgelo et al. | 1995 |
| Star duckweed | <i>Lemna trisulca</i> | Not reported | 14 d | EC50 | final yield (oven dry weight) | 327 | 20.37 | 1256 | 7.8-8.3 +/- 0.3 | Huebert and Shay | 1992 |
| Star duckweed | <i>Lemna trisulca</i> | Not reported | 14 d | EC50 | multiplication rate (number of fronds) | 915.6 | 20.37 | 3518 | 7.8-8.3 +/- 0.3 | Huebert and Shay | 1992 |
| Water flea | <i>Ceriodaphnia dubia</i> | Less than 24hrs | 4 weeks | LOEC | Reproduction - Number of young per adult | 25 | 97.6 | 26 | 6 | Belanger et Cherry | 1990 |
| Water flea | <i>Ceriodaphnia dubia</i> | Less than 24hrs | 4 weeks | LOEC | Reproduction - Number of young per adult | 25 | 97.6 | 26 | 8 | Belanger et Cherry | 1990 |
| Water flea | <i>Ceriodaphnia dubia</i> | Less than 24hrs | 4 weeks | LOEC | Reproduction - Number of young per adult | 50 | 97.6 | 51 | 9 | Belanger et Cherry | 1990 |
| Water flea | <i>Ceriodaphnia dubia</i> | Less than 24hrs | 4 weeks | LOEC | Reproduction - Number of young per adult | 100 | 113.6 | 90 | 6, 8, 9 | Belanger et Cherry | 1990 |
| Water flea | <i>Ceriodaphnia dubia</i> | Less than 24hrs | 4 weeks | LOEC | Reproduction - Number of young per adult | 100 | 182 | 60 | 8 | Belanger et Cherry | 1990 |
| Water flea | <i>Ceriodaphnia dubia</i> | Less than 24hrs | 4 weeks | LOEC | Reproduction - Number of young per adult | 50 | 182 | 30 | 6 | Belanger et Cherry | 1990 |
| Water flea | <i>Ceriodaphnia dubia</i> | Less than 24hrs | 4 weeks | MATC | Reproduction - Number of young per adult | 35 | 97.6 | 36 | 9 | Belanger et Cherry | 1990 |
| Water flea | <i>Ceriodaphnia dubia</i> | Less than 24hrs | 4 weeks | MATC | Reproduction - Number of young per adult | 71 | 113.6 | 64 | 6, 8, 9 | Belanger et Cherry | 1990 |
| Water flea | <i>Ceriodaphnia dubia</i> | Less than 24hrs | 4 weeks | MATC | Reproduction - Number of young per adult | 71 | 182 | 43 | 8 | Belanger et Cherry | 1990 |
| Water flea | <i>Ceriodaphnia dubia</i> | Less than 24hrs | 4 weeks | NOEC | Reproduction - Number of young per adult | 25 | 97.6 | 26 | 9 | Belanger et Cherry | 1990 |
| Water flea | <i>Ceriodaphnia dubia</i> | Less than 24hrs | 4 weeks | NOEC | Reproduction - Number of young per adult | 50 | 113.6 | 45 | 6, 8, 9 | Belanger et Cherry | 1990 |
| Water flea | <i>Ceriodaphnia dubia</i> | Less than 24hrs | 4 weeks | NOEC | Reproduction - Number of young per adult | 100 | 182 | 60 | 9 | Belanger et Cherry | 1990 |
| Water flea | <i>Ceriodaphnia dubia</i> | Less than 24hrs | 4 weeks | NOEC | Reproduction - Number of young per adult | 50 | 182 | 30 | 8 | Belanger et Cherry | 1990 |
| Water flea | <i>Ceriodaphnia dubia</i> | 60-84 h | 96 h | ChV | Mortality | 70 | 169 | 45 | 7.8-8.2 | Masters et al. | 1991 |
| Water flea | <i>Ceriodaphnia dubia</i> | Less than 24hrs | 7 d | ChV | Mortality | 90 | 169 | 58 | 7.8-8.2 | Masters et al. | 1991 |
| Water flea | <i>Ceriodaphnia dubia</i> | 60-84 h | 96 h | ChV | Reproduction - Brood size | 45 | 169 | 29 | 7.8-8.2 | Masters et al. | 1991 |
| Water flea | <i>Ceriodaphnia dubia</i> | Less than 24hrs | 7 d | ChV | Reproduction - Brood size | 105 | 169 | 67 | 7.8-8.2 | Masters et al. | 1991 |
| Water flea | <i>Ceriodaphnia dubia</i> | Juvenile | 9 d | EC50 | Immobility | 354 | 280 | 148 | 7.8 | Muyssen and Janssen | 2002 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | EC16 | Repro - Number of young per survivor | 70 | 49 | 128 | 7.4-8.2 | Biesinger and Christensen | 1972 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | EC50 | Repro - Number of young per survivor | 102 | 49 | 187 | 7.4-8.2 | Biesinger and Christensen | 1972 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | LC50 | Immobility | 158 | 49 | 289 | 7.4-8.2 | Biesinger and Christensen | 1972 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | EC10 | Reproduction - Number of young per adult | 92.1 | 26.5 | 283 | 7.3 | De Schampelaere et al. | 2005 |

Table C.1: Summary of long-term effects of zinc on a variety of aquatic organisms

| Species Common Name | Scientific Name | Life Stage | Duration | Endpoint | Observed Effect | Effect Concentration (ug/L) | Test Hardness (mg/L) | Effect Concentration (ug/L) at 100 mg/L Hardness ¹ | pH | Authors | Year |
|---------------------|----------------------|-----------------|----------|----------|------------------------------------------|-----------------------------|----------------------|---------------------------------------------------------------|------|-------------------------|------|
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | EC10 | Reproduction - Number of young per adult | 378 | 122.4 | 319 | 6.8 | De Schamphelaere et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | EC10 | Reproduction - Number of young per adult | 59.2 | 124.7 | 49 | 8.4 | De Schamphelaere et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | EC10 | Reproduction - Number of young per adult | 265 | 183.2 | 159 | 8 | De Schamphelaere et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | EC10 | Reproduction - Number of young per adult | 171 | 189.3 | 100 | 8 | De Schamphelaere et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | EC10 | Reproduction - Number of young per adult | 126 | 196.4 | 71 | 8.2 | De Schamphelaere et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | EC10 | Reproduction - Number of young per adult | 196 | 250.5 | 90 | 7.2 | De Schamphelaere et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | EC50 | Reproduction - Number of young per adult | 112 | 26.5 | 344 | 7.3 | De Schamphelaere et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | EC50 | Reproduction - Number of young per adult | 536 | 122.4 | 452 | 6.8 | De Schamphelaere et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | EC50 | Reproduction - Number of young per adult | 171 | 124.7 | 142 | 8.4 | De Schamphelaere et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | EC50 | Reproduction - Number of young per adult | 473 | 183 | 284 | 8 | De Schamphelaere et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | EC50 | Reproduction - Number of young per adult | 313 | 189 | 183 | 8 | De Schamphelaere et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | EC50 | Reproduction - Number of young per adult | 242 | 196.4 | 137 | 8.2 | De Schamphelaere et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | EC50 | Reproduction - Number of young per adult | 299 | 250.5 | 137 | 7.2 | De Schamphelaere et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | NOEC | Reproduction - Number of young per adult | 62.6 | 13.8 | 334 | 6 | De Schamphelaere et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | NOEC | Reproduction - Number of young per adult | 94.5 | 26.5 | 291 | 7.3 | De Schamphelaere et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | NOEC | Reproduction - Number of young per adult | 491 | 122.4 | 414 | 6.8 | De Schamphelaere et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | NOEC | Reproduction - Number of young per adult | 72.7 | 124.7 | 60 | 8.4 | De Schamphelaere et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | NOEC | Reproduction - Number of young per adult | 251 | 183 | 151 | 8 | De Schamphelaere et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | NOEC | Reproduction - Number of young per adult | 244 | 189.3 | 142 | 8 | De Schamphelaere et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | NOEC | Reproduction - Number of young per adult | 143 | 196.4 | 81 | 8.2 | De Schamphelaere et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | NOEC | Reproduction - Number of young per adult | 155 | 250.5 | 71 | 7.2 | De Schamphelaere et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Population | 21 d | EC10 | Population growth inhibition | 420 | 225 | 211 | 8.1 | Enserink et al. | 1991 |
| Water flea | <i>Daphnia magna</i> | Population | 21 d | EC50 | Population growth inhibition | 570 | 225 | 287 | 8.1 | Enserink et al. | 1991 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | LC50 | Mortality | 840 | 225 | 423 | 8.1 | Enserink et al. | 1991 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | LOEC | Length | 120 | 225 | 60 | 8.1 | Enserink et al. | 1991 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | LOEC | Mortality | 1000 | 225 | 504 | 8.1 | Enserink et al. | 1991 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 21 d | LOEC | Repro - Number of young per survivor | 1000 | 225 | 504 | 8.1 | Enserink et al. | 1991 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | EC10 | Reproduction | 328 | 35 | 797 | 7.25 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | EC10 | Reproduction | 233 | 110 | 215 | 8 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | EC10 | Reproduction | 277 | 110 | 256 | 6.5 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | EC10 | Reproduction | 313 | 110 | 289 | 6.5 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | EC10 | Reproduction | 557 | 110 | 514 | 8 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | EC10 | Reproduction | 179 | 240 | 85 | 7.25 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | EC10 | Reproduction | 308 | 240 | 147 | 7.25 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | EC10 | Reproduction | 331 | 240 | 158 | 7.25 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | EC10 | Reproduction | 394 | 240 | 188 | 7.25 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | EC10 | Reproduction | 502 | 240 | 239 | 7.25 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | EC10 | Reproduction | 634 | 240 | 302 | 8.5 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | EC10 | Reproduction | 911 | 240 | 434 | 7.25 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | EC10 | Reproduction | 423 | 240 | 202 | 6 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | EC10 | Reproduction | 114 | 370 | 38 | 6.5 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | EC10 | Reproduction | 341 | 370 | 113 | 6.5 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | EC10 | Reproduction | 600 | 370 | 198 | 8 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | EC10 | Reproduction | 90 | 370 | 30 | 8 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | NOEC | Reproduction | 445 | 35 | 1082 | 7.25 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | NOEC | Reproduction | 209 | 110 | 193 | 8 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | NOEC | Reproduction | 320 | 110 | 295 | 6.5 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | NOEC | Reproduction | 445 | 110 | 411 | 6.5 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | NOEC | Reproduction | 630 | 110 | 581 | 8 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | NOEC | Reproduction | 1000 | 240 | 477 | 7.25 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | NOEC | Reproduction | 209 | 240 | 100 | 7.25 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | NOEC | Reproduction | 320 | 240 | 153 | 7.25 | Heijerick et al. | 2003 |

Table C.1: Summary of long-term effects of zinc on a variety of aquatic organisms

| Species Common Name | Scientific Name | Life Stage | Duration | Endpoint | Observed Effect | Effect Concentration (ug/L) | Test Hardness (mg/L) | Effect Concentration (ug/L) at 100 mg/L Hardness ¹ | pH | Authors | Year |
|---------------------|----------------------|-------------------|----------|----------|------------------------------------------|-----------------------------|----------------------|---------------------------------------------------------------|---------|-------------------------|------|
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | NOEC | Reproduction | 575 | 240 | 274 | 7.25 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | NOEC | Reproduction | 575 | 240 | 274 | 7.25 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | NOEC | Reproduction | 630 | 240 | 300 | 8.5 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | NOEC | Reproduction | 425 | 240 | 203 | 6 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | NOEC | Reproduction | 320 | 370 | 106 | 8 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | NOEC | Reproduction | 320 | 370 | 106 | 6.5 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | NOEC | Reproduction | 630 | 370 | 208 | 6.5 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Not reported | 21 d | NOEC | Reproduction | 630 | 370 | 208 | 8 | Heijerick et al. | 2003 |
| Water flea | <i>Daphnia magna</i> | Adult | 21 d | EC50 | Reproduction | 196 | 50 | 352 | 7 | Heijerick et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Adult | 21 d | EC50 | Reproduction | 202 | 50 | 363 | 6.5 | Heijerick et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Adult | 21 d | EC50 | Reproduction | 218 | 50 | 392 | 6 | Heijerick et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Adult | 21 d | EC50 | Reproduction | 233 | 50 | 419 | 7.5 | Heijerick et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Adult | 21 d | EC50 | Reproduction | 239 | 50 | 430 | 5.5 | Heijerick et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Adult | 21 d | EC50 | Reproduction | 262 | 50 | 471 | 8 | Heijerick et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Adult | 21 d | NOEC | Reproduction | 117 | 50 | 210 | 8 | Heijerick et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Adult | 21 d | NOEC | Reproduction | 133 | 50 | 239 | 7.5 | Heijerick et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Adult | 21 d | NOEC | Reproduction | 154 | 50 | 277 | 7 | Heijerick et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Adult | 21 d | NOEC | Reproduction | 161 | 50 | 289 | 5.5 | Heijerick et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Adult | 21 d | NOEC | Reproduction | 162 | 50 | 291 | 6.5 | Heijerick et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Adult | 21 d | NOEC | Reproduction | 168 | 50 | 302 | 6 | Heijerick et al. | 2005 |
| Water flea | <i>Daphnia magna</i> | Less than 48h old | 21 d | IC10 | Reproduction - Number of young per adult | 67.6 | 64.9 | 97 | 7.6-7.8 | Münzinger and Monicelli | 1991 |
| Water flea | <i>Daphnia magna</i> | Less than 48h old | 21 d | LOEC | Mortality | 150 | 64.9 | 216 | 7.6-7.8 | Münzinger and Monicelli | 1991 |
| Water flea | <i>Daphnia magna</i> | Less than 48h old | 21 d | LOEC | Reproduction - Number of young per adult | 150 | 64.9 | 216 | 7.6-7.8 | Münzinger and Monicelli | 1991 |
| Water flea | <i>Daphnia magna</i> | Less than 48h old | 21 d | MATC | Mortality | 122 | 64.9 | 176 | 7.6-7.8 | Münzinger and Monicelli | 1991 |
| Water flea | <i>Daphnia magna</i> | Less than 48h old | 21 d | MATC | Reproduction - Number of young per adult | 122 | 64.9 | 176 | 7.6-7.8 | Münzinger and Monicelli | 1991 |
| Water flea | <i>Daphnia magna</i> | Less than 48h old | 21 d | NOEC | Mortality | 100 | 64.9 | 144 | 7.6-7.8 | Münzinger and Monicelli | 1991 |
| Water flea | <i>Daphnia magna</i> | Less than 48h old | 21 d | NOEC | Reproduction - Number of young per adult | 100 | 64.9 | 144 | 7.6-7.8 | Münzinger and Monicelli | 1991 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | IC10 | Reproduction - Brood size | 29.8 | 51.9 | 52 | 8.39 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | IC10 | Reproduction - Brood size | 32.8 | 51.9 | 57 | 8.39 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | IC10 | Reproduction - Brood size | 55.7 | 51.9 | 97 | 8.39 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | IC10 | Reproduction - Brood size | 65.8 | 101.8 | 65 | 8.32 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | IC10 | Reproduction - Brood size | 158 | 197 | 89 | 8.29 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | IC10 | Reproduction - Brood size | 214 | 197 | 121 | 8.29 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | MATC | Mortality | 111.8 | 51.9 | 195 | 8.39 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | MATC | Mortality | 120.2 | 51.9 | 209 | 8.39 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | MATC | Mortality | 86.6 | 51.9 | 151 | 8.39 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | MATC | Mortality | 124.4 | 101.8 | 123 | 8.32 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | MATC | Mortality | 178.7 | 197 | 101 | 8.29 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | MATC | Mortality | 237.2 | 197 | 134 | 8.29 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | MATC | Reproduction - Brood size | 21.7 | 51.9 | 38 | 8.39 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | MATC | Reproduction - Brood size | 99.2 | 51.9 | 173 | 8.39 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | MATC | Reproduction - Brood size | 86.6 | 101.8 | 85 | 8.32 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | MATC | Reproduction - Brood size | 174.6 | 197 | 98 | 8.29 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | MATC | Reproduction - Brood size | 224.7 | 197 | 127 | 8.29 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | NEC | Mortality | 112.5 | 51.9 | 196 | 8.39 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | NEC | Mortality | 120.8 | 51.9 | 210 | 8.39 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | NEC | Mortality | 87.5 | 51.9 | 152 | 8.39 | Paulauskis and Winner | 1988 |

Table C.1: Summary of long-term effects of zinc on a variety of aquatic organisms

| Species Common Name | Scientific Name | Life Stage | Duration | Endpoint | Observed Effect | Effect Concentration (ug/L) | Test Hardness (mg/L) | Effect Concentration (ug/L) at 100 mg/L Hardness ¹ | pH | Authors | Year |
|---------------------|-----------------------------|-----------------|----------|----------|---------------------------|-----------------------------|----------------------|---------------------------------------------------------------|---------|-----------------------|------|
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | NEC | Mortality | 125 | 101.8 | 123 | 8.32 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | NEC | Mortality | 179.2 | 197 | 101 | 8.29 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | NEC | Mortality | 237.5 | 197 | 134 | 8.29 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | NEC | Reproduction - Brood size | 100 | 51.9 | 174 | 8.39 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | NEC | Reproduction - Brood size | 25 | 51.9 | 44 | 8.39 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | NEC | Reproduction - Brood size | 87.5 | 101.8 | 86 | 8.32 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | NEC | Reproduction - Brood size | 175 | 197 | 99 | 8.29 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 50 d | NEC | Reproduction - Brood size | 225 | 197 | 127 | 8.29 | Paulauskis and Winner | 1988 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 134 d | LOEC | Growth | 100 | 145 | 73 | 8.2-9.5 | Winner | 1981 |
| Water flea | <i>Daphnia magna</i> | Less than 24hrs | 134 d | LOEC | Reproduction - Brood size | 300 | 145 | 219 | 8.2-9.5 | Winner | 1981 |
| zebra mussel | <i>Dreissena polymorpha</i> | Adult | 10 weeks | EC50 | Filtration rate | 131 | 268 | 57 | 7.9 | Kraak et al. | 1994 |
| zebra mussel | <i>Dreissena polymorpha</i> | Adult | 10 weeks | LC10 | Mortality | 517 | 268 | 225 | 7.9 | Kraak et al. | 1994 |
| zebra mussel | <i>Dreissena polymorpha</i> | Adult | 10 weeks | LC50 | Mortality | 1065 | 268 | 463 | 7.9 | Kraak et al. | 1994 |
| zebra mussel | <i>Dreissena polymorpha</i> | Adult | 10 weeks | LOEC | Filtration rate | 382 | 268 | 166 | 7.9 | Kraak et al. | 1994 |
| zebra mussel | <i>Dreissena polymorpha</i> | Adult | 10 weeks | MATC | Filtration rate | 196 | 268 | 85 | 7.9 | Kraak et al. | 1994 |
| zebra mussel | <i>Dreissena polymorpha</i> | Adult | 10 weeks | NOEC | Filtration rate | 101 | 268 | 44 | 7.9 | Kraak et al. | 1994 |

¹ If adjusted from another hardness, value was calculated using the following equation: EXP(LN(EFFECT conc)- (0.846)*(LN(measured water hardness)-LN(desired water hardness)))

most sensitive effect end-point

APPENDIX D

Hardness – Toxicity Relationships For Zinc and Cadmium

Table D.1: Data used to develop hardness-zinc toxicity relationships for long-term exposures to zinc.

| Family | Species Common Name | Scientific Name | Duration | Endpoint | Life Stage | Observed Effect | Test Hardness (mg/L) | Effect Concentration (µg/L) | Mean Effect Concentration if Applicable (ug/L) | pH | Author(s) | Year |
|---------------|---------------------|----------------------------------------|----------|-----------|--------------|---------------------------------------------|----------------------|-----------------------------|------------------------------------------------|-----------|-----------------------------|------|
| Algae | Green Algae | <i>Pseudokirchneriella subcapitata</i> | 72 h | EC50 | Population | biomass | 212.4 | 17.5 | ← | 7.5 | Heijerick et al. | 2002 |
| | | | 72 h | EC50 | Population | biomass | 212.4 | 34.4 | ← | 7.5 | Heijerick et al. | 2002 |
| | | | | Mean EC50 | | | | | 24.5 | | | |
| | | | 72 h | EC50 | Population | biomass | 162.3 | 15.4 | ← | 7.5 | Heijerick et al. | 2002 |
| | | | 72 h | EC50 | Population | biomass | 162.3 | 33.2 | ← | 7.5 | Heijerick et al. | 2002 |
| | | | | Mean EC50 | | | | | 22.6 | | | |
| | | | 72 h | EC50 | Population | biomass | 112.3 | 14.9 | ← | 7.5 | Heijerick et al. | 2002 |
| | | | 72 h | EC50 | Population | biomass | 112.3 | 27.9 | ← | 7.5 | Heijerick et al. | 2002 |
| | | | | Mean EC50 | | | | | 20.4 | | | |
| | | | 72 h | EC50 | Population | biomass | 62.3 | 10.9 | ← | 7.5 | Heijerick et al. | 2002 |
| | | | 72 h | EC50 | Population | biomass | 62.3 | 11.1 | ← | 7.5 | Heijerick et al. | 2002 |
| | | | | Mean EC50 | | | | | 11.0 | | | |
| | | | 72 h | EC50 | Population | biomass | 37.2 | 8.44 | ← | 7.5 | Heijerick et al. | 2002 |
| | | | 72 h | EC50 | Population | biomass | 37.2 | 10.5 | ← | 7.5 | Heijerick et al. | 2002 |
| | | | | Mean EC50 | | | | | 9.41 | | | |
| | | | 72 h | EC50 | Population | biomass | 262.6 | 54.9 | | 7.5 | Heijerick et al. | 2002 |
| Invertebrates | Water Flea | <i>Daphnia magna</i> | 21 d | EC10 | < 24 hrs | Reproduction - # young / adult | 196.4 | 126 | | 8.2 | De Schampelaere et al. | 2005 |
| | | | 21 d | EC10 | < 24 hrs | Reproduction - # young / adult | 189.3 | 171 | | 8 | De Schampelaere et al. | 2005 |
| | | | 21 d | EC10 | < 24 hrs | Reproduction - # young / adult | 183.2 | 265 | | 8 | De Schampelaere et al. | 2005 |
| | | | 21 d | EC10 | < 24 hrs | Reproduction - # young / adult | 124.7 | 59.2 | | 8.4 | De Schampelaere et al. | 2005 |
| | | | 21 d | EC10 | Not reported | Reproduction (1) | 110 | 233 | ← | 8 | Heijerick et al. | 2003 |
| | | | 21 d | EC10 | Not reported | Reproduction (1) | 110 | 557 | ← | 8 | Heijerick et al. | 2003 |
| | | | | Mean EC10 | | | | | 360 | | | |
| | | | 21 d | EC10 | Not reported | Reproduction (1) | 240 | 634 | | 8.5 | Heijerick et al. | 2003 |
| | | | 21 d | EC10 | Not reported | Reproduction (1) | 370 | 600 | ← | 8 | Heijerick et al. | 2003 |
| | | | 21 d | EC10 | Not reported | Reproduction (1) | 370 | 90 | ← | 8 | Heijerick et al. | 2003 |
| | | | | Mean EC10 | | | | | 232 | | | |
| | | | 50 d | IC10 | < 24 hrs | Reproduction - Brood size | 51.9 | 29.8 | ← | 8.39 | Paulauskis and Winner | 1988 |
| | | | 50 d | IC10 | < 24 hrs | Reproduction - Brood size | 51.9 | 32.8 | ← | 8.39 | Paulauskis and Winner | 1988 |
| | | | 50 d | IC10 | < 24 hrs | Reproduction - Brood size | 51.9 | 55.7 | ← | 8.39 | Paulauskis and Winner | 1988 |
| | | | | Mean IC10 | | | | | 37.9 | | | |
| | | | 50 d | IC10 | < 24 hrs | Reproduction - Brood size | 101.8 | 65.8 | | 8.32 | Paulauskis and Winner | 1988 |
| | | | 50 d | IC10 | < 24 hrs | Reproduction - Brood size | 197 | 158 | ← | 8.29 | Paulauskis and Winner | 1988 |
| | | | 50 d | IC10 | < 24 hrs | Reproduction - Brood size | 197 | 214 | ← | 8.29 | Paulauskis and Winner | 1988 |
| | | | | Mean IC10 | | | | | 184 | | | |
| | | <i>Ceriodaphnia dubia</i> | 21 d | EC10 | Population | Population growth inhibition | 225 | 420 | | 8.1 | Enserink et al. | 1991 |
| | | | 21 d | EC16 | < 24 hrs | Reproduction - Number of young per survivor | 49 | 70 | | 7.4-8.2 | Biesinger and Christensen | 1972 |
| | | | | IC10 | < 48 hrs | Reproduction - Number of young per adult | 64.9 | 67.6 | | 7.6-7.8 | Münzinger and Monicelli | 1991 |
| | | | 7 d | ChV | 24hrs | Reproduction - Brood size | 169 | 105 | | 7.8-8.2 | Masters et al. | 1991 |
| | | | 4 weeks | MATC | < 24 hrs | Reproduction - Number of young per adult | 97.6 | 35 | ← | 9 | Belanger et Cherry | 1990 |
| | | | 4 weeks | MATC | < 24 hrs | Reproduction - Number of young per adult | 113.6 | 71 | ← | 6, 8, 9 | Belanger et Cherry | 1990 |
| | | | 4 weeks | Mean MATC | 24hrs | Reproduction- Number of young per adult | 106 | | 50 | | | |
| | | | 4 weeks | MATC | 24hrs | Reproduction - Number of young per adult | 182 | 71 | | 8 | Belanger et Cherry | 1990 |
| | Amphipod | <i>Hyalella azteca</i> | 7 d | LC50 | 1- 11 days | Mortality (1) | 18 | 56 | ← | 6.44-8.68 | Borgmann et al | 2005 |
| | | | 7 d | LC50 | 1- 11 days | Mortality (1) | 18 | 70 | ← | 6.44-8.68 | Borgmann et al | 2005 |
| | | | 7 d | Mean LC50 | 1- 11 days | Mortality (1) | | | 63 | | | |
| | | | 7 d | LC50 | 1- 11 days | Mortality (1) | 124 | 222 | | 7.23-8.98 | Borgmann et al | 2005 |
| Fish | Rainbow Trout | <i>Oncorhynchus mykiss</i> | 30 d | LC10 | Juvenile | Mortality (1) | 31.72 | 34.5 | | 7.58-7.87 | De Schampelaere and Janssen | 2004 |
| | | | 30 d | LC10 | Juvenile | Mortality (1) | 104.99 | 171 | | 7.58-7.87 | De Schampelaere and Janssen | 2004 |
| | | | 30 d | LC10 | Juvenile | Mortality (1) | 190.35 | 290 | | 7.58-7.87 | De Schampelaere and Janssen | 2004 |
| | | | 30 d | LC10 | Juvenile | Mortality (1) | 398.68 | 337 | | 7.58-7.87 | De Schampelaere and Janssen | 2004 |
| | | | 30 d | LC10 | Juvenile | Mortality (1) | 29.6 | 38.4 | ← | 7.45 | De Schampelaere and Janssen | 2004 |
| | | | 30 d | LC10 | Juvenile | Mortality (1) | 29.1 | 46.1 | ← | 7.65 | De Schampelaere and Janssen | 2004 |
| | | | 30 d | LC10 | Juvenile | Mortality (1) | 29.1 | 73.6 | ← | 7.58 | De Schampelaere and Janssen | 2004 |
| | | | | Mean LC10 | | | 29.3 | | 50.7 | | | |
| | | | 30 d | LC10 | Juvenile | Mortality (1) | 29.1 | 99.1 | ← | 6.78 | De Schampelaere and Janssen | 2004 |
| | | | 30 d | LC10 | Juvenile | Mortality (1) | 28.2 | 185 | ← | 6.8 | De Schampelaere et al. | 2005 |
| | | | | Mean LC10 | | | | | 135 | | | |
| | | | 30 d | LC10 | Juvenile | Mortality (1) | 31.5 | 219 | | 7.08 | De Schampelaere et al. | 2005 |
| | | | 30 d | LC10 | Juvenile | Mortality (1) | 176.3 | 578 | | 8.13 | De Schampelaere et al. | 2005 |
| | | | 30 d | LC10 | Juvenile | Mortality (1) | 103.7 | 902 | | 7.76 | De Schampelaere et al. | 2005 |
| | Fathead Minnow | <i>Pimephales promelas</i> | 32 d | ChV | Larva | Mortality | 47 | 188 | | 7.4-8.2 | Norberg-King | 1989 |
| | | | 7 d | LC50 | Larva | Mortality | 190 | 780 | | 8.3-8.7 | Magliette et al. | 1995 |
| | | | NR | MATC | Egg | Mortality | 46 | 207 | | 7-8 | Benoit and Holcombe | 1978 |
| | | | 7 d | MATC | Larva | Mortality | 48 | 125 | | | Norberg and Mount | 1985 |
| | | | 7 d | TLM | Fry | Mortality | 186 | 870 | | 7.5-7.6 | Pickering and Vigor | 1965 |

"← " values included in mean

Table D.2: Data used to develop hardness-cadmium relationships for long-term exposure to cadmium.

| Family | Species Common Name | Scientific Name | Duration | Endpoint | Life stage | Observed Effect | Average Hardness | Effect Concentration (ug/L) | Mean Effect Concentration if Applicable (ug/L) | pH | Authors | Year |
|---------------|---------------------|----------------------------------------|----------|-----------|-----------------|------------------------------------------|------------------|-----------------------------|------------------------------------------------|----------------|-------------------------------------|------|
| Algae | Green Algae | <i>Pseudokirchneriella subcapitata</i> | 72 h | EC10 | Population | Growth rate | 3.42 | 2.8 | ← | 6.71 | Kallqvist | 2007 |
| | | | 72 h | EC10 | Population | Growth rate | 6.21 | 7.5 | ← | 6.85 | Kallqvist | 2007 |
| | | | 72 h | EC10 | Population | Growth rate | 16.21 | 8.5 | ← | 6.74 | Kallqvist | 2007 |
| | | | 72 h | EC10 | Population | Growth rate | 46.21 | 6 | ← | 6.65 | Kallqvist | 2007 |
| | | | | | | | 18.01 | | 5.7 | | | |
| Invertebrates | Water Flea | <i>Daphnia magna</i> | 72 h | EC50 | Population | Growth | 250 | 43.5 | | 8.1 | Benhra et al. | 1997 |
| | | | 21 d | EC16 | Less than 24hrs | Reproduction | 45.3 | 0.17 | | 7.74 (7.4-8.2) | Biesinger and Christensen | 1972 |
| | | | 21 d | MATC | Less than 24hrs | Reproduction - Number of young per adult | 53 | 0.15 | | 7.5 +- 0.2 | Chapman et al | 1980 |
| | | | 7 d | MATC | Neonate | Growth | 90 | 1.2 | | | Winner | 1988 |
| | | | 21 d | MATC | Not reported | Reproduction - Number of young per adult | 130 | 0.64 | | | Borgmann et al | 1989 |
| | | | 7 d | EC10 | Adult | Repro - brood mass | 179 | 0.13 | | 8.07 +- 0.07 | Barata and Baird | 2000 |
| | | | 21 d | MATC | 24h | Reproduction | 249.8 | 1.09 | | 8.0 +- 0.2 | Kuhn et al | 1989 |
| | | | 14 d | MATC | Less than 24hrs | Reproduction - Number of young per adult | 240 | 4.3 | | 8.0 +- 0.3 | Elnabarawy et al | 1986 |
| Fish | Brown Trout | <i>Salmo trutta</i> | 30 d | IC20 | Swim-up fry | Biomass, decrease in | 29.2 | 0.87 | | 7.54 (0.13) | Brinkman and Hansen | 2007 |
| | | | 61 d | MATC | Larva | Biomass, decrease in | 45 | 2 | ← | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| | | | 60 d | MATC | Larva | Biomass, decrease in | 45 | 6.7 | ← | 7.6 (7.2-7.8) | Eaton et al. | 1978 |
| | | | | mean MATC | | | 45 | | 3.7 | | | |
| | | | 30 d | IC20 | Swim-up fry | Biomass, decrease in | 67.6 | 2.18 | | 7.60 (0.10) | Brinkman and Hansen | 2007 |
| | | | 30 d | IC20 | Swim-up fry | Biomass, decrease in | 151.0 | 6.62 | | 7.51 (0.12) | Brinkman and Hansen | 2007 |
| | | | 50 d | ChV | Embryo-larval | | 250 | 16.49 | | | Brown et al. (as cited by EPA 2001) | 1994 |

"←" values included in mean

Table D.3: Hardness-Toxicity Relationships for Zinc and Cadmium

| Parameter | Species | n | Slope | <i>p</i> -value | <i>r</i> ² |
|-----------|----------------------------------------|----|-------|-----------------|-----------------------|
| Zinc | <i>Pseudokirchneriella subcapitata</i> | 6 | 0.785 | 0.007 | 0.869 |
| | <i>Daphnia magna</i> | 13 | 1.027 | 0.004 | 0.541 |
| | <i>Ceriodaphnia dubia</i> | 3 | 0.983 | 0.432 | 0.606 |
| | <i>Hyalella azteca</i> | 2 | 0.656 | - | 1.000 |
| | <i>Oncorhynchus mykiss</i> | 9 | 0.719 | 0.046 | 0.455 |
| | <i>Pimephales promelas</i> | 5 | 1.135 | 0.006 | 0.944 |
| | Combined Slope (<i>p</i> = 0.909) | 38 | 0.846 | 0.000 | 0.804 |
| Cadmium | <i>Pseudokirchneriella subcapitata</i> | 2 | 0.771 | - | 1.000 |
| | <i>Daphnia magna</i> | 7 | 1.159 | 0.148 | 0.370 |
| | <i>Salmo trutta</i> | 5 | 1.179 | 0.022 | 0.864 |
| | Combined Slope (<i>p</i> = 0.830) | 14 | 1.023 | 0.830 | 0.810 |

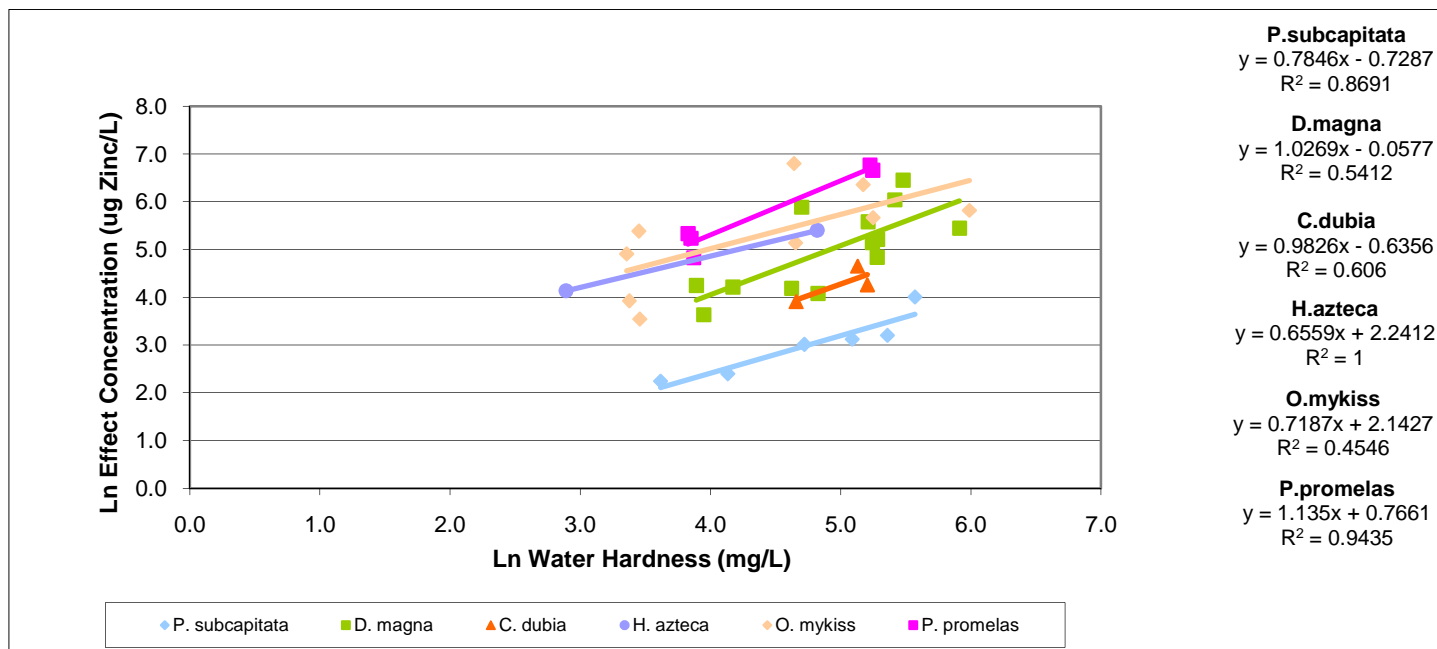


Figure D.1: Comparison of zinc hardness-toxicity relationships for various aquatic species in long-term exposures. Data in Appendix Table D.1.

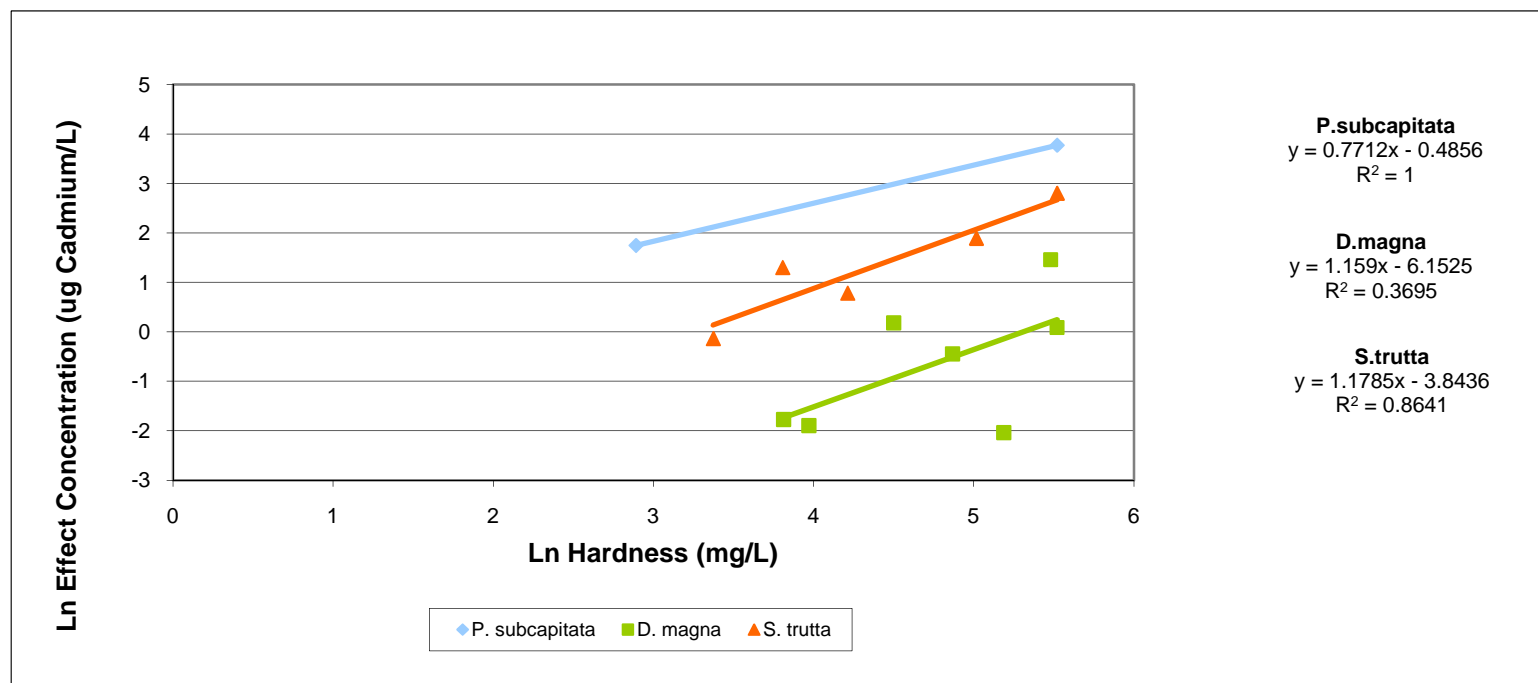


Figure D.2: Comparison of cadmium hardness-toxicity relationships for various aquatic species in long-term exposures (Data in Appendix Table D.2).